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(54) Title: GENES FOR ALTERING MITOCHONDRIAL FUNCTION AND FOR HYBRID SEED PRODUCTION

(57) Abstract: The present invention relates to isolated nucleic acid molecules which restore fertility to cytoplasmic male sterile plants and modify expression of toxic mitochondria proteins by the plant. The present invention also relates to methods of identifying a candidate plant suitable for breeding with a cytoplasmic male sterile plant and methods of identifying a candidate gene restoring fertility in plants by analyzing for the candidate plant and candidate gene, respectively, for the presence of the nucleic acid molecule of the present invention. Also disclosed are methods of producing hybrid plant seed, methods of directing gene expression to plant mitochondria, and method of expressing a gene preferentially in roots of a plant. Promoters and terminators from plant genes which restore fertility to cytoplasmic male sterile plants and modify expression of toxic mitochondria proteins are also disclosed. Finally, methods of producing plants with a cytoplasmic male sterile plant restoration system are disclosed.

## GENES FOR ALTERING MITOCHONDRIAL FUNCTION AND FOR HYBRID SEED PRODUCTION

[0001] This application claims the benefit of U.S. Patent Application Serial No. 60/347,996, filed January 10, 2002, which is hereby incorporated by reference in its entirety.

[0002] This invention arose out of research sponsored by the USDA NRI (Grant No. 98-35300-6171). The U.S. Government may have certain rights in this invention.

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### FIELD OF THE INVENTION

[0003] The present invention relates to improving productivity or usefulness of plants by altering mitochondrial gene expression and to the production of hybrid seed. Specifically, the present invention relates to the use of genes that affect mitochondrial gene expression, some of which ameliorate male sterility and others which cause male sterility or altered floral development. The invention also provides a method of facilitating the identification of genes with similar functions in other plant species.

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### BACKGROUND OF THE INVENTION

[0004] A widely used method for producing hybrid seeds involves crossing a cytoplasmic male sterile (CMS) plant line with a fertile plant line. Typically, the fertile line contains a fertility restorer gene in its nuclear genome, so that all of the progeny are male fertile. All seeds collected from a CMS plant must result from cross-pollination. However, the hybrid seed so generated will itself be male sterile unless the male parent has brought a nuclear fertility-restorer gene into the next generation. The fertility of the progeny is important for productivity in plant varieties where self-pollination is responsible for production of the desirable crop. For example, a fruit crop of a self-pollinated species requires male fertility, while an ornamental species will produce attractive flowers or plant morphology even when no pollen is produced.

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[0005] While a number of naturally occurring CMS/restorer systems exist and are currently in use for hybrid seed production, there are a number of crop species which lack known CMS and fertility restorer genes. For example, a hybrid seed of tomato is typically made by hand emasculation of plants to be used as female parents.

5 This hand-made method of cross-pollination is quite labor intensive and cost-prohibitive for many crops. In addition, certain naturally occurring CMS/restorer systems have some drawbacks. For example, corn plants carrying the CMS-T cytoplasm are more susceptible to a blight disease.

[0006] Fertility restorer genes that have been particularly useful for hybrid seed production are active as single dominant alleles at a locus, though multigenic systems are sometimes used. A *Petunia* fertility restorer locus termed *Rf* is known to be effective with no additional helper genes to restore fertility (Edwardson et al., "Fertility Restoration in Cytoplasmic Male Sterile *Petunia*," *J. Hered.*, 58:195-196 (1967); Izhar, "Cytoplasmic Male Sterility in Petunia. III. Genetic Control on 10 Microsporogenesis and Male Fertility Restoration," *J. Hered.*, 69:22-26 (1978)).

[0007] Nuclear fertility restoration genes confer normal pollen development upon plants carrying sterility-encoding mitochondria. The mitochondrial genes responsible for causing the male sterility have been identified in a number of species, including *Petunia*, maize, *Brassica*, and common bean. The expression of these 15 CMS-encoding mitochondrial genes is affected by the nuclear restorer genes, as shown for *Rf* in *Petunia* (Pruitt et al., "Cytochrome Oxidase Subunit II Sequences in *Petunia* Mitochondria: Two Intron-Containing Genes and an Intron-Less Pseudogene Associated With Cytoplasmic Male Sterility," *Curr. Genet.*, 16:281-91 (1989); Nivison et al., "Identification of a Mitochondrial Protein Associated With 20 Cytoplasmic Male Sterility in *Petunia*," *Plant Cell*, 1:1121-30 (1989); Nivision et al., "Sequencing, Processing, and Localization of the *Petunia* CMS-Associated Mitochondrial Protein," *Plant J.*, 5:613-623 (1994); Hanson et al., "Mitochondrial Gene Organization and Expression in Petunia Male Fertile and Sterile Plants," *J. Hered.*, 90:362-368 (1999)); *RfI* in CMS-T maize (Dewey et al., "Novel 25 Recombinations in the Maize Mitochondrial Genome Produce a Unique Transcriptional Unit in the Texas Male-Sterile Cytoplasm," *Cell*, 44:439-49 (1986); Wise et al., "Mitochondrial Transcript Processing and Restoration of Male Fertility in T-Cytoplasm Maize," *J. Hered.*, 90:380-385 (1999); Kennell et al., "Influence of 30

Nuclear Background on Transcription of a Maize Mitochondrial Region Associated With Texas Male Sterile Cytoplasm," Mol. Gen. Genet., 210:399-406 (1987); Kennell et al., "Initiation and Processing of *atp6*, T-*urf13*, and *ORF221* Transcripts From Mitochondria of T Cytoplasm Maize," Mol. Gen. Genet., 216:16-24 (1989)); *Rfp1* and 5 *rfp1* in *Brassica* (Singh et al., Suppression of Cytoplasmic Male Sterility by Nuclear Genes Alters Expression of a Novel Mitochondrial Gene Region," Plant Cell, 3:1349-1362 (1991); Singh et al., "Nuclear Genes Associated With a Single Brassica CMS Restorer Locus Influence Transcripts of Three Different Mitochondrial Gene Regions," Genetics, 143:505-516 (1996)); restorers in radish (Krishnasamy et al., 10 "Organ-Specific Reduction in the Abundance of a Mitochondrial Protein Accompanies Fertility Restoration in Cytoplasmic Male-Sterile Radish," Plant Molec. Biol., 26:935-946 (1994)); restorers in sunflower (Horn et al., "A Mitochondrial 16 kDa Protein is Associated With Cytoplasmic Male Sterility in Sunflower," Plant Molec. Biol., 17:29-36 (1991); Laver et al., "Mitochondrial Genome Organization and 15 Expression Associated With Cytoplasmic Male Sterility in Sunflower (*Helianthus annuus*)," Plant J., 1:185-193 (1991); Monéger et al., "Nuclear Restoration of Cytoplasmic Male Sterility in Sunflower is Associated With the Tissue-Specific Regulation of a Novel Mitochondrial Gene," EMBO J., 13:8-17 (1994); Smart et al., "Cell-Specific Regulation of Gene Expression in Mitochondria During Anther 20 Development in Sunflower," Plant Cell, 6:811-825 (1994)); restorers in rice (Akagi et al., "A Unique Sequence Located Downstream From the Rice Mitochondrial *atp6* May Cause Male Sterility," Curr. Genet., 25:52-58 (1994); Kadowaki et al., "A Chimeric Gene Containing the 5' Portion of *atp6* is Associated With Cytoplasmic Male Sterility of Rice," Mol. Gen. Genet., 224:10-16 (1990)); and *Fr2* in broad bean 25 (Chase, "Expression of CMS-Unique and Flanking Mitochondrial DNA Sequencs in *Phaseolus vulgaris*," L. Curr. Genet., 25:245-251 (1993); He et al., "Pollen Fertility Restoration by Nuclear Gene *Fr* in CMS Bean: Nuclear-Directed Alteration of a Mitochondrial Population," Genetics, 139:995-962 (1995)). The expression of various nuclear restorer genes has been reported to be either enhanced in reproductive 30 tissue, as in the case of sunflower, or, as in the case of *Petunia*, expressed in both vegetative and reproductive tissues. Thus, different fertility restorer genes carry different promoters and nuclear expression regulatory elements which may confer very limited tissue-specific expression or very broad expression in the plant.

[0008] Reduction in the amount of the protein product of the CMS-encoding gene is the usual effect of these restorers whose target mitochondrial genes are known. These genes may possibly act by affecting the transcription or translation rate, the transcript or protein stability, processing, splicing, etc.

5 Alleles of some restorer genes may up-regulate while others may down-regulate the expression of particular mitochondrial genes. Fertility restorer genes and their alleles or homologous counterparts in other species may thus be extremely valuable in engineering the expression of genes introduced into higher plant mitochondria.

[0009] The cloning and sequencing of the restorer gene *Rf2* in maize has been 10 reported in Cui et al., "The *rf2* Nuclear Restorer Gene of Male-Sterile T-Cytoplasm Maize," Science, 272:1334-1336 (1996) and U.S. Patent No. 5,981,833 to Wise et al. This restorer gene acts in conjunction with a second required gene, *Rf1*, the gene that reduces the amount of the toxic protein, to restore fertility to plants carrying the maize CMS-T cytoplasm (Dewey et al., "Novel Recombinations in the Maize Mitochondrial 15 Genome Produce a Unique Transcriptional Unit in the Texas Male-Sterile Cytoplasm," Cell, 44:439-49 (1986); Dewey et al., "A Mitochondrial Protein Associated With Cytoplasmic Male Sterility in the T Cytoplasm of Maize," Proc. Natl. Acad. Sci. USA, 84:5374-78 (1987); Wise et al., "Urf13-T of T Cytoplasm

20 Maize Mitochondria Encodes a 13kD Polypeptide," Plant Mol. Biol., 9:121-26 (1987)). Plants of genotype *Rf1rf2*, though sterile, have greatly reduced amounts of the URF13 protein. In contrast, sterile plants of genotype *rf1Rf2* have abundant amounts of the URF13 protein. The *Rf2* gene is, thus, unusual in that no effect on the expression of the maize T-CMS-associated protein, URF13, has been detected. The sequence of the gene bore out the absence of observable effect on mitochondrial gene 25 expression; according to sequence analysis, *Rf2* is apparently an aldehyde dehydrogenase (Liu et al., "Mitochondrial Aldehyde Dehydrogenase Activity is Required for Male Fertility in Maize," The Plant Cell, 13:1063-1078 (2001)). It has been proposed that *Rf2* acts by compensating for a metabolic defect caused by the low levels of the URF13 protein that remain despite the presence of *Rf1*, the gene that

30 reduces the amount of the toxic protein (Dewey et al., "A Mitochondrial Protein Associated With Cytoplasmic Male Sterility in the T Cytoplasm of Maize," Proc. Natl. Acad. Sci. USA, 84:5374-78 (1987)) and also alters the T-*urf13* transcript profile (Kennell et al., "Influence of Nuclear Background on Transcription of a Maize

Mitochondrial Region Associated With Texas Male Sterile Cytoplasm," Mol. Gen. Genet., 210:399-406 (1987)).

[0010] An abnormal recombinant mitochondrial gene in *Petunia* CMS lines (termed *pcf*) has been genetically correlated with CMS (Young et al., "A Fused 5 Mitochondrial Gene Associated With Cytoplasmic Male Sterility is Developmentally Regulated," Cell, 50:41-49 (1987)). Because plant mitochondrial RNA is edited from C to U in some locations, the edited RNA sequence for the *pcf* gene has been determined, allowing the prediction of the *pcf*-encoded protein (Wintz et al., "A Termination Codon is Created by RNA Editing in the *Petunia* Mitochondrial *atp9* 10 Gene Transcript," Curr. Genet., 19:61-64 (1990); Sutton et al., "Editing of Pre-mRNAs Can Occur Before *cis*- and *trans*-Splicing in *Petunia* Mitochondria," Mol. Cell Biol., 11:4274-4277 (1991); Nivision et al., "Sequencing, Processing, and Localization of the *Petunia* CMS-Associated Mitochondrial Protein," Plant J., 5:613-623 (1994); Hanson et al., "Mitochondrial Gene Organization and Expression in 15 Petunia Male Fertile and Sterile Plants," J. Hered., 90:362-368 (1999)). Antibodies to synthetic peptide sequences have revealed the presence of a 19.5 kD PCF protein located in both the membrane and soluble fraction of mitochondria (Nivison et al., "Identification of a Mitochondrial Protein Associated With Cytoplasmic Male Sterility in *Petunia*," Plant Cell, 1:1121-30 (1989)). The PCF protein is processed 20 from a longer precursor protein and is entirely encoded by the *urfS* region of the *pcf* gene (Nivision et al., "Sequencing, Processing, and Localization of the *Petunia* CMS-Associated Mitochondrial Protein," Plant J., 5:613-623 (1994)). The PCF protein is strongly expressed in sporogenous cells of premeiotic petunia anthers in CMS lines, but undetectable in CMS-*Rf* lines (Conley et al., "Tissue-Specific Protein Expression 25 in Plant Mitochondria," Plant Cell, 6:85-91 (1994)). Abnormalities in *Petunia* pollen development are first observed in meiosis, and by the developmental stage where fertile plants are releasing pollen, CMS anthers are hollow shells (Conley et al., "Effects of *Petunia* Cytoplasmic Male Sterile (CMS) Cytoplasm on the Development of Sterile and Fertility-Restored *P. parodii* Anthers," Am. J. Bot., 81:630-640 (1994)). 30 It is evident that the *pcf* gene product is disrupting mitochondrial function, leading to death of the sporogenous cells, though the exact mechanism at the molecular level is not known.

[0011] In maize T, *Petunia*, rice, and *Brassica* Pol CMS systems, particular transcripts of CMS-associated genes have been reported to be altered in restored lines (Pruitt et al., "Transcription of the *Petunia* Mitochondrial CMS-Associated *pcf* Locus in Male Sterile and Fertility-Restored Lines," *Mol. Gen. Genet.*, 227:348-355 (1991); 5 Dewey et al., "Novel Recombinations in the Maize Mitochondrial Genome Produce a Unique Transcriptional Unit in the Texas Male-Sterile Cytoplasm," *Cell*, 44:439-49 (1986); Kennell et al., "Initiation and Processing of *atp6*, T-*urf13*, and *ORF221* Transcripts From Mitochondria of T Cytoplasm Maize," *Mol. Gen. Genet.*, 216:16-24 (1989); Kennell et al., "Influence of Nuclear Background on Transcription of a Maize 10 Mitochondrial Region Associated With Texas Male Sterile Cytoplasm," *Mol. Gen. Genet.*, 210:399-406 (1987); Singh et al., "Suppression of Cytoplasmic Male Sterility by Nuclear Genes Alters Expression of a Novel Mitochondrial Gene Region," *Plant Cell*, 3:1349-1362 (1991); Singh et al., "Nuclear Genes Associated With a Single Brassica CMS Restorer Locus Influence Transcripts of Three Different Mitochondrial 15 Gene Regions," *Genetics*, 143:505-516 (1996); Wise et al., "Mitochondrial Transcript Processing and Restoration of Male Fertility in T-Cytoplasm Maize," *J. Hered.*, 90:380-385 (1999)). In *Brassica*, the presence of either one of two restorer genes results in monocistronic transcripts of *atp6*, instead of the dicistronic *orf224/atp6* transcripts found in CMS lines (Singh et al., "Suppression of Cytoplasmic Male 20 Sterility by Nuclear Genes Alters Expression of a Novel Mitochondrial Gene Region," *Plant Cell*, 3:1349-1362 (1991)). A UG-rich sequence appears to be the target of the *Brassica* restorer alleles (Singh et al., "Nuclear Genes Associated With a Single *Brassica* CMS Restorer Locus Influence Transcripts of Three Different Mitochondrial Gene Regions," *Genetics*, 143:505-516 (1996)). In *Petunia*, *pcf* transcripts with 5' termini at -121 are specifically reduced in restored lines (Pruitt et al., "Transcription of the *Petunia* Mitochondrial CMS-Associated *pcf* Locus in Male Sterile and Fertility-Restored Lines," *Mol. Gen. Genet.*, 227:348-355 (1991)), while 25 transcripts terminating at -266 and -522 remain at normal levels. In maize T cytoplasm, a sequence unlike either the *Brassica* restorer target or the *Petunia* -121 transcript terminus is the putative recognition signal for the *Rf1* gene (Dill et al., "*Rf8* and *Rf\** Mediate Unique T-*urf13*-Transcript Accumulation, Revealing a Conserved 30 Motif Associated With RNA Processing and Restoration of Pollen Fertility in T-cytoplasm Maize," *Genetics*, 147:1367-1379 (1997)).

[0012] The steady-state amounts of the *Petunia pcf*-encoded protein and the maize *urf13*-encoded protein decrease greatly in restored lines compared to unrestored lines (Nivison et al., "Identification of a Mitochondrial Protein Associated With Cytoplasmic Male Sterility in *Petunia*," Plant Cell, 1:1121-30 (1989); Dewey et al., 5 "Novel Recombinations in the Maize Mitochondrial Genome Produce a Unique Transcriptional Unit in the Texas Male-Sterile Cytoplasm," Cell, 44:439-49 (1986); Wise et al., "Urf13-T of T Cytoplasm Maize Mitochondria Encodes a 13kD Polypeptide," Plant Mol. Biol., 9:121-26 (1987)). Abundance of CMS-associated proteins is also reduced in sunflower and radish (Horn et al., "A Mitochondrial 16 10 kDa Protein is Associated With Cytoplasmic Male Sterility in Sunflower," Plant Mol. Biol., 17:29-36 (1991); Laver et al., "Mitochondrial Genome Organization and Expression Associated With Cytoplasmic Male Sterility in Sunflower (*Helianthus annuus*)," Plant J., 1:185-193 (1991); Krishnasamy et al., "Organ-Specific Reduction in the Abundance of a Mitochondrial Protein Accompanies Fertility Restoration in 15 Cytoplasmic Male-Sterile Radish," Plant Mol. Biol., 26:935-946 (1994)). The mechanism behind the reduction in quantity of CMS-associated proteins in restored lines is not understood. For example, absence of transcripts that could potentially encode the PCF protein is not the explanation; only the shortest transcript is reduced in restored lines (Pruitt et al., "Transcription of the *Petunia* Mitochondrial CMS- 20 Associated *pcf* Locus in Male Sterile and Fertility-Restored Lines," Mol. Gen. Genet., 227:348-355 (1991)).

[0013] In *Petunia* and in some other CMS/restorer systems, the abnormal gene is co-transcribed with known mitochondrial genes. One possible mechanism for CMS in *Petunia* and its restoration, which is also consistent with current data, is that the 25 restorer gene not only results in decrease in the expression of PCF, but also improves the expression of the co-transcribed genes *nad3* and *rps12* in some way. For example, it remains possible that an RNA processing event results in little translation of PCF but enhanced production of NAD3 and RPS12 protein.

[0014] In sum, with the exception of maize *Rf2*, in those systems where 30 analysis has reached the molecular level, restorer genes have been found to affect the abundance of mitochondrial-encoded DNAs, RNAs, and proteins.

[0015] Cytoplasmic male sterility/restorer systems have been proven to be an invaluable tool in the production of hybrid seeds. Despite their importance for both

the production of major crops such as rice and sunflower and the study of organelle/nuclear interactions in plants, none of the nuclear fertility-restorer genes that reduce the expression of aberrant mitochondrial proteins have been cloned.

5 [0016] The present invention is directed to overcoming these deficiencies in the art.

## SUMMARY OF THE INVENTION

[0017] The present invention relates to an isolated nucleic acid molecule which restores fertility to cytoplasmic male sterile plants and modifies expression of 10 toxic mitochondria proteins by the plant. The nucleic acid molecule encodes a protein having an amino acid sequence of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, or 41. Alternatively, the nucleic acid molecule encodes a protein containing a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino 15 acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input. Alternatively, the nucleic acid molecule hybridizes to a nucleotide sequence of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ 20 ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, or SEQ ID NO: 40 under stringent conditions of a hybridization buffer containing 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C. Alternatively, the nucleic acid molecule has a nucleotide 25 sequence of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, or SEQ ID NO: 40.

[0018] Another aspect of the present invention relates to a method of identifying a candidate plant suitable for breeding with a cytoplasmic male sterile 30 plant. The method involves analyzing the candidate plant for the presence, in its genome, of the above nucleic acid molecule of the present invention.

**[0019]** Yet another aspect of the present invention relates to a method of identifying a candidate gene restoring fertility in plants. The method involves analyzing the candidate gene for the presence of the above nucleic acid molecule in accordance with the present invention.

5 **[0020]** The present invention also relates to a method of producing hybrid plant seed. The method first involves providing a cytoplasmic male sterile plant. Next, a second plant containing the above nucleic acid molecule in accordance with the present invention is provided. Finally, the cytoplasmic male sterile plant and the second plant are bred under conditions effective to produce hybrid progeny seed  
10 which yield fertile plants.

15 **[0021]** Another aspect of the present invention relates to a method of producing plant seeds for an inbred line of plants. The method first involves providing a cytoplasmic male sterile plant. Next, a second plant containing the above nucleic acid molecule in accordance with the present invention is provided. Then, the cytoplasmic male sterile plant and the second plant are bred under conditions effective to produce hybrid progeny seed which yield fertile plants. Next, hybrid fertile plants are produced from the hybrid progeny seeds. Finally, the hybrid fertile plants and the second plant are backcrossed to produce seed which yield inbred progeny plants.

20 **[0022]** Yet another aspect of the present invention relates to a method of directing gene expression to plant mitochondria. The method involves transforming a plant with a chimeric nucleic acid molecule containing a transgene operatively linked to a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of  
25 the transformed plant. The promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1. The terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

30 **[0023]** The present invention also relates to a promoter from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant. The promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1.

5 [0024] Another aspect of the present invention relates to a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant. The terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

10 [0025] Yet another aspect of the present invention relates to a nucleic acid construct. The nucleic acid construct includes: (i) a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant and (ii) a nucleic acid heterologous to and operatively coupled to the promoter or the terminator. The promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1. The terminator has a nucleotide sequence of from nucleotide 3761 to 15 4593 of SEQ ID NO: 1.

20 [0026] The present invention also relates to a method of expressing a gene preferentially in roots of a plant. The method involves transforming a plant with a nucleic acid construct. The nucleic acid construct includes a promoter suitable for driving expression preferentially in roots having a nucleotide sequence of from 1 to 1388 of SEQ ID NO: 44; a nucleic acid heterologous to the promoter, where the promoter is operatively coupled 5' to the nucleic acid to induce transcription of the nucleic acid; and a terminator having a nucleotide sequence of from nucleotide 3168 to 4016 of SEQ ID NO: 44, where the terminator is operably coupled 3' to the nucleic acid.

25 [0027] Another aspect of the present invention relates to a method of altering plant floral morphology in ornamental plants. The method involves transforming an ornamental plant with the above nucleic acid molecule in accordance with the present invention.

30 [0028] Another aspect of the present invention relates to a method of producing plants with a cytoplasmic male sterile plant restoration system. The method first involves transforming a first plant in its chloroplast genome with a nucleic acid which causes the plant to become male sterile. Next a second plant is transformed with the above nucleic acid molecule in accordance with the present

invention whose protein product is targeted to the chloroplast. Finally, the first and second plants are crossed to produce progeny plants possessing a cytoplasmic male sterile plant restoration system.

[0029] Another aspect of the present invention relates to a method of  
5 producing plants with a cytoplasmic male sterile plant restoration system. The  
method first involves mutagenizing a first plant having a nucleic acid which encodes a  
protein. The protein has a motif having an amino acid sequence corresponding to any  
10 of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME  
software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid  
sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST  
software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input. Next,  
15 the mutagenized first plant is crossed with a wild-type plant having mitochondrial  
DNA polymorphisms compared to mitochondrial DNA in the mutagenized first plant  
to produce progeny plants. Finally, it is determined if the progeny plants are fertile,  
whereby fertile progeny plants can be used as a fertile maintainer line, where the  
15 mutagenized first plant, the fertile maintainer line, and a wild-type allele present in  
the first plant before mutagenesis comprises a new cytoplasmic male sterile plant  
restoration system.

[0030] The present invention also relates to an isolated nucleic acid sequence  
20 corresponding to SEQ ID NO: 42 or SEQ ID NO: 44.

[0031] The present invention identifies nucleic acid sequences which encode  
the gene for restoration of fertility to cytoplasmic male sterile plants. This gene  
modifies the expression of the mitochondrial genome and is the first such gene  
sequence that has been identified. In petunia, the gene may be transferred to lines  
25 lacking the gene in order to restore fertility. More importantly, the gene sequence has  
characteristics that can be used to identify comparable genes from economically  
important species. Thus, the gene and the sequence information may be used to  
develop hybrid seed production systems in economically important plants.  
Furthermore, the information may be used in crop improvement by controlling  
30 mitochondrial gene expression.

**BRIEF DESCRIPTION OF THE DRAWINGS**

**[0032]** Figures 1A-D show that the *Rf* locus contains two tandem mitochondrially targeted PPR motif genes. Figure 1A illustrates the genomic organization of the region containing the *Rf-PPR592* and *Rf-PPR591* genes.

5 Duplicated blocks are indicated by similar shading. Arrows indicate the direction of transcription. 1 and 2 show locations of the primers used to amplify the *rf-PPR592* gene from a CMS plant. Figure 1B shows a single onion epidermal cell expressing a known mitochondrially targeted green fluorescent protein (GFP) after DNA bombardment. Figure 1C shows a single onion epidermal cell transiently expressing

10 44 N-terminal amino acids of *Rf-PPR592* fused to GFP. Figure 1D shows a comparison of PPR motifs found in *Rf-PPR592* with the MEME-derived consensus from 1,303 PPR motifs. The 14 PPR repeats are sorted by decreasing statistical significance, with PPR 230-264 showing the highest match to the consensus motif that is generated by retaining only the amino acids that occur at least in 6 of the

15 14 repeats.

**[0033]** Figures 2A-B show the genetic structure of the *rf-PPR592* gene. Figure 2A illustrates that a comparison of *Rf-PPR592* and *rf-PPR592* reveals a size polymorphism. The first lane was loaded with the *Rf-PPR592* PCR amplicon obtained from a restorer line (*Rf/Rf*), the adjacent lane was loaded with the *rf-PPR592*

20 PCR amplicon obtained with the same primer pair from a CMS line (*rf/rf*). Figure 2B illustrates that a comparison of *Rf-PPR592*, *Rf-PPR591*, and *rf-PPR592* reveals five similarity blocks. For each block, (I to V), the two blocks that exhibit the greatest similarity are shown with the same shading. Overall all three sequences are greater than 90% identical at the nucleotide level except in block V, where *Rf-PPR591*

25 exhibits only 23% identity to the other two genes. The locations of 47- and 49-nt deletions in *Rf-PPR591* and 47- and 530-nt deletions in *rf-PPR592* with respect to the *Rf-PPR592* sequence in blocks I and II are shown as lines.

**[0034]** Figures 3A-B show the expression pattern of *rf-PPR592* and *Rf-PPR592*. Figure 3A depicts the examination of floral bud RNA for expression of *rf-PPR592* and *Rf-PPR592*. RT-PCR of floral bud RNA of a CMS plant (S) with primers specific to *rf-PPR592*, and RT-PCR of floral bud RNA of an *Rf/Rf* (nontransgenic) fertile plant with primers specific for *Rf-PPR592* (R). DNA, positive

control for the amplification where the substrate is leaf DNA from a CMS plant; M, mass markers; 0, no template added, negative control. Figure 3B depicts the examination of different tissues for expression of *rf-PPR592*. RT-PCR of RNA from different tissues of a CMS plant with primers specific to *rf-PPR592*. DNA, M, and

5 0 are same as in Figure 3A.

[0035] Figures 4A-D illustrate the restoration of fertility to CMS *Petunia* lines by transformation with a 4.6-kb genomic sequence carrying *Rf-PPR592*. Figure 4A shows the flower of *P. parodii* CMS line 3688. Figure 4B shows the regenerant carrying *Rf-PPR592*. Figure 4C shows the *P. hybrida* CMS line 2423. Figure 4D shows the regenerant carrying *Rf-PPR592*.

[0036] Figures 5A-B illustrate the cosegregation of the *Rf-PPR592* transgene, restoration of fertility, and reduction of PCF. Figure 5A shows the DNA blot hybridized with an *npt* II transgene-specific probe. Lane 1, *P. parodii* CMS line 3688; lanes 2-and 3, sterile T<sub>1</sub> progeny of transformed *P. parodii*; lanes 4-9, fertile T<sub>1</sub> progeny. Figure 5B shows the immunoblot of floral bud proteins probed with anti-PCF antibody. Lanes are as in Figure 5A.

[0037] Figures 6A-B illustrate abnormal flowers on plants obtained by introducing *Rf-PPR592* into a CMS background. Figure 6A shows a petaloid flower on a plant carrying a recombination event near the *Rf* locus, affecting the region 5' to 20 *Rf-PPR592*. Figure 6B shows an abnormal flower on a plant carrying the CMS cytoplasm and the 4.5 kb *Rf-PPR592* transgene.

[0038] Figures 7A-B show methods for creating a new CMS/restorer system.

[0039] Figure 8 shows a two-line method for hybrid rice production, using an engineered inducible restorer gene.

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## DETAILED DESCRIPTION OF THE INVENTION

[0040] The present invention relates to an isolated nucleic acid molecule which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant.

30 [0041] One form of the nucleic acid molecule of the present invention is a nucleotide sequence of from nucleotide 1982 to 3760 of SEQ ID NO: 1, identified herein as *Rf-PPR592*, as follows:

1 ATATATATATACAAACTGATTTCTGTCTATTGCACAGTGTATTTACATACCCCTGAAAAAGGGTAGCTCCGCT  
 81 AATAATGTTATCTTACAAAAAAACAATAATTTTACATAATATACAAAACCTATTGTTATGTATTGAAATAT  
 161 GATAAAAATATTGTTATTTGTAAATATAGCTATTAGGTAGTCATGTGGTGTAAATATTCTAAATATTACCTGAG  
 241 TCGGCCATTGGCTAAAATATTTATTTAGTATTGTATACCCCAATAGTATACAAGTTGACACCGCAATAGTGTACC  
 5 321 TACGATATCTCTATTACCTTTAGTATTGTATACCCCAATAGTATACAAGTTGACACCGCAATAGTGTACC  
 401 CCAATGTTGTGGGTGTGGCTATAAAATGTTAAACAAAATTGAGTGATGGTAGTGTAAATTTAGTGTAAAGCTGGG  
 481 TAGTTTAAACATTCTTTGAAATTGAGTCAAGTCAGTACAGTACAAAAAACTGAAATATTATGTTCTGATT  
 561 TTGGCTGGTCTCTAAAATTTGAAATGCTGGTAGTTTCAATTAGCGAGGGCAAATAGACTACATGCCAAATTT  
 641 TACGTTAAAGAAGTGTGTGGAAAGTATTGAAAGATTGACGGACAAGTGTGCTAGACAACACGTCAAATT  
 10 721 TGTAGAAAAATCTAGGAAGAAATTCAAAAGCAAATATTGCTTAAGCAGTCAGTCAGTCAGTCAGTCAGTCAG  
 801 TGAGAAATGGGCATCACTATAAGATTGTTTCATTGATATTTCATTATAAACTTAAGGAAAGTGTAAAGGAAA  
 881 GCCACTTTGGCTTGCCTTACCGTGAAGCTACTTCAGCAAGAAAAGAGCTAGTTTAGCTTTGAACTTTAAT  
 961 CATTGTGGGCCAGCTTCAGACCTTGTGGCCGAATTCACTACATTACAAGTAAAAATTAGTCACAGGCCACTTTA  
 1041 CCACTAGTATTGGTTGAAGTCATTGTTTACATGAGAGACCACTTTGAACTTCATCTTGTGCG  
 15 1121 TTGAACCTCATGCCCTAGTTAAAGTTCAACTCAATCCGTAAGGGCTGAATTAGGCTAGATGCGTAAACTTCAA  
 1201 CCTTGTGGACTGAAGTTGAACCTCGCCCTTATGGTGGCCTGAAGTTGAACCTCAATCCTGTGGCTGAACCTGTG  
 1281 AGTTCAACCCACAAGGATTAAAGTTCAAAATGACCTCTCAAGCAAATCTGCAAAAAAAAGTGGTCTCATGCACT  
 1361 TTTACCCATTCCAAGTAGGCTGAAGTCAGTCCAGCCCACAATTTCAGTCCAAAAAATTTCACAATATACCTCTTA  
 1441 TCTCGGTTATGATCTTGTATGATTAGCAGGAAAGTGGACTTCAGTCCAGGGCCATGGACTGAAGTTG  
 20 1521 GGTACAGGCCACTAATACCAAAATTAGTTAGTTAGGCTACTTTGCTTAAAGAGATAGAACCTCAGTCCAGAGGCC  
 1601 TTGAAGTTCAGTCCTTAAAGATTGAACCTCGATCCAGTGCCATATGGACTGAAGTCAGTCAGTCAGTCAG  
 1681 TCAGTCCAGGCCATGGACTGAAGTTCAATCCTAAAGATAGAACCTCAGTCCAGGGCCATGGACTGAAGTTCA  
 1761 TCAATTATCAGAACTTAAGTCAGTATTAGTAAAGGCCAAAGTGGTAGTATAAGACCAATAAAATAGAGGCC  
 1841 TAAAACAAATAACAGTGTAAAAGTGGCTGATGGACGAAATTCTACAAAATGGACTCGAGGTAGCAATTCAACTTCAA  
 25 1921 CCTATGGTGTCAAGTCGTACAATTCTCCAATCACCCCTACTAAGTGAAGTGAAGCGAAGATGATGAGAAATTGCA  
 2001 GTTACTGTCATGGTAATCCCTTTCTCATTGCTTCAATTGACCCGACATTATTCTACCAATACATGT  
 2081 TCCATTCAGTTAAAGGAATTGGGGTTCTAATGAATTGAGAATGTTAGTGTAGTGTGCTTCAGTTG  
 2161 CCGTCAAATGGTTACAACTAACCCCTTCCTGCTGTCTCTAAATTGTTGAAAGCTTGGTACATATGAAGC  
 2241 ATTACTCTCTGTTCTATTTCGAGAAATCCACAAATTACGTTACGTTGATGCTTCGCCCTGAGCACTGTG  
 30 2321 GTTAACAGTTGTTGCCCTATGCATCGTACCGATCTGGATTCTGTATTAGCCATTCAACTCAAGAAAGGTATTCCATA  
 2401 TAATGAAGTCACCTTACTACCTAATAAGGGACTTTGCTGAAAATAAGGTCAAAGATGCTGTTCAATTGTTCAA  
 2481 AGTTGGTGAGGGAGAATATATGTGAGCCTGATGAGTCAGTGTGCTGGACGGTCATGGATGGCTTGCAAGAAGGCC  
 2561 ACTCAAAAGCTTTGATTGCTCCGGTAATGGAACAAGGAATTACTAACGGCGATACATGCATCTACAAACATTGTTAT  
 2641 CGATGCCCTTGCAAAGATGGGATGGCTAGATGGTTGACGGAGATGAAACAAAAACATTCTCCAG  
 35 2721 ACATTATTACATACCTCATTGATCGATGGTTGGGTAAGTTAGTCAGTGGAAAAGGTTAGGACTTTGTTCTGAG  
 2801 ATGATACATCTTAATATTGATCCAGATGTGTCACCTCAACTCCGTATTGATGGACTATGCAAAGAGGGAAAGTGA  
 2881 AGATGCCGAGGAATAATGACATACATGATCGAAAAGGTTGAGAACCTAATGAGATAACCTACAATGTGTTAATGGATG  
 2961 GATATTGCTTGCCTGGTCAAATGGTAGAGCGAGGGAGAATTGGATTCATGATAGATAAGGGCATTGAGCCTGATATC  
 3041 ATTAGCTATACCGCACTAATAAAATGGATACCTGAGAAAAAGGATAAGGCCATGCAATTGTTGAAATTTC  
 40 3121 TCAAAATGGATTGAAACCTAGTATTGTTACCTGCAGTGTCTCTGGTGTCTTTGAAAGTGGAGAAACTGAATGTG  
 3201 CAAAAATATTCTTGTGATGAGATGCAAGCTGCCGGCACATACCTAATTATACACTCATTGCACTTGTGTTGGTTAT  
 3281 TTTAAGAATGGACTTGTGAAAGAGGCTATGTCACACTCCATAAGTGGAAAGGAGGAGAGAAGATAACAAATATTCAAAT  
 3361 TTACACGGCTGTCATTAAATGGATTGCAAAAGTGTGAAAGCTGACAAAGCTCATGCTACGTTGAGAAGCTTCCCTG  
 3441 TAGGCTTACATCCTGATGTGATAACACACTGCAATGATTAGTGGATATTGTCAAGAAGGGTTGTTAGATGAAGCTAA  
 45 3521 GATATGCTAAGGAAATGGAGGACAATGGTTGTTGCCAGACAACCGAACATACAATGTTATTGCGGGGATTTCAG  
 3601 AAGCAGTAAAGTTAGTGAAGGCTTCTGAGGAAATAGCTGGGAAGAGCTCTCATTGAGGCACTACTGTAG  
 3681 AGTTATTGATGGATATTAGCAGAGGATCCTTGTAAACATGATTCCAGAATTTCACCGGATAATAAGAAGTGA

3761 ATAACTTTGCACCTGTTTTTGACGATATCACCATTATTCTGCTATTCCTTCATCTTAGCAAAAGAAATTGCATC  
 3841 CAGTGGAAATTGCGGAAGCTGAAAAAATGGCAAGAAGAACATTGCTTAAGCTTCCTGGCAAGCTTATATCGGAGGGACAT  
 3921 CATTGGTGTGTTGGCTCTCTCTTATCTGGAAATCAAATGTTCTGCGCTCTTAATATCAGAAACAATGTGAACCT  
 4001 CCATATATGTACGAGTTATAAGTTCGGAATATGATTCAATGGTTAGTATTCTATTTGATATGGAATTAATTTT  
 5 4081 GAGCGACCCAGTGTTGACCATTGCCTACCTTCGGTTATTATATGATTGAAATTCCCTCCAATCTCCAATACTCACTTCAT  
 4161 TTTGTCTGTTGAATTTCATTTCTTTCTGTTACGATTGTCACTTCACCGCCTGAGTATCCATCAGGTTCCA  
 4241 GTTGGAAAAAGAACATCATTGGCCATGACCATCATGCTTCTGAGTGCAAGATCAAGAGAGGTACTTTCTCTAAGAA  
 4321 CCTCTGGTTTTAAGTGTCTGGGTCTTCAGTACTTTAAGCTATTTCTAATCCTTGAAGAGATTACATACATAT  
 4401 CTGTGCACTGTGTTGTTCTTTCTGGGTGATACTTGTGTTAGCTAAGGATTGAAAGGTAATTTCATTTCAT  
 10 4481 TAGCAATAGATATGAAACAGCTTGTAAAGGACTCTGGAGTCCTAAAATTTGGCTATGCAAATAGCCTATTGCATCA  
 4561 ATTTGTCGTTGAAATCCATGTATCATAAAAAAA

*Rf-PPR592*, isolated from *Petunia* has an open reading frame (“ORF”) of 1779 bp, extending between nucleotides 1982-3760.

15 **[0042]** The nucleic acid molecule of the present invention which has the nucleotide sequence of from nucleotide 1982 to 3760 of SEQ ID NO: 1 encodes a protein or polypeptide having a deduced amino acid sequence of SEQ ID NO: 2, as follows:

20 MMRIAVRYCLNGNPFFSFAYSIAPRHYSTNTCSISVKGNFGVSNEFENVKCLDDAFSLFRQMVTTKPLPSAVSFS  
 KLLKALVHMKHYSSVVSIFREIHKLRIPVDAFALSTVVNSCLMHRTDLGFSVLAIHFKKGIPYNEVTFTTLIRGL  
 FAENKVKDAVHLFKKLVRENICEPDEVMYGTVMMDGLCKKGHTQKAFDLLRLMEQGITKPDTCTYNTIVIDAFCKDGM  
 LDGATSLNEMKQKNIPPDIIITYTSILDGLGKLSQWEKVRTLPLEMIHLNIYPDVCTFNSVIDGLCKEGKVEDAEE  
 IMTYMIEKGVEPNEITYNVMDGYCLRGQMGRARRIFDSMIDKGIEPDIISYTALINGYVEKKKMDKAMQLFREIS  
 25 QNGLKPSIVTCVSLRGLFEVGRTECAKIFFDEMQAAGHIPNLYTHCTLLGGYFKNGLVEEAMSHFKLERRREDT  
 NIQIYTAVINGLCKNGKLDKAHATFEKLPLIGLHPDVITYTAMISGYCQEGLLDEAKDMLRKMEDNGCLPDNRTYN  
 VIVRGFFRSSKVSEMKAFLKEIAGKSFSFEAATVELLMDIIAEDPSLLNMIPEFHRDNKK

30 **[0043]** As shown in Figure 1D, most of the predicted mature protein (87%) of *Rf-PPR592* consists of 14 pentatricopeptide repeat motifs (PPRs). These repeats extend from the amino acid in position 54 to the amino acid in position 544 and are organized in two sets of tandem repeats, one set containing 3 PPRs from amino acid 54 to amino acid 158, the other set containing 11 PPRs from amino acid 160 to amino acid 544. Thus, another suitable nucleic acid molecule in accordance with the present 35 invention encodes a protein containing a motif having an amino acid sequence corresponding to any of the PPR motifs (SEQ ID NOs: 3 to 18), where SEQ ID NO: 3 is as follows:

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E E A . . L Y . . M . . . G . . P N . . T Y N A L I N A Y A K . G . .

where SEQ ID NO: 4 is as follows:

5 D . A . . . F . . M . . . G . . P D . . T Y . . L I . G L C K . G . .

where SEQ ID NO: 5 is as follows:

D D A F S L F R Q M V T T K P L P S A V S F S K L L K A L V H M K H Y

10

where SEQ ID NO: 6 is as follows:

S S V V S I F R E I H K L R I P V D A F A L S T V V N S C C L M H R T

15

where SEQ ID NO: 7 is as follows:

D L G F S V L A I H F K K G I P Y N E V T F T T L I R G L F A E N K V

where SEQ ID NO: 8 is as follows:

20

D A V H L F K K L V R E N I C E P D E V M Y G T V M D G L C K K G H T

where SEQ ID NO: 9 is as follows:

25

Q K A F D L L R L M E Q G I T K P D T C I Y N I V I D A F C K D G M L

where SEQ ID NO: 10 is as follows:

D G A T S L L N E M K Q K N I P P D I I T Y T S L I D G L G K L S Q W

30

where SEQ ID NO: 11 is as follows:

E K V R T L F L E M I H L N I Y P D V C T F N S V I D G L C K E G K V

35

where SEQ ID NO: 12 is as follows:

E D A E E I M T Y M I E K G V E P N E I T Y N V V M D G Y C L R G Q M

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where SEQ ID NO: 13 is as follows:

G R A R R I F D S M I D K G I E P D I I S Y T A L I N G Y V E K K K M

5

where SEQ ID NO: 14 is as follows:

D K A M Q L F R E I S Q N G L K P S I V T C S V L L R G L F E V G R T

10 where SEQ ID NO: 15 is as follows:

E C A K I F F D E M Q A A G H I P N L Y T H C T L L G G Y F K N G L V

where SEQ ID NO: 16 is as follows:

15

E E A M S H F H K L E R R R E D T N I Q I Y T A V I N G L C K N G K L

where SEQ ID NO: 17 is as follows:

20

D K A H A T F E K L P L I G L H P D V I T Y T A M I S G Y C Q E G L L

and where SEQ ID NO: 18 is as follows:

D E A K D M L R K M E D N G C L P D N R T Y N V I V R G F F R S S K V

25

A PPR motif-containing gene can be identified if it contains the consensus sequence (SEQ ID NOs: 3 or 4) or if it is found with a MEME software (Bailey et al., "Fitting a Mixture Model by Expectation Maximization to Discover Motifs in Biopolymers," Proceedings of the Second International Conference on Intelligent Systems for Molecular Biology, pp. 28-36, AAAI Press, Menlo Park, California (1994), which is hereby incorporated by reference in its entirety). To find whether a protein has a PPR motif with the MEME software, the parameters for motif searching should be set as minimum width=35, maximum width=35. MEME (Multiple Em for Motif Elicitation) is a software tool for discovering motifs in a group of related DNA or protein sequences (Bailey et al., "Fitting a Mixture Model by Expectation Maximization to Discover Motifs in Biopolymers," Proceedings of the Second

International Conference on Intelligent Systems for Molecular Biology, pp. 28-36, AAAI Press, Menlo Park, California (1994), which is hereby incorporated by reference in its entirety). MEME takes as input a group of DNA or protein sequences (the “training set”) and outputs as many motifs as requested. MEME uses statistical modeling techniques to automatically choose the best width, number of occurrences, and description for each motif. MEME represents motifs as position-dependent letter-probability matrices which describe the probability of each possible letter at each position in the pattern. Individual MEME motifs do not contain gaps. Patterns with variable-length gaps are split by MEME into two or more separate motifs.

10 [0044] Another suitable nucleic acid molecule for the present invention is a nucleic acid molecule which encodes a protein having an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input. Meta-MEME is a software toolkit for building and using motif-based hidden Markov models of DNA and proteins. The input to Meta-MEME is a set of 15 similar protein sequences, as well as a set of motif models discovered by MEME. Meta-MEME combines these models into a single, motif-based hidden Markov model and uses this model to produce a multiple alignment of the original set of sequences and to search a sequence database for homologs (Grundy et al., “Meta-MEME: Motif-based Hidden Markov Models of Biological Sequences,” Computer Applications in the Biosciences, 13(4):397-406 (1997), which is hereby incorporated by reference in its entirety).

20 [0045] Also suitable for the present invention is a nucleic acid molecule which encodes a protein having an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input for comparison (Fulton et al., “Identification, Analysis, 25 and Utilization of Conserved Ortholog Set Markers for Comparative Genomics in Higher Plants,” Plant Cell, 14:1457-1467 (2002), which is hereby incorporated by reference in its entirety).

30 [0046] Also suitable in the present invention is a nucleic acid molecule which has a nucleotide sequence of SEQ ID NO: 19, an 8.5 kb fragment containing *Rf-PPR592* capable of transforming cytoplasmic male sterile plants, as follows:

GGATCCAAAATTCACTAAAGGTTAACCGCGAGGAACTGAAGTTGGAGAGCAATGTGGTATCTGGTCATGGAC  
GGAGTCATGGGGTATAGTTGCTGGCTATTAGCTAACTGGAGAACTGTTGTGAGGAATTATTTAAAGAACATGCTAC  
TTTCTCGTCCACATAAACATGTCAAATATTTCTACTGTGATAGAGAGTCAAGGAAATAGTGTGGATTCTCC  
AAACACAAACGATTGAGAAAAGTGAAGTGAAGGCTGAAGAGAAAAGTAAAGAGAACTGGAAGCTAAGAACACAG  
5 AAGCACAAACCTATAAACATAGACACTGGCATGTCAGAAAATTAACTTCGATTCTCCAGTAGAAAGACACA  
AATACATCAGTAAATTCTTAGGCTCAAGCAAGGATACTCTGGTAGAATTGCAATTACCAACATAATAAG  
CTCAAAAAAAATAACTAAGCTGCAACTAGCTACTTGGTCCGAGAAGCTTGTATCAGGAAGTCCACAGTCCAA  
AACCAAGGTAGACTATCAAGCTAGTCCCACCAGTCATTTCTTAGACTTGTCTCACGATAAACTAAGATCATT  
TTTTATGATACATGGATTCAACGACAAATTGATGCAATAGGCTATTGCAAGCAGAAAATTAGGAGACTCC  
10 AGAGTCCTTACAAAGCTGTTCATATCTATTGCTAATGAAAATGAAAATTACCTTTCAATCCTAGCTATAAAC  
AAAGTATCACCCGAAAAAAAGAAACAAACATGCACAGATATGTATGAATCTCTCAAAGGATTAGAAAATAGCT  
TAAAAGTACTGAAAGAACCCAGAACACTAAAAACCAAGAGGTTCTAGAGAGAAAAGTACCTCTTGATCTG  
CACTCAGAAAGCATGATGGTCATGGCAAAATGATTCTTCAACTGGAACCTGATGGATACTCAAGGCAGTGA  
AAATGACAATCGTAACAGAAAAAGAAAATTGAAAAATTCAACAAGACAAAATGAAGTGAATGGAGATTGGAG  
15 GGAATTCAATCATATAATAACCGAAGGTAGGCAATGGCAACACTGGTCGCTCAAAATTAAATTCCATATCAA  
AATAGAATACTGAAACCATTGAAATCATATTCCGAAACTTATAACTCGTACATATGGGAGTTCACATTGTTCT  
GATATTAAGAGCGCAGAACATTGATTCCAAGATAAAAGAGAGAGCCAAACACCAAAATGATGTCCTCCGAT  
ATAAGCTTGCAGGAAAGCTTAAGCAATGTTCTTGCATTTTCACTGCTTCCGCAATTCCACTGGATGCAATT  
TCTTGTCAAGATGAAAGGAAATAGCAGAATAATGGTGTATCGTCAAAAAAAACAGGTGCAAAAGTTATTCACT  
20 TCTTATTATCCGGTGAATTCTGGAATCATGTTAAGCAAAGAAGGATCCTCTGCTATAATATCCATCAATAACTC  
TACAGTAGCTGCCTCAAATGAGAAGCTTCCCAGCTATTCTCAGAAAAGCCTTCATTCACTAACTTACTG  
CTTCTGAAAATCCCCGACAATAACATTGTATGTCGGTTGTCTGGCAAACACCAATTGTCCTCCATTCTTCA  
GCATATCTTAGCTCATCTAACACCTCTTGACAATATCCACTAATCATGTCAGTGTATGTATCACATCAGG  
ATGTAAGCCTATCAAGGGAAAGCTCTCAAACGTAGCATGAGCTTGTGAGCTTACCAATTGGCACAATCCATTA  
25 ATGACAGCGTGTAAATTGAATATTGTATCTCTCTCCCTTCCAACCTATGGAGTGTGACATAGCCTCTT  
CAACAAGTCATTCTAAAATAACCAAGCAAAGTCAATGAGTGTATAAATTAGGTATGTGCCCGCAGCTG  
CATCTCATCAAAGAATATTTCGACATTCAAGTCTTCAACTTCAAAAGACCACGCAAGAGAACACTGCAGGTA  
ACAATACTAGGTTCAATCCATTGGAGAAATTTCACGAAACAATTGCAATGCCCTATCCATTTCCTTCG  
CGTATCCATTATTAGTGCAGGTTAGCTAATGATATCAGGCTCAATGCCCTATCTATCATGGAATCAAATTCT  
30 CCTCGCTTACCCATTGACCACGCAAGCAATATCCATTACCAATTGTAGGTTATCTCATTAGGTTCTACA  
CCTTTTCGATCATGTATGTCATTATTCCTCGGCATCTCAACATTCCCTCTTGCACTGTCATGACGG  
AGTTGAAGGTGCACACATCTGGATAAATATTAAGATGTATCATCTCAAGGAACAAAGCTCTAACCTTCCACTG  
ACTTAACCTACCAAACCATCGATCAATGAGGTATATGTAATAATGTCGGAGGAATGTTTTGTTCTATCTCG  
TTCAAAAGGCTGGTAGCACCCTAGCATCCCATTGCAAAAGGCATCGATAACAATGTTGAGATGCATGTAT  
35 CGGGCTTAGTAATTCTGTTCCATTAACCGGAGCAAATCAAAGCTTTGAGTATGCCCTCTGCAAAGCCC  
ATCCATGACCGTCCCACATGACTTCATCAGGCTCACATATATTCTCCCTACCAACTTTGAACAAATGAACA  
GCATCTTGACCTTATTCAGCAAAGTCCCTTATTAAGGTAGTAAAGGTGACTTCATTATATGGAATACCTT  
TCTTGAAGTGAATGGCTAATACAGAAAATCCGAGATCGGTACGATGCATAAGGCAACAATGTTAACCAAGCT  
CAAGGCAGAACATCAACAGGAATCGTAATTGTCGGATTCTCGAAAATAGAAACAACAGAAGAGTAATGCTTC  
40 ATATGTACCAAGCTTCAACAATTAGAGAAAGAGACAGCAGAAGGAAGGCTTAGTTGTAACCATTGACGGA  
ACAAACTGAAAGCATCATCTAAACACTAACATTCTCAAATTCAATTAGAAACCCAAAATTCCCTTAACGAAAT  
GGAACATGTATTGGTAGAATAATGTCGGGTGCAATTGAAATAAGCAAAGAATGAGAAAAAGGGATTACATTGAGA  
CAGTAACGCACTGCAATTCTCATCTCGCTTCACTTAGTAGGGTGATTGGAAGAATTGTACGACTAT  
GACACCATAGGTTGAAGTTGAATTGCTACCTCGAGTCCATTGAGAAATTGTCATCAGCCACTTTAACAC

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TGTTATTTAGTTAGGCCTCTATTTTATTGGCTTATACTAACCACTTTGGGCCTTACTAAATAAAACTGA  
CTTAAGTTCTGATAATTGACTGAACCTCAGTCCATACGGCCCTGGACTGAAGTTCTATCTTAAGGATTGAACCT  
CAGTCCATATGGCTCTGGACTGAAGTCCATCTTAAGGACTTGACTGAACCTCAGTCCATATGGCACTGGATCGAA  
GTTCAATCTTAAGGACTGAACCTCAATCCGGCTCTGGACTGAAGTTCTATCTCTTAAGCAAAAGTAGGCCACAA  
5 ACTAAATATTTGGTATTAGTGGCCTGTACCCAAAGACCAATGGCAAAAGTGGTCTTCGTGCACTTCCCGTCC  
ATTTGCTAAATCATAACAAAGATCATAACCGAGATAAGGAGGTATATATTGTGAAATTTTTGGAACCTGAATAA  
TTGTGGGCTGAACCTCAGCCTACTTGCGAATGGTAAAGTGCATGAGAGACCACTTTTTGCAAGATTTGCT  
TGAGAGGTCACTTTGAAACTTAAATCCTGTGGGTGAACCTCACACAAGTCAGGCCACAAGGATTGAAGTTC  
AACTCAGGCCACCAAAAGGGCGAAGTTCAACTCAGTCCACAAGGTTGAAGTTACGCATCTATGCCTAAAAAT  
10 TCAGCCCTTACGGATTGAAGTTGAACCTAAACTTAGGCATGAAGTTCAAGCGCACAAAGATTGAAGTTCCAAAA  
AGTGGTCTCTCATGTAACCAATAAAAGATGACTCAAACCAAATACTAGTGGTAAAGTGGCTGTGAGCTAA  
TTTTTACTTGTGAATGTATGAAGTTGGCCCACAAGGTCTGAAGTTGGCCACAATGATTAAAGTTCCAAAAAA  
GCTAAAAACTAGCTCTTTCTTGAAAGTAGCTCACCGTAAAGGCAAGGCCAAAAGTGGCTTCTGCAC  
TTTCCTTAAGTTATAATGAATAAAATCGAATGGAAATAACATCTTATAGTGTATGCCCTACACTCCCTAAG  
15 GCAGCATTGTCTTGACTGCCTTTGCTTAAGCAATATTGCTTTGAATTCTCCACATTTCTACATAATT  
TGACGTGTTGCTAGGCAACACTGTCCGTACAATCTTCTGAATACTTCCCAGACAACACTTCTTAACGTA  
AAATTGGCCATGTAGTCTATTGCCCCCTCGCTAAATGAAAAGTGCAGCATTCAAAATTTAGAAGGACAGC  
CAAAATCAAGAACATATAATATTCAAGTTCTGACTATGACTACACTACAATTTGTTAAACATTTATAGCCACACAAC  
20 CACAACATTGGGTACACTATTGGGTGTCAAACCTGTATACTATTGGGGTATACGAATAACTAAAGTAAATA  
ATAGAAGATATCGTATAGGTATACTGCGCTATGGATATAACAGCTTGGAGTATATGCGACTATAAAATAAA  
TTTTTAGCAAATGGCGACTCAGGTAATATTAGGAAATATTACACCACATGACTACCTAATAGCTATATTA  
CAAAAATAACAATATTTCATATTACAATACATAACAATGAGTTGTATATTATGTAAGGAAATTATT  
GTTATTTTTGAAAGATAACATTATTAGCGGAGCTACCTTTCAAGGGTATGTAATATAACACTGTGCAAAT  
25 AGACAGAAAAACTAGTTGTATATATATCAGATATTGATCCCCCTCATTTCGTATGTTACTTTTA  
TATTATATATCCCTTAGTAAAAACTGGCTCCGCCACTGCCAGTAAGGTAGTATTAGTTGGCTCGCTAATAA  
AGTAACATCTACGTTATTTCATCAACATTAAAAGGAAGATTCACTATCCACATAGGCATCATCATTATCAA  
AGAATATCAGTTCATACATTGTATATATAACTTCTCAAATAACTAATTAAAAGTAAAGTACATTAAAAGG  
AAGATTCACTATCCTTTAATATTTCGTATATTACTTATTTGATACTCCTAGTAAAAACTGGCTC  
30 CGCCACTACCAGTGTAAATTAAATTGCGTCCCTCACTAAAGTAACACCTATAATTAAATTTCATGAAGTCA  
GAGTTAGCATTGGAAAGGGATATAAGCACATGCATTGTGTATATATATAACTTGCTCAAATAAAACTAACTTAA  
AATGAAATTTCATTTCTAGTACAATGAACATGCATCAATGCGTAATTAGTTGAGGTGGCTATATGAATATG  
TTATTAATTGAAAGCAAACATAAAACTGATAGAAGAATTTCACCTAAAATTGAACTTGAGCTGCTTCAGT  
TACTATCTCATTTCACTATATATGTTGTATCAGCTAATTCTATGATTAATTAAACAAATTGTAAGTATTAAAC  
35 AAAATAACGAATAAAATGGAAAATAAGTACTTGATGAACGTAGGGCCGGAGTTGGCCGAGGTGACCGGAGACAAT  
GGAAAGCAGAGTTACTATTGGACTAAATGCCACAAAAGAATCATTGTTTACAATGTAGCAAGTTGGCACGA  
TTATGATTCTGACACAATGCCACATTAGAAAGATAATGTCGACTAATGAGGTAAATTTCATTATGGAATGA  
TAACAAACAAATAAGTACAGGATAAAACAAACTGAAAGGGTTGCGCGATAATTAACTTGTGTTTACGAAAA  
TACTCATCAAATCAAATATTATCTGCCCTGCGATGTAAGTTCTATTTACTCGCTTGCCTAAATTATG  
40 AAAAATTACTCACACCGGATATATACCTAACCATAGCAATTAACTATGTAATGTGATGTAATGAAGAGAGA  
ATGTCTATTAATTAAATTATGGCTAAGTGATAGTGTATTGTAACAAATGACGTGCAATTGTTGATTAGACACT  
TACAAAATACCCACGAAATCTAAATAATTACAGCCACTATCCACTACTTCAAATATTATCTGGCCTACCCATT  
AAAATATTACTCACTACCCCTCCAGACTTATATTAAAGGTATAAAAGGTAAACAAAGGTATAACAAATAAAATGGCCT  
CCAGACTTTATACCATAATTGCAAGCCTAAAGGTACACACCTATAACAAAGGTATAACAAATAAAATGGGT

ATGTTGGTAAATACTTTAGTTTATGGGTAGAGTAATTTAATGGGTATGACTTGAAATACTTAAATTCA  
 TGGGTATAGAGTGTAAAATTCTTGTGATTGGGTATATACACCCGATGTGGGTAAAGTACTTGCTAATTTG  
 CCCTAAGGTAAATATAGACTTATAGTATAAAGGGTACACATTGAAAGCTCAATATTTATT  
 5 CCAAAGAATAAGAGACCAAACAATGTGTTGAGATTATTACTTGTGTCACAAACTAAAAGAAAATTAG  
 AAGTCTAAATTACAATACTTAACATGCATTTACGAATAAATATCACAAATCTCAAACATTAGAGATAATGT  
 CGTGGATGATGTTAACATATTGACTACACAACCCATTGTACAATAATTGAAGCATGTATATGCACGACCAAGA  
 CTCCATCATCATAGATCAAATGAATGTCATTTAATGCATGAAACCTAAGTAGAACATTATGCCTTAATGAAC  
 AAAACCAAGCAAAAGATACTACTTGTGCAATTGAATGAATTCTACCGTATATACTAATATACACCAGAGGTT  
 10 AGTTAACACTTGGAACTTCAAAAGGTGTACAACCAGAGTTCTTACATTGATGGTTCTTCATTCACCA  
 ACTGATAAAATGAAGGCTGGTATAGTCTACAAATCCCTAGTCCCTGTGAACTTGATCCCTCTAGCTACATGC  
 AGAACATGTCCTTAGATCCATAGGTGTATTGCCATTGCCACTGAACAATGGAGGACAATGTATAATTGTCCTC  
 CTCACCCATTGCACATACTCTGTCATTGCTGCACATCTACATGCCTTCTGAATATTCTCTGAGTCACAAAG  
 CATCATGAGACATCTGCAAGTATTTGAATGGGATAATCACATTCCAAATCGAAAGGTTCTGTCTTAACAAAG  
 15 TCAAGCTGCATCTGACAAAGAGACTCGTTGATGAAATGCCACATAAGAGCACATGCAAACCAAGTTGTTAAA  
 GCATTGTACACATATAACCTACTTCTGATGTGAATAAAGAAAGTCGATCAATGACAGAGGAAACAGTCACATCTAT  
 AACACAAAAAAATCTTTCTTAAAGTACGACCAACATAGATAACTATAATTCCCGTAGATGTCACAAAGGATTCTTAATGATGGTAA  
 GAATAAAAATAAACAAACATGTAATTGCTCATTATCCGACTGTACAAGGATTCTTAATGATGGTATAA  
 TAGGAGCAATCCCTTATGACAGATGCACTAATTGTTGGGTGCATATTCAATGCAAGACTGTGGGTATATA  
 20 ATCTAAAATATCATTCAAATCAAACCTGGGACGATTGAGAGAAGATTAGCATGGCCTCTGCACAAGGATGACACG  
 CATAAATCGAGAAATGTCACAAATAAGGAAATATATATTACCTGTTCAATTGCATAGTTCTAAAGAAGTT  
 TTGGCAGTTAAAGTATTAATAGTTACCTGTTGATTGACAGGAGGAACTGGCTGGGACGGACCTG  
 TGATTATTCTGCTAATCTCCTGTATATTGCAATGTGCAGTTAATCCAGTGCATTTCGCTGTTATGGATGG  
 ATCC

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**[0047]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr1*, which has a nucleotide sequence of SEQ ID NO: 20, as follows:

30 ATGGCGCGCCGCGTCCCTACCCGCCCGCGCGGGTGGCGGGCGGCGTCCCACGCTCGGAGGGCTCGATCCAAG  
 GGCGAGGAGGCCGCGGGGGCAGTGGCGCCGAGGACGCACGCCACGTGTTGACCAATTGCTCCGGCGTGGCAG  
 GGGCGCCTCGATCTACGGCTGAAACCGCCCTCGCCGACGTGCGCGTACAGCCCCGCGGCCCGTGTCCC  
 TACAACCGCATGGCCCGAGCCGGCGCCGCAAGGTAACCTCCACCGTGCACACCTATGCCATCCTCATGGCGTCT  
 GCTGCCGTGCGGGCGCTTGGACCTCGGTTCGCGGCTTGGCAATGTCGTCAAGAAGGGATTAGAGTGGATGC  
 35 CATCACCTTCACTCCTCTGCTCAAGGGCCTCTGCGACAAGAGGACGAGCGACGCAATGGACATAGTGC  
 AGAACATGACCGAGCTCGGCTGCATACCAAGATGTTCTCCTACAATAATCTCTCAAGGGCTGTGATGAGAAC  
 GAAGCCAAGAAGCTCTCGAGCTGCTGCACATGATGGCTGATGATCGAGGAGGAGGTAGCCCACCTGATGTGG  
 GTATAACACTGTCCTCAATGGCTCTCAAAGAGGGGATTGACACAAAGCTTACAGTACATACCATGAAATGCT  
 GACCGGGGGATTACCAAGATGTTGACCTACAGCTCTATTATTGCTGCCATTGCAAGGCTCAAGCTATGGACA  
 40 AAGCCATGGAGGTACTTAAACACCATGTTAAGAATGGTGTGATGCCTGACATGACATATAATAGTATTCTGCA  
 TGGATATTGCTCTTCAGGGCAGCCAAAGAGGCTATTGGAACACTCAAAAGATGCGCAGTGTGGCGTCAACCA  
 AATGTTGTTACTTATAGTTCACTGATGAATTATCTTGCAAGAATGAAAGATCCACCGAAGCTAGAAAGATTTC  
 ATTCTATGACCAAGAGGGCCTAGAGCCTGATATTGCTACCTATCGTACCCGCTTCAGGGGTATGCTACCAAGG

AGCCCTGTTGAGATGCATGCTCTGGATTGATGGTACCAAATGGTATCCAACCGGATCATCATGTATTCAAC  
 ATTCTAATATGTGCATACGCTAAACAAGAGAAAGTAGATCAGGCAATGCTTGTATTCAAGAAAATGAGGCAGCATG  
 GATTGAATCCGAATGTAGTGTGCTATGGAACAGTATAGATGACTTCAAGTCAGGCAGTGAGATGATGCTAT  
 GCTTATTGAGCAGATGATCGATGAAGGACTAACCCCTAACATTATTGTGTATACCTCCCTAACATGGCTG  
 5 TGACACCTGTGACAAATGGGACAAGGCTGAAGAGTTAACATTCTGAAATGTGGATCGAGGCATCTGTGAACACTA  
 TTTCTTAATTCAATAATTGACAGTCATTGCAAAGAAGGGAGGGTATAGAATCTGAAAAACTTTGACTGAT  
 GGTACGAATTGGTGTGAAGCCGATATCATTACGTACAATACTCATCGATGGATGCTGCTAGCTGGTAAGATG  
 GATGAAGCAACGAAGTTACTTGCAGCATGGTCTAGTTGGGTGAAACCTGATATTGTTACCTATGGCACCTTGA  
 TTAATGGCTACTGTAGAGTTAGCAGGATGGATGACGCATTAGCTTTCAAAGAGATGGTGAGCAGTGGTGTAG  
 10 TCCTAATATTACGTATAACATAATTCTGAAAGGTTATTTCATACCAGAAGAACTGCTGCTGCAAAAGAACTC  
 TATGTCAGTATTACCAAAAGTGGAACACAGCTGAACTTAGCACGTACAACATAATCCTTCATGGACTTGC  
 ACAATCTCACTGACGAGGCACCTCGAATGTTAGCAGGATTTACAGCTGGAGACTAGGACTTT  
 TAACATTATGATTGGTGCCTTACTTAAATGTGGAAGAATGGATGAAGCTAAGGATTGTTGCTGACTCGGCT  
 AACCGTTAGTGCCAGATGTTAGGACCTACAGTTAACAGGAGAACTTATAGAGCAGGGTCGCTAGAAGAAT  
 15 TGGATGATCTATTCTTCATGGAGGAGAATGGCTGTTCCGCCACTCCGCATGCTAAATTCCATTGTTAGGAA  
 ACTGTTACAGAGGGGTGATATAACCAGGGCTGGCACTTACCTGTTACGATTGATGAGAAGCATTCCCTCGAA  
 GCATCCACTGCTTCCTTCTGTTAGAATCTCCCCAATCGCTGGAGCAAATATCAAGAATATCACACTTGTCTG  
 TAAATTGAAATTAAAGCAGCCAAATGCACCTGTGAGTTAGGCCAAAGTGGTCCAAATCTGCCTAAACC  
 TGGCACAAATTGGTCGGTAGTGTGCGCACAGTTCACTTATCGCGGGGGTATCGCCTTACCGCGGGGTACG  
 20 ACGTTACCGCACTACCGCAGGGTGACGTTAACCCGGCCAAACGATAAGTAAACCTGGTCGACAAATTGG  
 CCCAAACCGACCAGTTATCGCCTACCGCGGGATGCCTCAGTAGGACCTTAG

*Rhpr1* is a rice homolog of the *Petunia Rf-PPR592* gene.

**[0048]** The nucleic acid molecule of the present invention which has the  
 25 nucleotide sequence of SEQ ID NO: 20 encodes a protein or polypeptide having a  
 deduced amino acid sequence corresponding to SEQ ID NO: 21 as follows:

MARRVPTRPRGGGGGGVPRSEGSIQGRGGRAGGSGAEDARHVFDELLRRGRGASIYGLNRALADVARHSPAAVSR  
 YNRMARAGAGKVTPTVHTYAILIGCCCRAGRDLGFAALGNVVKKGFRVDAITFTPLLKGLCADKRTSDAMDIVLR  
 30 RMTELGCIPDVFSYNNLLKGLCDENRSQEAELELLHMMADDRGGSPPDVSYNTVLNGFFKEGDSKAYSTYHEML  
 DRGILPDVVTYSSIIAALCKAQAMDKAMEVLNTMVKNVGMPDCMTYNSILHGYCSSGQPKEAIGTLKKMRSDGVEP  
 NVVTYSSLNYLCKNKRSTEARKIFDSMTKRGLEPDIATYRTLLQGYATKGALVEMHALLDLMVRNGIQPDHHVFN  
 ILICAYAKQEVDQAMLVFSKMRQHGLNPNVVCYGTIDVLCKSGSVDDAMLYFEQMIIDEGLTPNIIVYTSLIHGL  
 CTCDKWDKAEEILEMLDRGICLNТИFFNSIIDSHCKEGRVIESEKLFIDLMVRIGVKPDIITYNTLIDGCCLAGKM  
 35 DEATKLLASMVSVGVKPDIVTYGTLINGYCRVSRMDDALALFKEMVSSGVSPNIITYNIILQGLFHTRRTAAAKEL  
 YVSITKSGTQLELSTYNIILHGLCKNNLTDEALRMFQNLCLTDLQLETRTFNIMIGALLKCGRMDEAKDLFAAHSA  
 NGLVPDVRTYSLMAENLIEQGSLEELDDLFLSMEENGCSADSRMLNSIVRKLLQRGDITRAGTYLFMIDEKHFSL  
 ASTASFLLESSPIVWEQISRISHLSVNLKLIKQPKCTCELGPKWSQNLPKPGTNSVGSVAQFHLRSRGGYRAYRGGT  
 TVTALPQGDGNPGPNDKVNPGRTNLAQNRPVIALPRDASVGP

**[0049]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr2*, which has a nucleotide sequence of SEQ ID NO: 22, as follows:

5 ATGGCGCGCCGCCGCTTCCCGCTCCGCCGGCGCTGGGCCCTCGCTGGAGGGCTGACCCAAGGGC  
GAGGGGGCGCAGGGGGGAGTGGCGCCGAGGACGCACGCCACGTGTCGACGAATTGCTCCGGCGTGGCAGGGG  
CGCCTCGATCTACGGCTTGAACCGCCCTCGCCGACGTCGCCGTACAGCCCCGCCGCCGTGCTCCGCTAC  
AACCGCATGGCCGAGCCCGCAGGTAACCTCCAACCTGTGACCTACGGCATTCTCATCGGTTCTGCT  
GCTGCGCGGGCCGCTTGGACCTCGGTTCGCGGCCCTGGCAATGTCATTAAGAAGGGATTAGAGTGGACGCCAT  
10 CGCCTCACTCCTCTGCTCAAGGGCCTCTGTGCTGACAAGAGGACGAGCGACGCAATGGACATAGTGTCTCCGAGA  
ATGACCCAGCTGGCTGCATACCAAATGTCTTCTACAATATTCTCTCAAGGGCTGTGTGATGAGAACAGAA  
GCCAAGAAGCTCTCGAGCTGCTCAAATGATGCCGTGATGGAGGTGACTGCCACCTGATGTGGTGTGCTATAC  
CACTGTCATCAATGGCTCTCAAGGAGGGATCTGGACAAAGCTTACGGTACATACCATGAAATGCTGGACCCG  
GGGATTTACCAAATGTTTACCTACAGCTCTATTATGCTGCGTTATGCAAGGCTCAAGCTATGGACAAAGCCA  
15 TGGAGGTACTTACCAAGCATGGTTAAGAATGGTGTGATGCCATTGCAAGGACGTATAATAGTATCGTCATGGTA  
TTGCTCTCAGGGCAGCGAAAGAGGCATTGGATTTCTCAAAGGATGACAGTGTGATGGTGTGAAACCAGATGTT  
GTTACTTATAACTCGCTCATGGATTATCTTGCAAGAACGGAAGATGACCGGAAGCTAGAAAGATGTTGCTATTCTA  
TGACCAAGAGGGCCTAAAGCCTGAAATTACTACCTATGGTACCCGCTCAGGGTATGCTACCAAAGGAGCCCT  
TGTGAGATGCATGGCTCTGGATTGATGGTACGAAACGTTACCCCTAATCATTATGTTTACGATTCTA  
20 ATATGTGCATACGCTAAACAAGGGAAAGTAGATCAGGCAATGCTGTGTTGCAAGGAGCTATGCTATGCTTA  
ATCCGGATACTGACCTATGGAACAGTTAGGCATACTTGCAAGTCAGGAGAGTAGAAGATGCTATGCTTA  
TTTGAGCAGATGATGAAAGACTAAGCCCTGGCAACATTGTTATACTCCCTAATTGATGCTCTGTATC  
TTTGACAAATGGGACAAGGCTAAAGAGTTAATTCTGAAATGTTGGATCAGGCACTGCTGGACACTATTTCT  
TTAATTCAATAATTGACAGTCATTGCAAGAAGGGAGGGTTATAGAATCTGAAAACCTTTGACCTGATGGTACG  
25 TATTGGTGTGAAGCCGATATCATTACGTACAGTACTCTCATCGATGGATATTGCTGGCAGGTAAAGATGGATGAA  
GCAACGAAGTTACTGCCAGCATGGCTCAGTTGGAAATGAAACCTGATTGTTACATATAATACCTTGTAAATG  
GCTACTGAAAATTAGCAGGATGAAAGATGCGTTAGTTCTTTAGGGAGATGGAGAGCAGTGGTGTAGCCTGA  
TATTATTACGTATAATATAATTCTGCAAGGTTATTCAAACCCAGAAGAACTGCTGTCGAAAGAAACTCTATGTC  
GGGATTACCGAAAGTGGACCGCTGAACTTAGCACATAACATAATCCTCATGGCTTGCAAAACAAATC  
30 TCACTGACGAGGCACCTCGAATGTTCAAGAACCTATGTTGACGGATTACAGCTGGAGACTAGGACTTTAACAT  
TATGATTGGTGCATTGCTTAAAGTTGGCAGAAATGATGAAGCCAAGGATTGTTGAGCTCTCGGCTAACGGT  
TTAGTGCCAGATGTTAGGACCTACAGTTAATGGCAGAAAATCTTATAGAGCAGGGTTGCTAGAAGAATTGGATG  
ATCTATTCTTCAATGGAGGAGAATGGCTGACTGCCAATCCCGCATGCTAAATTCCATTGTTAGGAAACTGTT  
ACAGAGGGGTGATATAACCAGGGCTGGCACTTACCTGTCATGATTGATGAGAAGCACTTCTCCCTGAAAGCATCC  
35 ACTGCTTCCCTGTTAGATCTTGTCTGGGGAAAATCAAGAATATCATAGTTGATTAGAGGAGGGATCT  
TCTCTTATGTTAAATAGCAGGTTCAAGAAAATCTTGTGATTGAGAATCTGGTGTCCATTCTTCTTAA  
ATTATTAAATCCTCCAGTGAATCTGTTGATTCCAAGCACCACATGATAGGTTCCAACATTCTTGAATCAGTAAA  
GTTCAAATGCTTAATGGATCAAATAAGGATTCTGACTGCATTGAGGAAATCCTTCAAAGTTGAAGAGATTC  
TCTTAAGCTGTCAGTGAATGTCAGTCGCTGACAAAGATGACAAGAAAACAAGGCCAGAACTGTCGAAAGTG  
40 GCTGCTTGTGACAATGGAAAATGCATGCTGTCTGCTGTTCAAGTGGAGACTTCTGACACAGTGTCCAGA  
GTTGGAGGAAATTAAAGAGACATTAAGGGAGATGGGAGGTCTGATAGTATTTGACGTTATGGTGGATTTC  
ATTCAACATTGGAGAATCTCATAAAGGATACATCCACTTCAGCTTGGACCGAAATGAAGGAACATTTGCAAAG  
TGCTGCTCCCTTGAATGTTGAAAATATTGAAATGCCATATTCTAAGCGATGATAACAAGACCCATTG

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CTTAATATGAGTAGAAAATTGAACCGAAACGCTCCCTGCTTCTTTGTTGGTGTCAATTATCAATACTATTGAGT  
 TATTATCAGCTCTTCATAACTTCAGAATTCTCTGTTGTTCCAGCTACATATCCGAAATCGTCTAAAGTCTC  
 TCAACAGAGTTACTCTGTGGTGATGGGGGGGGCGACCGTGGCCGAGGCGTGGAGTGCCTACCGCATCAGGGTGA  
 TCGGCCGCGCTGCTCCGCCCTGGTCCGAGGCTTGGCGCGAGCTGGCGGGAGGGAGACTGTGGTGAGATCGG  
 5 ATTCGCCGCTGGTGGTGTGCTACCATGGGGATTGCCGCAGGCGCTCAGATCGTTATGCCGGAGCGAAC  
 AAAAGTATGGCGTGGCGCGCGAGTGGACGGCCAGGCCTCGCGGAATGGGGCTGGGGACCGAGCCAGTCT  
 CGCTTGCCGGTAACCGGAACCGAGCTCAGCACTACATTGCAAAGATTGGCAACTCTGACAATTCCATGTTCT  
 ACAAGCTTGACGTCGAGGGATGGAGAACCTGCCACCGAATAGTAGCCCTGCTATCTATGTTGCGAACCATCAGAG  
 TTTTTGGATATCTATACCCCTCTAACTCTAGGAAGGTGTTCAAGTTATAAGCAAGACAAGTATATTATGTT  
 10 CGAATTATTTGA

*Rhpr2* is a rice homolog of the *Petunia Rf-PPR592* gene.

[0050] The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 22 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 23 as follows:

MARRAASRVAGAVGALRSEGSTQGRGGRTGGSGAEDARHVFDELLRRGRGASIYGLNCALADVARHSPAAVSRY  
 NRMARAGADEVTPLNLCTYGLIGSCCCCAGRDLGFAALGNVIKKGFRVDAIAFTPLLKGLCADKRTSDAMDIVLRR  
 MTQLGCIPNVFSYNNILLKGLCDENRQEAELELLQMMFDGGDCPPDVSVTTVINGFFKEGDLKDKEYHMLDR  
 20 GILPNVVITYSSIITAALCKAQAMDKAMEVLTSMVKNGVMPNCRTYNSIVHGYCSSGQPKEAIGFLKKMHSDGVEPDV  
 VTYNSLMDYLCKNGRCTEARKMFDSMTKRLKPEITTYGTLQGYATKGALVEMHGLLDLMVRNGIHPNHYVFSIL  
 ICAYAKQGKVDQAMLVFSKMRQQGLNPDTVTYGTIVIGILCKSGRVEDAMRYFEQMIIDERLSPGNIVYNSLIHSLCI  
 FDKWDKAKELILEMLDRGICLDTIFFNSTIDSHCKEGRVIESEKLFIDLVRIGVKPDIITYSTLIDGYCLAGKMD  
 ATKLLASMVSVGMKPDCVTYNTLINGYCKISRMEALVLFREMESSGVSPDIITYNIILQGLFQTRRTAAKELYV  
 25 GITESGTQLELSTYNIILHGLCKNNLTDEALRMFQNLCLTDLQLETRTFNIMIGALLKVRNDEAKDLFAALSANG  
 LVPDVRTYSLMAENLIEQGLLEELDDFLSMEENGCTANSRMLNSTVRKLLQRGDITRAGTYLFMIDEKHFSLEAS  
 TASLFLDLLSGGKYQEYHSCIRGGIFSLCVNSEVQENHLLDSESGVHFLKLLNPVNLDKAPSIGSKLLGISK  
 VQMLNGSNKDSDCISEEILSKVEEILSCQVIKSLDKDDKTTTRPELCPKWLALLTMENACLSAVSVEETSDTVSR  
 VGGNFKETLREMGGGLDSIFDVMVDFHSTLENLIKDTSTSALDRNEGTSLSQAAALLKCLKILENAIFLSDDNKTHL  
 30 LNMSRKLNPKRSLLSFVGIINTIELLSALSILQNSSVSSSTYPKSSKVSQQSYVVMAGGDRGRGVECHPHQGV  
 SAALLRPGPQALAASWRRRETVRSDFAAGGVATMGDPQALSDRLCGSATKVWRGGAETAEAFARNGAAGPSQS  
 RLPVTRNRAQHYIAKIWATLTISMFYKLDVEGMENLPPNSSPAIYVANHQSFLDIYTLLTLGRCFKFISKTSIFMF  
 RII

[0051] Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr3*, which has a nucleotide sequence of SEQ ID NO: 24, as follows:

40 ATGGCGCGCCGCGCCGCTTCCCGCGCTGTTGGCGCCCTCGCTCGACGGCTCGATCCAAGGGCGAGGAGGCCGCG  
 CGGGGGCAGTGGCGCCGAGGACGCACGCCACGTGTCAGCAATTGCTCCGGCGTGGCAGGGCGCCTCGATCTA

CGGCTTGAACCGCGCCCTCGCCGACGTCGCGCGTACAGCCCCGGCCCGTGTCCCGCTACAACCGCATGGCC  
 CGAGCTGGCGCCGACGAGGTAACCTCCGACTTGTGACCTACGGCATTCTCATCGGTTGCTGCTGCCGCGCGGGCC  
 GCTTGGACCTCGGTTCGGGCCCTGGGCAATGTCAATTAAAGAAGGGATTAGAGTGGAAAGCCATCACCTTCACCTCC  
 TCTGCTCAAGGGCCTCTGTGCCGACAAGAGGACGAGCGACGCAATGGACATAGTGCTCCGCAGAATGACCGAGCTC  
 5 GGTTGCATACCAAATGTCTCTCCTACAATAATCTTCTCAACGGGCTGTGTGATGAGAACAGAACAGCAAGAAGCTC  
 TCGAGTTGCTGCCACATGATGGCTGATGATCGAGGAGGAGGTAGGCCACCTGATGTGGTGTGCTATACCACTGTCAT  
 CAATGGCTTCTCAAAGAGGGGATTACAGACAAAGCTTACAGTACATACCATGAAATGCTGGACCGGGGGATTTA  
 CCTGATGTTGTGACCTACAGCTCTATTATTGCTGCCATTCAAGGGTCAAGCTATGGACAAGCCATGGAGTCATT  
 GCAAAGAAGGGAGGGTTATAGAACATCTGAAAAACTCTTGACCTGATGGTACGTATTGGTGTGAAGCCTGATATCAT  
 10 TACATACAGTACACTCATCGATGGATTGCTGGCAGGTAAGATGGATGAAGCAATGAAGTTACTTCTGGCATG  
 GTCTCAGTTGGGTTGAAACCTAATACTGTTACTTATAGCACTTGATTAATGGCTACTGCCAAAATTAGTAGGATGG  
 AACACGCGTTAGTTCTTTAAGGAGATGGAGAGCAGTGGTGTAGCCTGATATTATTACGTATAACATAATTCT  
 GCAAGGTTATTCAAACACAGAACAGAACGCTGCTGCAAAAGAACACTATGTCAGGATTACCGAACAGTGGAAACGCAG  
 ATTGAACCTAGCACATACACATAATCCTTCATGGACTTGCAAAACAAACACTCACTGATGATGCACTTCAGATGT  
 15 TTCAGAACCTATGTTGATGGATTGAAAGCTTGAGGCTAGGACTTCAACATTATGATTGATGCAATTGCTTAAAGT  
 TGGCAGAAATGATGAAGCCAAGGATTGTTGCTTCCTCGTCTAACGGTTAGTGGCAATTATTGGACGTAC  
 AGGGTATGGCTGAAATATTATAGGACAGGGTTGCTAGAAGAATTGGATCAACTCTTCTTCATGGAGGACA  
 ATGGCTGTACTGTTGACTCTGGCATGCTAAATTTCATTGTTAGGGAACTGTTGCAGAGAGGAGTAGTGGTGGTGGT  
 GAGTGGTGAATCTGCCACACCCACCAACTCTCAAAATTCTGACATGTGGGATCACTGTCAATCCCTCTCC  
 20 AAGACATGTGGATCACTGCAATCCCTCTCCAAACCAATTGTCAGACAGGTGCTTGGGTCAAGTTAAAGAAG  
 TTGGCAAAATGCTTCTGAAGAAAGGTTATTGTTGCTTCATCTCAGGAGATTCCAGATGATCCAGTGTCTCCAAC  
 AATTGAGGCGCTTATTTGCTCCATAGTAAAGCAAGTACACTTGCTGAGAACCCACCAACTTGACAAACACGGTTGTT  
 GTACCATCAAACAAACTTGGTTGATTCTGGGAAAGGTGAAAGTAATTACTGAAATGAGAACGGACTGGGG  
 CTGAAATCCGAGTCACTCAAAAGCAGATAAACCTAAGTACCTGTTGATGAGGAGCTTGTGCAAGCATATCAG  
 25 CCTTATCTTGGTTGATCGGCATGCTGGACGAGCACATCTGTTGTCGATCAACTGCTGACTGCTATATATGTGCTG  
 GTGCTGAATCGATCGATTGTCGTCGGAACTGAAGAACACCAACGGCACTGCTGCCGCTGGCTCTAGCCGCA  
 TCAGTTATAACCGTACAAACTCAGTGAATTGCTGGTTACATTGGTTATAATAAAAGGCCCTCGTTTTAGTT  
 CACCGCTGGGCTTCAGAATCTCAGGACCGGCCCTGCTCATGATCCTTACACCGTGTACCTGTAGAGTACTTCTCT  
 AAAAGAGAGTACCCTAGTGGAAACTGCAAAGTTGCACCATCTGCTTCATACGAAAGATATGCAGCAACTACTCGCT  
 30 TGCCTAATGGAGAACTGCCCTCATCTATTAGTCCTGGTGGCATTATATGCTCTGCCGTTCTATCTTGACCAAGT  
 ACCTACTGATAGGTACTCTAATAGGTTACACTACAATTAGGCCCTCGAGAGGCCGGAATAGTAATGTGCAACAA  
 TTAGGAATCACCAGAGCTGAAATTCCAATGCTTATGATTACTGAGGCTGCTGAGCACATGGACGTGAG  
 ATTACCGAAGACTGTCAGGCTCACTGGTATCCAGGTGGCTTCGAATTGTGGATTCCAATAGTTAACCTGGAG  
 TCTGTCATTGGTGGTGGTGAATCTGGTGCAGAGGTGAAGTTGCACGAAGCCCACAGAGCCTCTGCAAGGCTTCATCGCGCAAGCAGCA  
 35 GTGGAGATCCAGGGCATTCCGGATCAAGTGAAGGCCGACAGAGCCTCTGCAAGGCTTCATCGCGCAAGCAGCA  
 ACAGCAGGCAGGCAGCCCACTCCTCTCGCATGGCCCATTATTTTAG

*Rhpr3* is a rice homolog of the *Petunia Rf-PPR592* gene.

**[0052]** The nucleic acid molecule of the present invention which has the  
 40 nucleotide sequence of SEQ ID NO: 24 encodes a protein or polypeptide having a  
 deduced amino acid sequence corresponding to SEQ ID NO: 25 as follows:

MARRAASRAVGALRSDGSIQGRGGRAGGSGAEDARHVFDELLRRGRGASTYGLNRALADVARHSPAAVSRYNRMA  
 RAGADEVTPLCTYGLILIGCCCRAGRLDLGFAALGNVIKKGFRVEAITFTPLLKGLCADKRTSDAMDIVLRRMTEL  
 GCTPNVFSYNLLNGLCDENRSQEALELLHMMADDRGGSPDVSYTTVINGFFKEGDSDKAYSTYHEMLDRGIL  
 PDVVTYSSIIAALCKGQAMDKPWSHCKEGRVIESEKLFDLVRIGVKPDIITYSTLIDGYCLAGKMDEAMKLLSGM  
 5 VSVGLKPNTVTYSTLINGYCKISRMEDALVLFKEMESSGVSPDIITYNIILQGLFQTRRTAAKELYVRITESGTQ  
 IELSTYNIIILHGLCKNKLTDALQMFQNLCMDLKEARTFNIMIDALLKVRGRNDEAKDLFVAFSSNGLVPNYWTY  
 RLMAENIIQGQLEELDQLFLSMEDNGCTVDSGMLNFIVRELLQRGVVVVSGESATPPPTLKILTCGITVNPFS  
 KTCGITVNPFSKPIVQTGACGQVKVEGKVNASEERLIVVSSQEIPDDPVSPTIEALILLHSKASTLAENHQLTTRLV  
 VPSNKVGICLGEKKVITEMRRRTGAEIRVYSKADPKYLSFDEELVQHISLILVDRHAGRAHLLSHQLLTAIYVL  
 10 VLNRSIVVAEVKNNHGTAAWCWALAAISYNRTNFSDLLVSHWIFIKASVFSFTLGLQNLRTGPAHPYTVYPVEYFS  
 KREYPSGSSKVAPSASYERYAATTRLPNGELPSSISPGADYMSCRSYLDQVPTDRYSNRVTLQLGLSRAGNSNVQQ  
 LGITRAGNSNAYDYTEAAEQIHGREDYRRLSGLTGYPGGSSNCGFQIVNWSLSLVLVISGARVHLHEAHPGSSEI  
 VEIQGIPDQVKAAQSLLQGFIGASSNSRQAPQSSRMAHYF

15 **[0053]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr4*, which has a nucleotide sequence of SEQ ID NO: 26, as follows:

ATGCCGCTCGCCACGCTGCTCGGCCACCTCGCCGCCGGCGCTTCGGCTCGTGCAGGCCTCACCGGCCGCCGA  
 20 CCGCGGGCGGCCGCACCGACTCCTCACCTCCTCCGCACAGCGCCGCCCTCCCGACCTCCCGTCCACGGGCTCCTCTCGCCGCCCTC  
 CCTCGCGGGTGGTCGCGCGCCCACTCCGCGCCGCTCCCGTCCACGTCCCTGCCGCCGCGCCACTCCCGCAT  
 GCCTCCAAGGGGCTCTACCCCTCCCGTCCGAGCTCCACGCCGCTCATGCCGACATGCTCGTCC  
 CCATCCTCCGCGCTCTCCCGTCCCGTCCGCGTCCGATCCACGCCGCTCATGCCGACATGCTCGTCC  
 CGCCCTCGCCAGGGCATCCAGCCCTCAGGGCGTACGACCGTTCTCTGCCGGGGAGAGCCACCCGCCAC  
 25 CGCCCTCCACCTCCGTGAACGCCCTCTCGCCGCCCTCGCGGCCAAGCGGGTCGACCTGCCGAGAAGG  
 CGTCAGGAGCGCCTGCGCGCGTGTACCGGACATCTACACCTCAACACCGTCATCTCCGGCTCTGCAG  
 GATCGGCCAGCTCGCAAAGCGCGATGTCGCAAAGGACATCAAGGCATGGGTCTGGCTCCCTCTGTGGCACC  
 TACAATAGCCTCATCGATGGTACTGCAAGAAGGGTGGAGCTGGAACATGTACCATGTCGACATGCTTTGAAGG  
 AGATGGTCAAGCCGGATCTCACCGACTGCAAGTTACATTGGTGTGTTGATCAATGGGATTGCAAGAACCTCGAA  
 30 TACTGCGGCCAGTGGAGCTTCGAGGAGATGAAGCAGCAGGGATCGCTGCGAGTGTGACGTATAATTG  
 CTAATTTCAGGTCTCTGCAGTGAGGGTAAGGTGGAGGAAGGGGTGAAGCTGATGGAGGAGATGGAGGATTGGGGC  
 TGTCACCCAATGAAATCACCTTGGCTGTGTTCTGAAAGGGTTTGTAAAGAAGGAATGATGGCAGATGCCAATGA  
 TTGGATTGATGGTATGACAGAGAGGAATGTGGAACCTGATGTGGTTATTACAATATCTTGATCGATGTGATCGC  
 CGTCTGGAAAAATGGAGGATGCAATGGCGTGAAGGAGGAATGGCAAGAACAGGGATCAGTCCAATGTCACAA  
 35 CATATAATTGCTTGATAACAGGGTTAGCCGAGTGGGATTGGAGGAGTGCTCTGGCCTCTGGATGAGATGAA  
 GGAGAAAGGTATTGAAGCAGACGTCGTCACTTACAATGTGCTTATTGGTGTGCTGCAAAGGTGAGGTACGG  
 AAAGCTGTAAGCTCTGGATGAAATGTCGGAAGTTGGATTGGAACCAAACCATCTGACCTACAATACCATAATAC  
 AGGGGTTCTGTGATAAGGTAACATTAAGTCTGCCTATGAAATTAGAACCGAGATGGAAAAATGTCGAAACGGGC  
 AAATGTGGTTACGTACAATGTGTTCATCAAGTATTCTGCCAGATAGGGAAAGATGGATGAGCTAATGATCTACTC  
 40 AATGAGATGTTGGACAAATGTCTAGTCCAAACGGGATCACTTATGAAACGATAAAAGAGGGGATGATGGAAAAG  
 GCTATACACCAGATATTAGAGGGTGCAGTGTCTACAAGCTCTGAAAACCCAGCAGCATCCTGA

*Rhpr4* is a rice homolog of the *Petunia Rf-PPR592* gene.

**[0054]** The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 26 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 27 as follows:

5

MPLATLLGHLAAGRFGLVQALTGAATAAAAHRLLHLLLRTAPPPLPDLVSLARWSRAHFRAPLPLRLHGLLLARL  
 ASKGLYPLLRSELHVLAAARLHSASIPLRSPSASASASTPLIADMVLALARASQPLRAYDAFLLAGESHPRH  
 RPSTSSVNALLAGLVGAKRVDLAEKAFRSALRRRVSPDIYTFNTVISGLCRIGQLRKAGDVAKDIKAWGLAPSAT  
 YNSLIDGYCKKGGAGNMYHVDMLLKEMVEAGISPTAVTFGVILINGYCKNSNTAAAVRVFEEMKQQGIAASVVTYNS  
 10 LISGLCSEGKVVEGVKLMEEMEDLGLSPNEITFGCVLKGFCKKGMADANDWIDGMTERNVEPDVVIYNILIDVYR  
 RLGKMEDAMAVKEAMAKKGISPNTTYNCLITGFSRSGDWRSASGLLDEMKEKGIEADVVTYNVLIGALCCKGEVR  
 KAVKLLDEMSEVGLEPNHLYNTIIQGFCDKGNIKSAYEIRTRMEKCRKRANVTVNVFIKYFCQIGKMDEANDLL  
 NEMLDKCLVPNGITYETIKEGMMEKGYTPDIRGCTVSQASENPASS

15

**[0055]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr5*, which has a nucleotide sequence of SEQ ID NO: 28, as follows:

20

ATGGCTGATGATGGTCGCTGCCACCTGATGTGGTGTGTTAATACCATCATTGATGGTCTCTCAAAGAGGGTG  
 ATGTGGACAAAGCTTACATCACATACCATGAAATGCTGGACCGGAGGGTTCTCCAGATGCTGTGACTTACAACTC  
 TATCATTGCTGCCCTAACAGCAAGGCTCAAGCTATGGACAGGGCCATGGAGGTACTTACAGTGATGTTATGCCAAT  
 TGCTTCACATATAATAGTATTATGCATGGATATTGTTCTTCAGGACAGTCGGAAAAGGCTATTGGTATTTCAGAA  
 AGATGTGCAGTGATGGTATTGAACCAAGATGTTACTTATAACTCGTGATGGACTATCTCTGCAAGAACGGAAA  
 ATGCACAGAACGCCAGAAAGATTTGATTCTATGGTCAAGAGGGCTCAAGCCTGATATTACTACCTATGGTACC  
 25 CTGCTTCATGGGTATGCTTCAAAGGAGCTTGTGAGATGCATGATCTCTAGCTTGATGGTACAAAATGGCA  
 TGCAACTTGATCATCATGCTTCAACATATTAATATGTGCATACACTAAACAAGAAAAGTAGACGAGGTCTGCT  
 TGTATTCAAGCAAAATGAGGCAGCAAGGATTGACTCCGAACGCAGTGAACTATAGAACAGTGATAGATGGACTTTGC  
 AAGTTAGGTAGACTAGATGATGCTATGCTTAATTGAGCAGATGATTGATAAAGGACTGACACCTAACGTTGTTG  
 TTTATACCTCCCTAACATTGCTCTGTACCTATGACAAATGGAGAAGGCCGAGGAGTTAATTGGAAATATT  
 30 GGATCAAGGTATCAATCCAAACATTGTTTTTAATACAATATTGGACAGTCTTGCAAGAACAGGGAGGTTATA  
 GAATCTAAAAAAACTCTTGACCTGTTGGACATATTGGTGTGAATCCTGATGTCATTACATACAGTACACTCATCG  
 ATGGATATTGCTTAGCTGGTAAGATGGATGGAGCAATGAAGTTACTCACTGGCATGGCTCAGTTGGTTGAAACC  
 TGATAGTGTACATATAGCACTTGTAAATGGTTACTGTAATAGAACAGTGATAGATGGACTTTGCTTTC  
 AAGGAGATGGAAAGCAATGGTGTAACTGATATTACATATAACATAATTCTGCATGGTTATTCGCACCA  
 35 GAAGAACTGCTGCTGCAAAAGAACTATATGCCAGGATTACCGAAAGTGGAACGCAGCTGAACCTAGCACATACAA  
 CATAATCCTCATGGACTTGCAAAACAAACTCACTGATGATGCACTTCGGATGTTCAAGAACCTATGTTGA

*Rhpr5* is a rice homolog of the *Petunia Rf-PPR592* gene.

[0056] The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 28 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 29 as follows:

5 MADDGRCPDVSYNTIIDGLFKEGDVKAYITYHEMLDRRVSPDAVTYNSIIAALSKAQAMDRAVELTVMVMPN  
CFTYNSIMHGYCSSGQSEKAIGIFRKMCSDGIEPDVVVTYNSLMDYLCKNGKCTEARKIFDSMVKRLKPDITTYGT  
LLHGYASKGALVEMHDLLALMVQNGMQLDHHVFNILICAYTKQEKFVDEVVLVFSKMRQQGLTPNAVNYRTVIDGLC  
KLGRLLDAMLNFEQMIKGLTPNVVYTSЛИHALCTYDKWEKAELIFEILDQGINPNIVFFNTILDSCKEGRVI  
ESKKLFDLLGHIGVNPDVITYSTLIDGYCLAGKMDGAMKLLTGMVSVGLKPDSTYSTLINGYCKINRMEDALALF  
10 KEMESNGVNPDIITYNIILHGLFRTRRTAAKELYARITESGTQLELSTYNIILMDFAKTNLSMMHFGCFRTYV

[0057] Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr6*, which has a nucleotide sequence of SEQ ID NO: 30, as follows:

15 ATGGCGCGCCCGCGCCGCTTCCCGCGCTGGCTGGAGGGCTCGATCCAAGGGCGAGGGGGCCGCGCGGGGGCA  
ATGGCGCCGAGGACGCACGCCACGTGTTGACCGAATTGCTCGGCGTGGCAAGGGCGCCACGATCTACGGCTTGAA  
CCCGCGCCCTCGACGACGTCGCGCGTACAGCCCCGCGGCCCGTGTCCCGTACAACCGCATGGCCGAGGCCGCG  
GCCGACGAGGTAACTCCAACTTGTACACCTACAGCGTTCTCATCGGTTGCTGCCGGCGGGCCGCTTGGACC  
20 TCGGTTCGGGCCTTGGCAATGTCATTAAGAAGGGATTAGAGTGGAAAGCCATCACCTTCACTCCTGCTCAA  
GGGCCTCTGTGCCGACAAGAGGACGAGCGACGCAATGGACATAGTGCTCTGCAGAATGACCCAGCTCGGCTGCATA  
CCAAATGTCCTCCTGCCACCATTCCTCAAGGGTCTGTGTGATGAGAACAGAACAGCAAGAACGCTCTCGAGCTGC  
TCCAAATGATGCCGTGATGATGGAGGGTGAECTGCCACCTGATGTGGTGTGACAACACCGTCATCAATGGCTTCTT  
CAAAGAGGGGATCCGGACAAAGCTACGCTACATACCATGAAATGTTGACCAGGGATTTGCCAGATGTTGTG  
25 ACTTACAGCTCTATTATCGCTGCCTATGCAAGGCTCAAGCTATGGACAAGGCCATGGAGGTACTAACACCATGG  
TTAAGAATGGTGTATGCCCTAATTGCAAGGACATATAATAGTATTGTCACGGATATTGCTCTTCAGGGCAGTTGAC  
AGAGGCTATTGGATTTCTAAATGATGTCAGTGATGGTGTGCAACCGAGATGTTTACTTGTAACTTGCTGATG  
GATTATCTTGCAAGAACAGAACGATGCACGGAAGCTAGAAAGATTTCAATTCTATGACCAAGTGTGGCTAAAGC  
CTGATATTACTACCTATTGTAACCTGCTTCAGGGTATGTCACCAAAGGAGGCCCTGTTGAGATGCACTGATCTCTT  
30 GGATTTGATGGTATGGAACGGTATCCAACCTAATCATCATGTATTCAACATTCTAATATGTCATACGCTAAACAA  
GAAAAAGTAGATGAGGCGATGCTGTATTCAAGGAAATGAGGCAAGGATTGAGTCCGAATGCACTGAGTGAAC  
GAACAGTCATAGATGTAACGTCAGGCAAGGATGAGTCAACGATGCACTGCTTACCTAAAGCAGATGATCAATG  
AGGACTAACCCCTGACATCATTGATATACCCCCCTAATTCAATGGTTTGACCTGTGACAAATGGGAGAAGGCT  
GAGGAGTTAATTTTAA

*Rhpr6* is a rice homolog of the *Petunia Rf-PPR592* gene.

[0058] The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 30 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 31 as follows:

MARRAASRAVGSEGSIQGRGGRAGGNGAEDARHVFDELLRRGKGATIYGLNRALDDVARHSPAAVSRYNRMARAG  
 ADEVTPNLYTYSVLIGCCCRAGRDLGFAALGNVIKKGFRVEAITFTPLLKGLCADKRTSDAMDIVLCRMTQLGCIP  
 PNFSCTILLKGLCDENRSQEALELLQMPDDGGDCPPDVLYNTVINGFFKEGDPDKAYATYHEMFDQGILPDVV  
 5 TYSSIIAALCKAQAMDKAMEVLNTMVNGVMPNCRTYNSIVHGYCSSGQLTEAIGFLKMMCSDGVEPDVVTCNLLM  
 DYLCNRRCTEARKIFNSMTKGLKPDITTYCTLLQGYATKGALVEMHDLDDLMVWNGIQPNHHVFNILICAYAKQ  
 EKVDEAMLVFSKMRQQGLSPNAVNRTVIDVLCKLGRVYDAVTLKQMINEGLTPDIIIVYTPLIHGFCTCDKWEKA  
 EELIF

10 **[0059]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr7*, which has a nucleotide sequence of SEQ ID NO: 32, as follows:

ATGGCACGCCCGTCGCTGCCCGCGCCCGCGCCGGCGTCCCGCGCTCGGAGGGTACGATCCAAGACC  
 15 GAGCACCGCGTTGGGAGCGGTGGCGCCGAGGACGACTCGACGTGTTCGACGAATTGCTCCGGCGAGGCATCGCCGC  
 TCCGATCCGCAGCTTGAACGGCGCTCTCGCCGACGTCGCGCGACAACCCCGGGCGCTGTGTCCCGCTTCAAC  
 CGCATGGCACGAGCTGGTGCAGCATGGTAACTCCCACCGTGCACACCTATGGCATCCTCATCGGCTGCTGCTGCA  
 GTGCGGGCCGCTTAGACCTCGGTTTCGGCCCTGGGCATGCGTTAGAAGGGATTCAAGAGTGGAAACCCATCAT  
 CTTAATCCTCTGCTCAAGGGCCTGTGCAGACAAGAGGACGGACGACGAATGGACATAGTGTCCGTGGAATG  
 20 ACCGAGCTCAGCTCGTGCCTGCCAATGCTTCTCCCACACCATTATTCTCAAGGGACTCTGTCATGAGAACAGAACCC  
 AAGAACGCTCTCGAGCTGCTCCACATGATGGCTGATGGAGGAGGCTGTTACCTAATGTTGTCATACAGCAC  
 CGTCATCGATGCCCTTGAAAGGAGGGATCCGGACAAAGCCTACGCTACATACCGTAAATGCTTGACCGGAGG  
 ATTTTGCCAAATGTTGTGATTACAGCTCATTATTGCTGCCCTATGCAAGGGTCAAGCAATGGACAAGGCCATGG  
 AGGTACACGATAGGATGGTTAGAATGGAGTTACACCAATTGCTCACGTATACTACTCTTGTGATGGATTITG  
 25 CTCTTCAGGGCAGTTGACAGAGGCTATTAATTCAGAAAAGATGTCAGCAATGGTGTGAACCAAATGTTGTT  
 ACTTATAGCTCGTTATGGACTATCTCTGCAAGAACGGAAAGATGCACAGAACGCTAGAAAGATTGATTCTATGG  
 TCAAGAGGGCCTAAAGCCTGATATTACTACCTACAGTAGCTTACTTCATGGGTATGCTATCGAAGGAGCTTGT  
 TGAGATGCATGGCTCTTGATTGATGGTACAAAGTGATATGCAACCCGATCATTATGCTTCAACACACTAATA  
 TATGCATCCGCCAAGCAAGGAAAAGTAGATGAGGCCATGCTGTATTAGCAAAATGAGGCAGCAAGGATTGAAAC  
 30 CTAATTGTTACGTATAGCACTTGTATTAATGGTACTGTAAAATTACTAGGATGGAGAATGCTTACTGCAAGGTT  
 CCAAGAGATGGTGAGCAATGGTGTAGCTTAATTTATCACATATAACATAATGCTGCAAGGTTATTGCTACA  
 GGAAGAACTGCTACTGCAAAAGAATTCTATGTACAGATTATCAAAAGTGGCAAAAAAGATCTTATAGAACAGGGT  
 TGCTAGAAGAATTGGATGATCTATTCTTCAATGGAGGACAATGACTGTAGTACTGTGTCGACTCCTGCATGCTA  
 A

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*Rhpr7* is a rice homolog of the *Petunia Rf-PPR592* gene.

**[0060]** The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 32 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 33 as follows:

MARRVAARARARAGGVPRSEGTIQDRARVGSGGAEDALDVDELLRRGIGAPIRSNLNGALADVARDNPAAAVSRFN  
 RMARAGASMVTPTVHTYGILIGCCSAGRDLGFAALGHVVKKGFRVEPIIFNPLLKGLCADKRTDDAMDIVLRGM  
 TELSCVPNVFSHTIILKGLCHENRSQEALLEHMMADDGGGCLPNVSYSTVIDGLLKGGDPDKAYATYREMLDRR  
 ILPNVVIYSSIIAALCKGQAMDKAMEVHDRVVKNGVTPNCFTYTSVLHGFCSSGQLTEAIKFLEKMCNSNGVEPNVV  
 5 TYSSFMDYLCKNGRCTEARKIFDSMVKRLKPDITTYSSLLHGYAIEGALVEMHGLFDLMVQSDMQPDHYVFNTLI  
 YASAKQGVDEAMLVFSKMRQQGLKPNCVTYSTLINGYCKITRMENALALFQEMVSNGVSPNFITYNIMLQGLFRT  
 GRTATAKEFYVQI IKSGKKDLIEQGLLEELDDLFLSMEDNDNSTVSTPAC

[0061] Another suitable nucleic acid molecule in accordance with the present  
 10 invention is isolated from rice and identified herein as *Rhpr8*, which has a nucleotide  
 sequence of SEQ ID NO: 34, as follows:

ATGGCGCGCCCGCGCCGCTTCCCGCGCTGCTGGCGCCCTCGCTCGAGGGCTCGATCCAAGGGCGAGGGGGCCGCG  
 CGGGGGCAGTGGCGGTGGCGCGGAGGACGCACGCCACGTGTCGACGAATTGCTCCGTCGTGGCATACCAAGATGT  
 15 CTTCTCCTACAAATTCTCTCAACGGGCTGTGTATGAGAACAGAACAGCCAAGAACGCTCTCGAGTTACTGCACATA  
 ATGGCTGATGATGGAGGTGACTGCCACCTGATGTGGTGTGTCGACAGCACCGTCATCAATGGCTCTTCAAGGAGG  
 GGGATCTGGACAAATGCTTGACCAGAGGATTTCGCCAAATGTTGTGACCTACAACCTCTATTATTGCTGCGCTATG  
 CAAGGCTCAAACGTGGACAAGGCCATGGAGGTACTTACCAACATGGTAAAGAGTGGTGTATGCCGATTGCGATG  
 ACATATAATAGTATTGTGCATGGTTTGTCTTCAGGGCAGCCGAAAGAGGCTATTGTATTTCAAAAAGATGC  
 20 GCAGTGATGGTGTGAAACCAGATGTTTACTTATAACTCGCTCATGGATTATCTTGCAAGAACGGAAGATGCAC  
 GGAAGCAAGAAAGATTTTGATTCTATGACCAAGAGGGCCCTAAAGCCTGATATTACTACCTATGGTACCCCTGCTT  
 CAGGGTATGCTACAAAGGAGCCCTGTTGAGATGCATGGCTCTGGATTTGATGGTACGAAACGGTATCCACC  
 CTAATCATTATGTTTACGCATTCTAGTATGTGCATACGCTAACAAAGAGAAAGTAGAAGAGGCAATGCTTGTATT  
 CAGCAAAATGAGGCAGCAAGGATTGAATCCGAATGCAGTGACCTATGGAACAGTTATAGATGTACTTGCAAGTCA  
 25 GGTAGAGTAGAAGATGCTATGCTTATTTGAGCAGATGATCGATGAAGGACTAACGACTGACAGCATTGTTATA  
 ACTCCCTAATTCTAGTCTCTGTATCTTGACAAATGGAGAAGGCTGAAGAGTTATTCCTGAAATGTTGGATCG  
 AGGCATCTGTCTTAGCACTATTTCTTAATTCAATAATTGACAGTCATTGCAAAAGAAGGGAGGGTTATAGAATCT  
 GGAAACTCTTGACTTGATGGTACGAATTGGTGTGAAGCCGATATCATTACCTTGGCAGGTTTGGGAGCG  
 CAAGGCGCAGACTACTCACTGTTGCTAACACATCTACCCATCTTCAACATGTCGAACACTGGAGACAAGGAGAA  
 30 GGAGACTCCCGTCAACACCAACGGAGGAATACTGCCTCAAACCTCCAGCGGAGGACCATTCTGGGCACATACAAC  
 ATAATCCTTCATGGACTTTGCAAAACAAACTCACTGATGATGCACTTCGAATGTTCAAGAACCTATGTTGATGG  
 ATTTGAAGCTTGAGGCTAGGACTTTCAACATTATGATTGATGTCATTGCTAAAGTGGCAGAAATGATGAAGCCAA  
 GGATTGTTGTTGCTTCTGCTAACGGTTAGTGGCAATTATTGGACGTACAGATTGATGGCTGAAATATT  
 ATAGGACAGGGTTGCTAGAAGAATTGGATCAACTCTTCAATGGAGGACAATGGCTGTACTGTTGACTCTG  
 35 GCATGCTAAATTCTATTGTTAGGAACTGTTGCAAGGAGGTGAGATAACCAGGGCTGGCACTTACCTTCCATGAT  
 TGATGAGAAGCACTTTCCCTCGAACGCATCCACTGCTTCTGTTATAGATCTTGCTGGGGAAAATATCAA  
 GAATATCATATATTCTCCCTGAAAATACAAGTCCTTATAGAATCTTGAGCTGCTGA

*Rhpr8* is a rice homolog of the *Petunia Rf-PPR592* gene.

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**[0062]** The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 34 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 35 as follows:

5 MARRAASRAAGALRSEGSIQGRGGRAGGSGGAEDARHVFDELLRRGIPDVFSYNILLNLCDENRSQEALELLHI  
MADDGGDCPPDVVSYSTVINGFFKEGDLKDMLDQRISPNVVTYNSIIAALCKAQTVDKAMEVLTVMKSGVMPDCM  
TYNSIVHGFCSGQPKEAIVFLKKMRSRGVEPDVVTYNSLMDYLCKNGRCTEARKIFDSMTKRGKPDITTYGTLL  
QGYATKGALVEMHGLLDLMVRNGIHPNHYVFSILVCAYAKQEKEEAMLVFSKMRQQGLNPNAVTYGTVIDVLCKS  
GRVEDAMLYFEQMIDEGLRPDSIVYNSLIHSLCFDKWEKAELFLEMLRGICLSTIFFNSIIDSHCKEGRVIES  
10 GKLFDLMVRIGVKPDIITLGRFLGSARRDYSLFVNIFYFIFTNMSNTGDKEKETPVNNTNGNTASNSSGGPFLGTYN  
IILHGLCKNKLTDALRMFQNLCMLDKLEARTFNIMIDALIKVGRNDEAKDLFVAFSSNGLVPNYWTYRLMAENI  
IGQGLLEELDQLFLSMEDNGCTVDSGMLNFIVRELLQRGEITRAGTYLSMIDEKHFSLEASTASLFIDLLSGGKYQ  
EYHIFLPEKYKSFIESLSC

15 **[0063]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr9*, which has a nucleotide sequence of SEQ ID NO: 36, as follows:

ATGGCGAGAAGAGGACGACGATACTGTAGAGCAGAGGAACAGAGGAGCGGGCACTGGTGCTCCGGTGGCTGGGA  
20 GGTGGCGACGTGGCGGCCAATGTGTTCCCGAGCGCCGCGCTGGAGAGCCCCGAGCTGCGACGGCATCACGCCGA  
CTACCGGCCGTGGCGGCCACATGGAGGCAAAGCCGGTCACTTCGCGTCAGGCGTGCCTCCGGCGGCCAG  
CTGCAGCAGCAGCTCGTCCGGCCACCCAACTCTGGCCGATTGGCCGATCTCAGCCTCCGGAGCGGAGACCGA  
TCTGGGCCGTCCATCCGCGCCGCCAGCCAATCGACGGTGGGTGATTACTGTACTGCCAGGTGGTACCCCTCC  
GCCGCCGGCGGCCGGCGGCCGGCGCAGGCATGGCGCCGGTGTACCACCCCTAACCGCGCCGCCACCCCGCC  
25 CGCGCGGGCGGCCTCCCCAGCGCGCAGGGTGGTACGACCCAAGACCTAGGGCGCGGGGGCAGTGGCACCGAGG  
GCGCACGCCACGTGCTCGACGAATTGGCCTACGGGCTGGGCGCTCGATCTACAGCTCAACCGCACCCCTCAC  
CGACGTCGCGCGTGCAGCCCCAGCCGAGCAGTTCGCTCTTCACCGCATGGCCCGAGCCGGCGACGAGGTA  
ACTCCGACTTGTGCACCTACAGCATTCTCATCGTTGCTGCCGCCGGCGCTTGGACCTCGGTTCGCGG  
CCTTGGCAATGTCATTAAGAAGGGATTAGAGTGGAAAGCCATCACCTCGCTCCTCGCTCAAGGGCTCTGTC  
30 CGACAAGAGGACGAGCGACGCAATGGACATAGTGTCCGCAGAATGACCGAGCTCAGCTGCATGCCAGATGTTTC  
TCCTGCACCATTCTCAAGGGTCTGTGATGAGAACAGAACGCAAGAGCTCTCGAGCTGCTGCACATGATGG  
CTGATGATCGAGGAGGAGGTAGCCCACCTGATGGTGTGCTGATACCAACTGTCATCAATGGCTTCTCAAAGAGGG  
GGATTCAAGACAAAGCTTACAGTACATACCATGAAATGCTGATGGAGGATTTCAACAAATGTTGACCTACAGC  
TCTATTATTGCTGCGTTATGCAAGGCTCAAGCTATGGACAAAGCCATGGAGGTACTTAACACCATGGTTAAGAATG  
35 GTGTCATGCCGATTGCAATGACATATAATAGTATTCTGCATGGATATTGCTTCAAGGGCAGCCAAAGAGGCTAT  
TGGAAACTCAAAAGATGCGCAGTGTGGCGTCAACCAAATGTTACTTATAGATCACTGATGAATTATCTT  
TGCAAGAACGAAAGATGCACCGAAGCTAGAAAGATTTGCTATGACCAAGAGGGGCCTGAGATGCATGCTCTTGGATTGAT  
CTACCTATCGTACCCCTGCTCAGGGGTATGCTACCAAAAGGAGCCCTGAGATGCATGCTCTTGGATTGATATTG  
GGATCCTGAGTTCTACAAGTATTGGAGAAGTGA

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*Rhpr9* is a rice homolog of the *Petunia Rf-PPR592* gene.

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**[0064]** The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 36 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 37 as follows:

5 MARRGRRYCRAEGTEERGTGAPVAGRWRRRPNVFPSSALESPELRRHADYRPWAHMEAKPVYFASRRASGRPE  
LQQQLVPRPTIPIWADWADSLPERRPIWAVHPRRPANRTVGVLILYCQVGDPFFFFAAAAAGMARRVTLTRARTRA  
RGGVPSAQGGTTQDLGRAGGSCTEGARHVLDELPLRGWGASIYSFNRTLTDVARDSPAASLNFNRMARAGADEV  
TPDLCTYSILIGCCCRAGRDLGFAALGNVIKKGFRVEAITFAPLLKGLCADKRTSDAMDIVLRRMTELSCMPDVF  
SCTILLKGLCDENRSQEAELELLHMMADDRGGSPPDVVSYTTVINGFFKEGDSDKAYSTYHEMLDRRISPNVVTYS  
10 SIIAALCKAQAMDKAMEVLNTMVKNGVMPDCMTYNSILHGYCSSGQPKEAIGTLKKMRSDGVEPNVVTYRSLMNYL  
CKNGRCTEARKIFDSMTKRGLEPDIATYRTLLQGYATKGALVEMHALLDLMDPEFYKYLEK

**[0065]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr10*, which has a nucleotide sequence of SEQ ID NO: 38, as follows:

ATGCCCTTCCGACCGCGCCTCCGCTCCCCCTCCCTCCTCCTGCCTCACCTCCGGGCCGCCCTCCCTCCC  
CCCGCCCGCCCGTGCCCGCTGGAGACCGCTCTCGTATTATCCCTCGGGCGGCCGCCGGAGGTGACGGA  
GTCCGAGGAGGACGCGGCGCTTGGCAGGGACACCCGAGCCCTCCCTCCATGGCGGGATTGCACGGGAGCG  
20 CCTAGGGTTGGCTGCAATGGCGGGGGCTGCCGATGACGAGGAGGTGAGAGGAAGGCCCGCCTGCGCGCGA  
TCAAGCTCTGCATGAGCTCTCGCGGAGAGGAGGTGGCGCGATGCGGGCAGCCTGGCGCAGCTGGTACTGA  
GCAAGGTGAGCATGCTATGAATTTCCTCATTCTGATTATCAACTCTACTCATGTGGTATCTGAATAACTATGGT  
ATTGGTGTGAGGAGGCGTAGGAATGGCATCGGTAGTTGAECTCTGATCGATATGAATGTGTGACACAGGATAT  
ATTGTTTCCAGAGGCATTATCAATTGATCATTACCATATAAAACAGTAAGAAAAGGGTCGAAAGCAATGCAT  
25 ACATAGTTGATTTGGTAGTATTACTGTAATTGTTTACTAGAAGGTCTTGCAAGTATGACAAACTA  
GTAACATAAAATTGTCGCTTAACTTATTGCGCTTCTGCTGTAGGATCTGGTCTGCAGCTGCTCTGTG  
ACATCTTATGAAACAGATTCAAGAGAGTGTGATTCCAACGGTTGTGTATGGGATGCTTAGCGAACAGTTATGCTAG  
AGCTCAGATGGTCATGATGCCCTTACGTTCTAGTAAAATGAGCAGCCTAACATGCAAATCTGGTGTTCACC  
TATGACAGTTATTGCAACGGCTTAAGGATGACAGACGTGGCATTGGAGCTTTGAAGAAATGGAGTCTTGTGGT  
30 TCTCTCCAGTGAATATTCGCATAGTATTATTAAATGGCCTCTGTAAGCAAGATAAGGTTGGAGAACGTTATC  
TTTCCTTCAGGAAGCTAGGAAGGAGGGAAAGTTAAACCTTGGGAATGACCTTAAACATTCTATGCTGCATTG  
TGTAATTGGGGTTGTTAGTCTGCAAAATCATTGCTGATGCTGAAATATGGATTAGTCCCTGACAGGT  
ATACCTTTCTACCCCTAACACGGCTATGTAAGTAGGTTCAATGGAGGAAGCATTGGATCTTCTGAGAGAGT  
GACAAAAGAAGGAATGGAACCTTGAGATTGTGACCTACAATAGCCTTATCAATGGTACCGATTGCTTGGTTAAC  
35 AAAGAAAATCCCTAAATCATCCAGATGATGAGAGGCCAAGGTGTTGAAACCTGATCTGTTACATATACTACTTA  
TTGCTGGTCACTGCGAAAGTGGTATGTTGAAGAAGGAATGAAGGTAAAGGAAGGATGTCCTAGACCAAGGTTGCA  
GTTGAATATTGTCACATATAGTGTCTTCTCAATGCTCTTCAAAAAGGCATGTTCTGCGAAATTGACAACCTA  
CTCGCGAGATCTACAATATTGGTTGGATATGGATGTTATCGCATATTCCATCCTTATCCATGGTATTGCAACC  
TAGGGGAAATTGAAAAGGCCTTCAAGTATGCAATGCAATGTCAGTTCTCAGAGGGTAATGCCAACATCACTGAA  
40 CCATTTTCTATTCTTAGGACTTGCAGAAAGGATTGTTAGTGAAGCAAGGTGGTATTGGAAAATGTAGCT  
AGAAAATATGCCAACTGATGTTAGTGTCTATAATGCGTATTGATGGTTATGCAAAACTGGTATATTGTA

ATGCTGTTGTTGTATGATCAGATCACTGTAGCTGGTATGCACCCAACCATTGTCACATGCAATTCTCTTCTATA  
 TGGTATTGAAAATTGGGGATCTGCAACTTGCCGAGAGCTATTTAGGGCTATTCACTAAGTGGACTTCTACCA  
 ACAGCAGTGACATACACTACCTGATGGATGCACTCTGAAAGCTGGAGAAGTTAATACCATGCTAAGTCTTTG  
 ATGAAATGGTGCAAAGAGGGATCAAGGCAAATGCAGTAACCTACAGTGTATTGTTAAAGGGCTTGTAAGCAGCT  
 5 CAGATTGAGGCTATCAATGTTCTCAAAGATATGGATAGCAAAGGTATTAATGCTGACCCGATAACCTACAAT  
 ACCCTTATACAAGGTTCTGTGAATCAGAAAACGTTAGATGGCTTCCACATACATGACATCATGTTATGCCGTG  
 GCCTGTCGCCACACCTGTTACTTATAACTTGCTTATTAAATGTGCTGTGTTGAAGGGAAAAGTTATTCAAGCAGA  
 AATACTTTGGAGTCCCTCAGAGAAAATGGCATTAAGTTGAGAAAATTGCGTACACAAACACTTATCAAAGCTCAG  
 TGCGCAAAGGAATGCCTATCAATGCTGTTGTTAGTTGGTAAGCTCTAGATGCAGGATTGAAGCTTCTATTG  
 10 AAGATTTCAGTGCAGCAATCAATGACTTGCAAAAGACAATTGCAAAAGCCTTATGTTGTCCCGATTAT  
 GCTATCTGTTGGTATTACCCAGATACTCAAATATATTGTGCTAGGCAGAGCTCTGCAGAAAAATAGTGAGCTT  
 GTCTATCTACCCATATTAAATGCACTGCTGTTAAACTGGTATTAA

*Rhpr10* is a rice homolog of the *Petunia Rf-PPR592* gene.

15 [0066] The nucleic acid molecule of the present invention which has the  
 nucleotide sequence of SEQ ID NO: 38 encodes a protein or polypeptide having a  
 deduced amino acid sequence corresponding to SEQ ID NO: 39 as follows:

20 MPFRPRLPLPLLLLLLPHLRRRSSPRPPVPAWRPLSYYPSAAAAAAEVTESEEDAAAVGRDTRAPPSIGGIARGA  
 PRVGNGGAADDEEVERKARAVARIKLCHELLRERRWRAMRAALAQLVTEQGSGSAALCDILWNRFRECDNSGC  
 VWDALANSYARAQMVKHDALYVLSKMSSLNMQISVFTYDSLLHGLRMTDVALELFEEEMESCGVSPSEYSHSIIINGL  
 CKQDKVGEALSLFLQEARKEGKFKPLGTMFNILMSALCNWGFVQSAKSFLCLMLKYGLVPDRYTFSTLIHGLCKVGS  
 MEEALDLFERVTKEGMELEIVTYNSLINGYRLLGLTKEIPKIIQMMRGQGVEPDLVTYTIAGHCESGDVEEGMK  
 VRKDVLQGLQLNIVTYSVLNALFKKGMFCEIDNLLGETYNIGLMDVIAYSILIHGYCKLGEIEKALQVCNAMC  
 25 SSQRVMPTSLNHSILLGLCKKGLLVEARWYLENVARKYQPTDVVFYNVIDGYAKLGDIVNAVRLYDQITVAGMH  
 PTIVTCNSLLGYCKIGDLQLAESYFRAIQLSGLLPTAVTYTTLMDALSEAGEVNTMLSLFDEMVAKRIKANAVTY  
 SVIVKGLCKQLRFDEAINVLKDMDSKGINADPITYNTLIQGFCESENVQMAFHIDIMLCRGLVPTVYNLLINV  
 LCLKGKVIAQELLESRLRENGIKLRKFAYTTLIKAQCAKGMPINAVLLVGKLLDAGEASIEDFSAAINRLCKRQF  
 AKEAFMFVPIMLSVGIVYPDTQIYCVLGRALQKNSELVYLPILNALAVKTG  
 30

[0067] Another suitable nucleic acid molecule in accordance with the present  
 invention is isolated from *Arabidopsis thaliana*, which has a nucleotide sequence of  
 SEQ ID NO: 40, as follows:

35 ATGAAGGCTTGAGATTGATTCAAGCCTCATCTCTGAAGACAGGTAGTCTTAGAACTGATTGCTCTGTACCAATT  
 CGAGTTCTTCTAGCTGCGAACGAGACTTTCAAGTATTAGCAATGGAAATGCTGTTCAAGAGAGATTGAG  
 AAGTGGTATTGTTGATATTAAGAAAGATGATGCTATTGCTCTGTTCCAAGAAATGATTAGGTCTCGTCCTCTCCT  
 AGCTTGTGTTAGTCAAGTATTCTTAGTGCCTATTGCCAGAACAAAACAGTTCAATCTCGTGTAGATTCTGCA  
 AGCAACTGGAATTGAATGGGATTGCTCATAACATCTACACTTGAATATCATGATCAACTGCTTTGCCGGTGTG  
 40 TAAAACTTGTTTGCTTATTCTGTTGGAAAAGTAATGAAGCTTGGGTATGAGCCTGACACAACCACGTTAAC

ACTCTGATCAAAGGACTTTCTTGAGGGTAAAGTGTCTGAAGCTGTGGTTAGTCGATAGGATGGTGGAAAACG  
 GATGTCACCTGATGTGGTTACTTATAATTGATTGAAATGGGATATGTAGATCAGGAGATACTTCTTGGCCTT  
 GGAGTTGCTCAGAAAGATGGAAGAAGAAATGTTAAGGCTGATGTGTTACTTACAGTACAATCATTGATAGTCTT  
 TGTAGAGATGGTGCATAGACGCTGCAATTAGCCTTTCAAGGAAATGGAGACGAAAGGGATTAAATCTAGTGTG  
 5 TTACGTATAATTCTCTTGAGAGGCTTTGTAAGCCGTAAGGATGATGGGCACTGTTGTTGAAGGATAT  
 GGTGAGTAGGAAATGTCCTAATGTCATCACTTCAATGTTACTTGTGTTGTCAAAGAAGGGAGCTT  
 CAGGAGGCTAATGAATTGTACAAAGAGATGATCACAAGAGGTATATCACCTAATATTATTACTTATAATACCTG  
 TGGATGGTATTGTATGAGACGGCTTACTGAGGCCAACAAATATGTTGATCTTGTGTTAGAATAAGTGCAG  
 TCCTGATATCGTACTTACAAGTCTCATCAAAGGATATTGTATGGTAAAAGAGTTGACGATGGTATGAAGGTC  
 10 TTCCGCAATATTCTAAGAGAGGCTTGGTGCACAGTTACTTATAGCATTCTGTCCAAGGGTTTGTCAAT  
 CCGGAAAATAAGCTCGCAGAGGAACCTTCCAAGGAAATGGTTCACACGGTGTCTCCTGATGTTATGACGTA  
 TGGTATTTGCTTGTGGCTGTGACAATGGGAAGCTTGAAAGGCATTGAAATTGGAGGATTTACAAAG  
 AGTAAGATGGATCTGGTATTGTTATGACACCATCATCGAGGGATGTGCAAGGGTGGAAAGTGGAAAGATG  
 CCTGGAATTATTCTGTAGCCTACCTGTAAGGAGTGAAGCCTAATGTTATGACATACACCGTGATGATTTCAGG  
 15 ATTATGTAAGAAAGGGTCACTGTCTGAAGCAAACATCTGCTTAGAAAATGGAGGAAGATGGGAATGCGCCAAAT  
 GATTGTACATACAACACACTAATCCGGGCACATCTCGAGATGGTACTTAACATGACGCTAAACTATTGAAG  
 AAATGAAGAGTGTGGGTTCTCAGCAGATGCTTCAGTATTAAGATGGTATCGATATGTTATTGAGTGGTGAATT  
 GGACAAAAGCTTCTAGATATGCTTCGTAA

20 SEQ ID NO: 40 is a *Arabidopsis* homolog of the *Petunia Rf-PPR592* gene.

**[0068]** The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 40 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 41 as follows:

25 MKALRLIQPHLLKTGSLRTDLLCTISSFFSSCERDFSSISNGNVCFRERLRSRGIVDIKKDDAIALFOEMIRSRPLP  
 SLVDFSRFFSAIARTKQFNVLDFCKQLELNGIAHNIIYTLNIMINCFCRCKTCFAYSVLGKVMKLYEPDTTTFN  
 TLIKGLFLEGKVSEAVLVDRMVENGCQPDVVTYNSIVNGICRSGDTSIALLRKMEERNVKADVFTYSTIIDS  
 CRDGCIDAAISLFKEMETKGKIKSSVVNTYNSLVRGLCKAGKWNDGALLKDMVSREIVPNVITFNVLLDVFKEGKL  
 QEANEELYKEMITRGISPNIITYNTLMDGYCMQNRSEANNMLDMVRNKCSPDIVFTSLIKGYCMVKRVDDGMKV  
 30 FRNISKRGLVANAVTYSILVQGFCQSGKIKLAEELFQEMVSHGVLVDVMTYGILLGLCDNGKLEKALEIFEDLQK  
 SKMDLGIVMYTTIIEGMCKGGKVEDAWNLFCSLPCKGVKPNVMTYTVMISGLCKKGSLSEANILRKMEEDGNAPN  
 DCTYNTLIRAHLRDGLTASAKLIEEMKSCGFSADASSIKMVIDMLLSGELDKSFLDMLS

**[0069]** Another nucleic acid molecule in accordance with the present invention has a nucleotide sequence of SEQ ID NO: 42, identified herein as *Rf-PPR591*, as follows:

1 ATATATATATACAAACTGATTTCTGTCTATTGACAGTGTATTTTACATACCCCTGAAAAAGGGTAGCTCCGCT  
 81 AATAATGTTATCTTACAAAAAATAACAATACTTTTACATAATATACAAACTCATTCTATGTATTGAAATAT  
 40 161 GATAAAAATATTGTATTTTGTAAATATAGCTATTAGGTAGTCATGTTGTGAAATTCTAAAGGGTAAATTTACCTGAG  
 241 TCGGCCATTGGCTAAAATATTCATTAGTCGATATACTCCAAGCTGTATATCCAGAGCGACAGTATACTTC

321 AATGGTATACTGATATCTTCTATTATACACCTTTAGTATTGATACCCCCAATAGTATAAGTTGACACCGCAATA  
 401 GTGTACCCCAATGTTGTGGTTGTGGCTACAAATTGAGTGATGGTAGTGTAACTTTAGTGTAAAGCTGGTAGTT  
 481 TGAAAACATTCTTTGAAAATTGAGTCAAGTCATAGTACAAAAAAACTGAAATATTATATGTTCTGATTCTGGCT  
 561 GGTCTTCTAAAATTGAAATGCTGGCTAGTTTCAATTAGCGAGGGCAATAGGCTACATGCCAAATTTACGT  
 5 641 AAAAGATAAGTGTGCTGGAAAGTATTGAAAAGATTGAGGGACAAGTGTAGGGCTAGACAACACGTCAAATTATGTA  
 721 GAAAATGCACGAAGAAATTCAAAAGCAAATATTGCTTAAGCAAGAGGCAGTCAGGACAAGTGTGCCTTAGGGAGTGA  
 801 GAAATGGGCATCACTATAAGATTGTATTCATCCGATATTATTATTGATTATAAACTTAAGGAAAAGTGCAGGAAAACCA  
 881 CTAGTTTAGCTATTTTGGAACTTAATCATTATGGGCTGAACCTCAGACTTGTGGGCCAACTTCATACATTGCG  
 961 AAGTAAAATTAGCTCACAGGCCACTTACACTAGTATTGGTTGAAGTCATTTTATGGTTTACATGAGAGA  
 10 1041 CCACCTTTGGAACCTCAATCTTGTGCGCTGAACTTCATGCCTAAGTTAAGTCACCTCAATCCGTAAGGGCTG  
 1121 AATTTTAGGCATAGATGCGTAAACCTCAACCTGTGGACTGAAGTTGAACCTCGCCCTTATGGTGGCCTGAAGTTGAA  
 1201 CTTCAATCCTGTGGCTGAACCTGTGAAGTTCAACCCACAAGGATTAAGTTCAAAAATGACCTCTCAAGCAAA  
 1281 TCTGTAAAAAAAGTGGTCTCTCATGCACTTTACCCATTGCAAAGTAGGCTGAAGTTAGGCCACAATTATTCAAGTT  
 1361 CCAAAAAATTTCACAATATATAACCTCCTTATCTCGGTTATGATTTGATGATTAGCAAATGGACGGGAAAGTGC  
 15 1441 ACGAAAGACCACTTTGCCATTGGCTTTGGGACAGGCCACTAATACAAAATATTAGTTGTGGCTACTTGCTTA  
 1521 AAGAGTTGAACCTCAGTCCAGAGGCCGATTGAAGTTCAAGTCTAAAGATTGAACCTCGATCCAGTGCATATGGACTG  
 1601 AAGTTCAAGTCCTTAAGATGGAACCTCAGTCCAGAGCCATATGGACTGAAGTTCAAGTCAATTATCAGAACTTAAGT  
 1681 CAATATTATTTAGTAAAGGCCAAAAGTGGTTACTATAAGACCAATAAAATAGCGGCCCTAAACTAAATAACAGTGT  
 1761 AAAAGTGGCTGATGGACGAAATTCTACAAAATGGACTCGAGGTAGCAATTCAACCTATGGTGTCAATGTTGA  
 20 1841 CAATTCTCAATCACCCCTACTAAGTGAAGTGAAGCGAAGATGATGAGAAATTCACTGCGTTACTGTCTCAATGGTAAT  
 1921 CCCTTTTCTCATTCTTGCTTATTCAATTGCCACCCGACATTATTCTACCAATACATGTTCCATTTCAGTAAAGGGAA  
 2001 TTTTGGGTTCTAATGAATTTCAGAATGTTAGATGTTAGATGATGCTTCAGTTGTTCCGCAAATGGTTAGAACTA  
 2081 AGCCTCTCCTCTGTTGCCTCTTCTCAAATTGTTGAAAGCTATGGTACATATGACGATTACTCTGTTGTTCT  
 2161 CTTTTCGAGAAATCCACAAATTACGAATTCTGTTCATGAATTCACTTGTGAGCATTGTGGTAAAGCTTGTGCTTAT  
 25 2241 GCATCGTACCGATCTGGATTTCTGTATTAGCATTCACTCAAGAAAAGGCATTCCATATAATGAAAGTCACCTTACTA  
 2321 CCTTAATAAGGGACTTTGCTGAAAATAAGGTCAAAGATGCTGTCATTGTCATAAGTTGGTAAAGTGGGAGAAATATA  
 2401 TGTGAGCCTAATGAAGTCATGTATGGAACGGCATGAATGGCTTGGCAAAAGGGCCATACTCAAAAGCTTTGATT  
 2481 GCTCCGGTTAATGGAACAAGGAACCAAGCCAAATACACGCACTTACACCATTGTCATAGACGCCCTTGCAAAGATG  
 2561 GGATGCTAGATGGTGTACCAAGCCATTGAAATGAGATGAAACAAAAAGCATTCTCCGACATTAACTATAGCACT  
 30 2641 TTAATTGATGCTTGTAAGTTAAGTCAGTGGAAAATGTTAGGACTTGTGCTTGAGATGATACTTAATATT  
 2721 TCCAAATGTGTGCACCTCACTCCGTATTGATGGACTATGCAAAGAGGGAAAGTAGAAAGACGCTGAGGAAATAATGA  
 2801 GATACATGATTGAAAAGGTGTAGACCCGTGATGTCATGACCTATAATATGATAATTGACGGATATGGCTGCGTGGTCAA  
 2881 GTGGATAGAGCACGGAAATTGGATTCCATGATCAATAAGAGCATTGAGCCGATATTAGCTATAATATACTAAT  
 2961 AAATGGATATGCCAGGCAAAGAAAATAGACGAGGCAATGCAAGTCTGCGTGAATTTCTCAAAGGGATTGAAACCTA  
 35 3041 GTATTGTTACCTGCAATGTTCTTGTGATGGCTTTGAACTGGAAAGAACTAAATCTGCACAAAATTCTTGTGAG  
 3121 ATGCTATCTGCGGGCACATCCCTGATTACACTCATTGACTTGTGCTTGGTGGTATTAAAGAATGGACTTGTGA  
 3201 AGAGGCTATGTCACACTCCATAAGTGGAAAAGAGGAGAGAAGATACAATATTCAAAATTACACGGCTGTCATTGATG  
 3281 GATTGTGCAAAATGGTAAGCTGACAAAGCTCATGCTACGTTGAGAAGCTTCCCTGATAGGCTTACATCTGATGTG  
 3361 ATAACATACACTGCAATGATTAGTGGATTGTCAGAAGAGGGTTGTTAGATGAAAGCTAAAGATATGCTAAGGAAAATGGA  
 40 3441 GGACAATGGTGTGGCAGACAACCGAACATACAATGTTATTGTCAGAAGCAATAAGTTAGTGA  
 3521 TGAAGGTTTCTGGAGGAAATAGCTGGAAAGAGCTTCTCATTTGAGGCAGCTACTGTAGAGTTGATGAGTATTATA  
 3601 GCAGAGGATCCATCCATAACACGCCAAATGCACTGGATTAAACTGCACATTGCAATTGAAATACAGGAGATTAGCAGAA  
 3681 AATCACAGGTCCGCCCCAGACAACCCAAAGGCTAAATCCCAATGAAACAAGGTAACATTAAACTTAACTGCCA  
 3761 AAACCTTTAAGAACTATGCAATTGAAACAGGTAATATATATTTCCTTATTGAAACATTCTGATTTATGCGT  
 45 3841 GTCATCCTGTGCAAGAGGCCATGCTAATCTCTCAATCGTCCAGGTTGATTGAAATGATTTAGATTATAC  
 3921 CCCACAGTTCTGCAATTGAAACACCCAAACATTAGTCATCTGCAAAAGGGATTGCTCCTATTATACCATCA  
 4001 TTAAGAAAATCCTGTGACAGTCGGATAAAATGAGCAAATAGTACATGTTGTTATTTCATAAGAGTTGACA

4081 TCTACGGGAAATTATAGTTATCTATGTGGTCGTACTTTAAAGAAAAGTATTTTGTGGTTATAGATTGACTGTTTCCT  
 4161 CTGTCATTGATCGACTTCTTTATTACACATCAGAAGTAGGTATATGTGTACAATGCTTAAACAACGGTTGCATGTG  
 4241 CTCTTATGGTCGCAATTCATCAACGAAGTCTTGTGAGATGCGAGCTGACTGTTAAGACAAGAACCTTCGATT  
 4321 GGAAATGTGATTATCCCATTCAAAGATACTTGACAGATGCTCATGATGCTTATTGACTCAGAAGAATATTCAGAAAAG  
 5 4401 GCATGTAGATGTGCAGCAAATGACAGAGTATGTCAATGGGTGAGGAGGACAATTATACATTGCTCCATTGTCAGTG  
 4481 GCAGGAAATGGCAATCACCTATGGATCTAAAGGACATGTTCTGCATGTAGCTAGAAGGGATGCAAGTTCACAGGGAACTAG  
 4561 GGATTTGGTAGACTATACCGCCTCATTATCAGTTAGTGAATGAAAGAAACCATCAATGTAAGGAAACTCTATGG  
 4641 TTGTACACCTTTGAAGTTCCAAGTGTAAACTAACCTCTGGTGTATATTAGTATATACGGTAGAATTCAATTGCA  
 4721 CAAGTAGATGTATCTTTGCTGGTTTAGTCATTAAGGCATAATGTTCTACTTAGGTTCATGCATTAAATGAAC  
 10 4801 ATTCAATTGATCTATGATGATGGAGTCTTGGTCGTGCATATACATGCTCAAAATTATTGTACAATGGGTTGTGACTCC  
 4881 AATATGTTAACATCATCCACGACATTATCTCTAATAGTTGAGATTTGTGATATTATCGTAAATGCATGTTAAGAT  
 4961 TATTGTAATTAGACTCTAAAGTTCTTTAGTTGGACACAAAGTAATAATCTCAAACACATGTTGGCT  
 5041 TCTTATTCTTGGAAATAAAATATTGAGCTTTACAATGTTGACCTTGGAAATATAAAAGTATTACCTTACTATACCTAT  
 5121 TAAAAATTACATTACTCATGAAATTCAAAGTATCTATCACACTGCGTATTTTTTACTATAAGTCTATATTACCT  
 15 5201 TAGGGCAAAATTAGGCAAGTACTTACCCACATGGGTGTATATACCAATCAACAAAGGAATTTCACACTCTATACCC  
 5281 ATGAAATTAAAGTATTACAGTCATACCCATTAAAAATTACTCTACCCATAAAACTAAAGTATTACCCAAACATACC  
 5361 CATTTCATTGTATACCTTGTGTTAGGTGTACCTTAAGGCTGCATAAAATTATGGTATAAAAGTCTGGAGGACCA  
 5441 TTTATTATTGTTACCTTTTACCTTATAACCTTATAATTATAAGTCTGGAGGGTAGAGTGTACTAATTATTTATGGTAGGC  
 5521 CAGATAATATTGAAAGTAGTGGATAGTGGCTGTAAATTATTTAGATTCGTGGTATTTGTAAAGTGTCTAATCAACA  
 20 5601 AATGCACGTCATTGTTACAATACACTACTACACTTAGCCATAATTAAATTAAAGACATTCTCTTCATTACATCAC  
 5681 ATTACCATAGTTAATTGCTATGGTTAGGTATATATATCCGGTGTAGTAAATTTCATATAAAATTATGGCAAGACGAG  
 5761 TAAATATGAAACTACATGCAGAGGCAGATAAAATTGGTGTAGTAAATTTCGTAAACATGATTAAATTATTC  
 5841 GCGCAAAACCCCTTCAGTTGTTAACGTTACTTATTCGTGTTACCTTACCCATAATGAAATTACCTCATTAGTG  
 5921 CCACATTATCTTCTATAATGTGGTATTGTCAAGAACATCAATCGTGCACCTGCTACATTGTAACATGATT  
 25 6001 CTTTGTGGCTATTAGTCAAAATAGTAACCTGCTTCCATTGTCCTCCGGTCACCTCGGCCACTCCGGCCCTACGTT  
 6081 CATCAAGTACTTATTCATTTTACCTTACGGTGTAAACTTACAATTGTTAATTAAATCATGAAATTAGCT  
 6161 ATACACACATATATAGTAAAAAGGAGATAGTAACCTGAAAGCAGCTCAAGTCAATTTCAGGTTACCTTACGGCAAAATCTTCTATCA  
 6241 GTTATTATGTTGCTTCAAATTAAATAACATATTCAATAGCCGACCTCAACTAATTACGCATTGATGCTAGTTCTATT  
 6321 GTACTAGGAAAAGTAAATTCACTTAAAGTTAAGTTAGTTATTCAGGCAAGTTATATATACACAAATGCAATGTGCTTAT  
 30 6401 ATCCCTTCAATGCTAACTCTGACTTCATGAAAATTAAATTATAGGTGTACTTGTGAGGGACGCGAATTAAATTAA  
 6481 CATCACTGGTAGTGGCGAGCCAGTATTTTACTAAGGAGTATCAAATATAAGTAAATACGAAATATTAAAG  
 6561 GATAGTGAATCTCCTTTAATGTACTTCATTAAAGTTAGTTATTCAGGAAAGTTATATATACACAAATGTTGAC  
 6641 TGATATTCTTGATAATGATGATGCCTATGTGGATAGTGAATCTCCTTTAATGTTGATGAAAAATAAA

35 *Rf-PPR591*, isolated from *Petunia*, has an open reading frame (“ORF”) of 1776 bp, extending between nucleotides 1882-3657, which is homologous to that of *Rf-PPR592*.

40 [0070] The nucleotide sequence of SEQ ID NO: 42 encodes a protein or polypeptide having a deduced amino acid sequence of SEQ ID NO: 43, as follows:

MMRISVRYCLNGNPFFSFAYSIAPRHYSTNTCSISVKGNFGVSNEFQNVKCLDDAFSLFRQMVRTKPLPSVASFS  
 KLLKAMVHMKHYSSVVSLFREIHKLRIPIPHEFILSIVVNSCLMHRDLSFVLAIFKKGIPYNEVTFTLIRGL  
 FAENKVKDADVHLFKKLVRENICEPNEVMYGTVMNGLCKKGHTQKAFDLLRLMEQGSTKPNTRTYTIVIDAFCKDGM  
 LDGATSLNEMKQKSIPPDIFTYSTLIDALCKLSQWENVRTLFLEMIHLNIYPNVCTFNSVIDGLCKEGKVEDAEE

IMRYMIEKGVDVDITYNMIIDGYGLRGQVDRAREIFDSMINKSIEPDIISYNILINGYARQKKIDEAMQVCREIS  
 QKGLKPSIVTCNVLLHGLFELGRTKSAQNFFDEMLSAGHIPLDLYTHCTLLGGYFKNGLVVEAMSHFKLERRREDT  
 NIQIYTAVIDGLCKNGKLDKAHATFEKLPLIGLHPDVITYTAMISGYCQEGLLDEAKDMLRKMEDNGCLADNRTYN  
 VIVRGFLRSNKVSEMKAFLLEEIAKSFSEAAVVELMDIIAEDPSITRKMHWIKLHIA

5

**[0071]** Yet another nucleic acid molecule in accordance with the present invention has a nucleotide sequence of SEQ ID NO: 44, identified herein as *rf-PPR592*, as follows:

10 ATGATGAGAATTGCAGTCGCGTTACTGTCTCAATGGTAATCCCTTTCTATTCTTGCTTATTCAATTGCACCCC  
 GACATTATTCTACCAATAACACGTTCCATTCACTAAAGGGATTGGGTTCTAATGAATTGAGAATGTTAA  
 GTGTTAGATGATGCTTCAGTTGTCGTCAAATGGTTAGAACTAAGCCTCTTCTGTGTTCTCTCT  
 AAATTGTTGAAAGCTTGGTACATATGAAGCATTACTCTTCTGTTGTTCTCTTCGAGAAATCCACAAATTAC  
 GTATTCCCTGTTCATGAATTCACTTGTGAGCATTGTTAACAGTTGTCGCTTATGCATCGTACCGATCTGGATT  
 15 TTCTGTATTAGCCATTCACTTCAGAAAGGTATTCCATTAACTAAGTTATCTTAAACACCTTACTAAGGGGACTC  
 TTTGCTGAAAATAAGGTTAAAGATGCTGTTATTGTTCAAAAAGTTGGTGAGGGAGAATATATGTGAGCCTAATG  
 AAGTCATGTATGGAACGGTCATGAATGGGCTTGCAAAAGGGCCATACTCAAAAGCTTTGATTGCTCCGGTT  
 AATGGAACAAGGAAGTACTAACGCCAATACATGTATCTATAGCATTGTTATCGATGCCTTTGCAAAGATGGGATG  
 CTAGATGGTGTACCAAGCCTTGAAATGAGATGAAACAAAAAGCATTCCCTCCGACATTAACTTATAGCACTT  
 20 TAATTGATGCTTGTAAGTCAAGTCAGTGGAAAATGTTAGGACTTGTGAGATGATACTATTTAATAT  
 TTATCCAAATGTTGTCACCTTCAACTCCGTATTGATGGACTATGCAAAGAGGGGAAAGTAGAAAGACGCTGAGGAA  
 ATAATGAGATACATGATTGAAAAGGTGTAGACCTGTGATGTCACCTATAATATGATAATTGACGGATATGGCT  
 TGCGTGGTCAAGTGGATAGAGCACGGAAATTGGATTCCATGATCAATAAGAGCATTGAGCCAATATTAG  
 CTATAATATACTAATAATGGATATGCCAGGCAAAGAAAATAGACGAGGCAATGCAAGTCTGCCGTGAAATTCT  
 25 CAAAAGGGATTGAAACCTAGTATTGTTACCTGCAATGTTCTTGATGGCTTTGAACTTGGAAAGAACTAAAT  
 CTGCACAAAATTCTTGATGAGATGCTATCTGGGGGACACATACCTGATTATACACTCATTGTTACTTGCTTGG  
 TGGTTATTAAAGAATGGACTTGTGAAAGAGGCTATGTCACACTTCCATAAGTTGGAAAAGAAGGGAGAGAAGATA  
 AATATTCAAATTACACGGCTGTCATTGATGGATTGTGCAAAAGGTAAAGCTCGACAAAGCTCATGCTACGTTG  
 AGAAGCTCCCTGATAGGCTTACATCCTGATGTGATAACACACTGCAATGATTAGTGGATATTGTCAAGAAGG  
 30 GTTGTGTTAGATGAAAGCTAAAGATATGCTAAGGAAAATGGAGGACAATGGTTGTTGGCAGACAACCGAACATACAAT  
 GTTATTGTCGGGGATTCTCAGAAGCAATAAGTTAGTGAAATGAAGGCTTCTGGAGGAAATAGCTGGAAAGA  
 GCTTCTCATTGAGGCAAGCTACTGTAGAGTTATTGATGGATATTAGCAGAGGATCCTTGTAAACATGAT  
 TCCAGAATTTCACCGGGATAATAAGAAGTGA

35 *rf-PPR592* is a gene homologous to *Rf-PPR592* and is isolated from a non-restoring *Petunia* line.

**[0072]** The nucleotide sequence of SEQ ID NO: 44 encodes a protein or polypeptide having a deduced amino acid sequence of SEQ ID NO: 45, as follows:

MMRIAVRYCLNGNPFFSFAYSIAPRHYSTNTRSISVKGNFGVSNEFENVKCLDDAFSLFRQMVRTKPLPSVVSFS  
KLLKALVHMKHYSVSVSLFREIHKLRIPIVHEFILSIVVNSCCLMHRTDLGFSVLAIHFKKGIPFNQVIFNTLLRGL  
FAENKVKDAVHLFKKLVRENICEPNEVMYGTVMNGLCKKGHTQKAFDLLRLMEQGSTKPNTCIYSIVIDAFCKDGM  
LDGATSLLNEMKQKSIPPDIIFTYSTLIDALCKLSQWENVRTLFLLEMIHLNIYPNVCTFNSVIDGLCKEGKVEDAEE  
5 IMRYMIEKGVDVDVITYNMIIDGYGLRGQVDRAREIFDSMINKSIEPNIISYNILINGYARQKKIDEAMQVCREIS  
QKGLKPSIVTCNVLLHGLFELGRTKSAQNFFDEMLSAGHIPDLYTHCTLLGGYFKNGLVEEAMSHFKLERRDE  
NIQIYTAVIDGLCKNGKLDKAHATFEKLPLIGLHPDVITYTAMISGYCQEGLDEAKDMLRKMEDNGCLADNRTYN  
VIVRGFLRSNKVSEMAKAFLEEIAAGKSFSFEAATVELLMDIIAEDPSLLNMIPEFHRDNKK

10 **[0073]** Also suitable in the present invention are other forms of the nucleic acid molecules shown above. An example of a nucleic acid suitable in the present invention is a nucleic acid molecule which hybridizes to a nucleotide sequence of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, 15 SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, or SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

**[0074]** For the purposes of defining the level of stringency, reference can conveniently be made to Sambrook et al., Molecular Cloning: A Laboratory Manual, 20 2nd Edition, Cold Spring Harbor, NY, Cold Spring Harbor Laboratory Press, at 11.45 (1989) which is hereby incorporated by reference in its entirety. An example of low stringency conditions is 4-6X SSC/0.1-0.5% w/v SDS at 37°-45° C for 2-3 hours. Depending on the source and concentration of the nucleic acid involved in the hybridization, alternative conditions of stringency may be employed such as medium 25 stringent conditions. Examples of medium stringent conditions include 1-4X SSC/0.25% w/v SDS at > 45° C for 2-3 hours. An example of high stringency conditions includes 0.1-1X SSC/0.1% w/v SDS at 60 C for 1-3 hours. The skilled artisan is aware of various parameters which may be altered during hybridization and washing and which will either maintain or change the stringency conditions. For 30 example, another stringent hybridization condition is hybridization at 4X SSC at 65° C, followed by a washing in 0.1X SSC at 65° C for about one hour. Alternatively, an exemplary stringent hybridization condition is in 50% formamide, 4XSSC, at 42° C. Still another example of stringent conditions include hybridization at 62° C in 6X SSC, .05X BLOTO, and washing at 2X SSC, 0.1% SDS at 62° C.

[0075] The isolated nucleic acid molecule of the present invention can be from petunia, *Arabidopsis thaliana*, or rice.

[0076] The present invention also relates to an isolated protein encoded by the isolated nucleic acid molecule of the present invention which restores fertility to 5 cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant.

[0077] The present invention also relates to an isolated expression system that contains the nucleic acid molecule of the present invention which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria 10 proteins by the plant. This involves incorporating the nucleic acid molecules of the present invention into host cells using conventional recombinant DNA technology. Generally, this involves inserting the nucleic acid molecule into an expression system to which the nucleic acid molecule is heterologous (i.e., not normally present). The 15 heterologous nucleic acid molecule is inserted into the expression system which includes the necessary elements for the transcription and translation of the inserted protein coding sequences. In one embodiment, the isolated expression system of the present invention contains the nucleic acid molecule of the present invention in proper sense orientation.

[0078] The nucleic acid molecules of the present invention may be inserted 20 into any of the many available expression vectors and cell systems using reagents that are well known in the art. Suitable vectors include, but are not limited to, the following viral vectors such as lambda vector system gt11, gt WES.tB, Charon 4, and plasmid vectors such as pBR322, pBR325, pACYC177, pACYC1084, pUC8, pUC9, pUC18, pUC19, pLG339, pR290, pKC37, pKC101, SV 40, pBluescript II SK +/- or 25 KS +/- (see "Stratagene Cloning Systems" Catalog (1993) from Stratagene, La Jolla, CA, which is hereby incorporated by reference in its entirety), pQE, pIH821, pGEX, pET series (see Studier et al., "Use of T7 RNA Polymerase to Direct Expression of Cloned Genes," Gene Expression Technology, vol. 185 (1990), which is hereby incorporated by reference in its entirety), and any derivatives thereof. Recombinant 30 molecules can be introduced into cells via transformation, particularly transduction, conjugation, mobilization, or electroporation. The DNA sequences are cloned into the vector using standard cloning procedures in the art, as described by Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Edition, Cold Spring Harbor, NY,

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Cold Spring Harbor Laboratory Press (1989); and Ausubel et al., Current Protocols in Molecular Biology, John Wiley & Sons, New York, N.Y., (1989) which are hereby incorporated by reference in their entirety.

[0079] In preparing a DNA vector for expression, the various DNA sequences 5 may normally be inserted or substituted into a bacterial plasmid. Any convenient plasmid may be employed, which will be characterized by having a bacterial replication system, a marker which allows for selection in a bacterium and generally one or more unique, conveniently located restriction sites. Numerous plasmids, referred to as transformation vectors, are available for plant transformation. The 10 selection of a vector will depend on the preferred transformation technique and target species for transformation. A variety of vectors are available for stable transformation using *Agrobacterium tumefaciens*, a soilborne bacterium that causes crown gall. Crown gall are characterized by tumors or galls that develop on the lower stem and main roots of the infected plant. These tumors are due to the transfer and 15 incorporation of part of the bacterium plasmid DNA into the plant chromosomal DNA. This transfer DNA (T-DNA) is expressed along with the normal genes of the plant cell. The plasmid DNA, pTi or Ti-DNA, for "tumor inducing plasmid," contains the vir genes necessary for movement of the T-DNA into the plant. The T-DNA carries genes that encode proteins involved in the biosynthesis of plant 20 regulatory factors, and bacterial nutrients (opines). The T-DNA is delimited by two 25 bp imperfect direct repeat sequences called the "border sequences." By removing the oncogene and opine genes, and replacing them with a gene of interest, it is possible to transfer foreign DNA into the plant without the formation of tumors or the multiplication of *Agrobacterium tumefaciens* (Fraley et al., "Expression of Bacterial 25 Genes in Plant Cells," Proc. Nat'l Acad. Sci., 80:4803-4807 (1983), which is hereby incorporated by reference in its entirety).

[0080] Further improvement of this technique led to the development of the binary vector system (Bevan, "Binary *Agrobacterium* Vectors for Plant Transformation," Nucleic Acids Res., 12:8711-8721 (1984), which is hereby 30 incorporated by reference in its entirety). In this system, all the T-DNA sequences (including the borders) are removed from the pTi, and a second vector containing T-DNA is introduced into *Agrobacterium tumefaciens*. This second vector has the advantage of being replicable in *E. coli* as well as *A. tumefaciens*, and contains a

multiclonal site that facilitates the cloning of a transgene. An example of a commonly used vector is pBin19 (Frisch et al., "Complete Sequence of the Binary Vector Bin19," Plant Molec. Biol., 27:405-409 (1995), which is hereby incorporated by reference in its entirety). Any appropriate vectors now known or later described for 5 genetic transformation are suitable for use with the present invention.

[0081] U.S. Patent No. 4,237,224 issued to Cohen and Boyer, which is hereby incorporated by reference in its entirety, describes the production of expression systems in the form of recombinant plasmids using restriction enzyme cleavage and ligation with DNA ligase. These recombinant plasmids are then introduced by means 10 of transformation and replicated in unicellular cultures including prokaryotic organisms and eukaryotic cells grown in tissue culture.

[0082] Certain "control elements" or "regulatory sequences" are also incorporated into the vector-construct. These include non-translated regions of the vector, promoters, and 5' and 3' untranslated regions which interact with host cellular 15 proteins to carry out transcription and translation. Such elements may vary in their strength and specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, may be used.

[0083] A constitutive promoter is a promoter that directs expression of a gene 20 throughout the development and life of an organism. Examples of some constitutive promoters that are widely used for inducing expression of transgenes include the nopaline synthase ("NOS") gene promoter, from *Agrobacterium tumefaciens* (U.S. Patent No. 5,034,322 issued to Rogers et al., which is hereby incorporated by reference in its entirety), the cauliflower mosaic virus ("CaMV") 35S and 19S 25 promoters (U.S. Patent No. 5,352,605 issued to Fraley et al., which is hereby incorporated by reference in its entirety), those derived from any of the several actin genes, which are known to be expressed in most cell types (U.S. Patent No. 6,002,068 issued to Privalle et al., which is hereby incorporated by reference in its entirety), and the ubiquitin promoter ("ubi"), which is the promoter of a gene product 30 known to accumulate in many cell types.

[0084] An inducible promoter is a promoter that is capable of directly or indirectly activating transcription of one or more DNA sequences or genes in response to an inducer. In the absence of an inducer, the DNA sequences or genes will not be

transcribed. The inducer can be a chemical agent, such as a metabolite, growth regulator, herbicide or phenolic compound, or a physiological stress directly imposed upon the plant such as cold, heat, salt, toxins, or through the action of a pathogen or disease agent such as a virus or fungus. A plant cell containing an inducible promoter 5 may be exposed to an inducer by externally applying the inducer to the cell or plant such as by spraying, watering, heating, or by exposure to the operative pathogen. An example of an appropriate inducible promoter for use in the present invention is a glucocorticoid-inducible promoter (Schena et al., "A Steroid-Inducible Gene Expression System for Plant Cells," Proc. Natl. Acad. Sci., 88:10421-5 (1991), which 10 is hereby incorporated by reference in its entirety). Expression of the transgene-encoded protein is induced in the transformed plants when the transgenic plants are brought into contact with nanomolar concentrations of a glucocorticoid, or by contact with dexamethasone, a glucocorticoid analog (Schena et al., "A Steroid-Inducible Gene Expression System for Plant Cells," Proc. Natl. Acad. Sci. USA, 88:10421-5 15 (1991); Aoyama et al., "A Glucocorticoid-Mediated Transcriptional Induction System in Transgenic Plants," Plant J., 11: 605-612 (1997), and McNellis et al., "Glucocorticoid-Inducible Expression of a Bacterial Avirulence Gene in Transgenic Arabidopsis Induces Hypersensitive Cell Death, Plant J., 14(2):247-57 (1998), which 20 are hereby incorporated by reference in their entirety). In addition, inducible promoters include promoters that function in a tissue specific manner to regulate the gene of interest within selected tissues of the plant. Examples of such tissue specific promoters include seed, flower, or root specific promoters as are well known in the field (U.S. Patent No. 5,750,385 issued to Shewmaker et al., which is hereby 25 incorporated by reference in its entirety). In the preferred embodiment of the present invention, a heterologous promoter is linked to the nucleic acid of the construct, where "heterologous promoter" is defined as a promoter to which the nucleic acid of the construct is not linked in nature.

[0085] The nucleic acid construct also includes an operable 3' regulatory region, selected from among those which are capable of providing correct 30 transcription termination and polyadenylation of mRNA for expression in the host cell of choice, operably linked to a DNA molecule which encodes for a protein of choice. A number of 3' regulatory regions are known to be operable in plants. Exemplary 3' regulatory regions include, without limitation, the nopaline synthase ("nos") 3'

regulatory region (Fraley et al., "Expression of Bacterial Genes in Plant Cells," Proc. Nat'l Acad. Sci. USA, 80:4803-4807 (1983), which is hereby incorporated by reference in its entirety) and the cauliflower mosaic virus ("CaMV") 3' regulatory region (Odell et al., "Identification of DNA Sequences Required for Activity of the 5 Cauliflower Mosaic Virus 35S Promoter," Nature, 313(6005):810-812 (1985), which is hereby incorporated by reference in its entirety). Virtually any 3' regulatory region known to be operable in plants would suffice for proper expression of the coding sequence of the nucleic acid of the present invention.

[0086] The vector of choice, suitable promoter, and an appropriate 3' 10 regulatory region can be ligated together to produce the nucleic acid construct which contains the nucleic acid molecule of the present invention, or suitable fragments thereof, using well known molecular cloning techniques as described in Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Edition, Cold Spring Harbor, NY, Cold Spring Harbor Laboratory Press (1989); Ausubel et al., Current Protocols in 15 Molecular Biology, John Wiley & Sons, New York, N.Y. (1989), which are hereby incorporated by reference in their entirety.

[0087] Once the nucleic acid construct has been prepared, it is ready to be incorporated into a host cell. Accordingly, in another embodiment, the present invention is an isolated host cell containing the nucleic acid molecule of the present 20 invention which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant. Basically, this method is carried out by transforming a host cell with the expression system of the present invention under conditions effective to yield transcription of the nucleic acid molecule in the host cell, using standard cloning procedures known in the art, such as described 25 by Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Edition, Cold Spring Harbor, NY, Cold Spring Harbor Laboratory Press, (1989), which is hereby incorporated by reference in its entirety. Suitable host cells include, but are not limited to, bacteria, virus, yeast, mammalian cells, insect, plant, and the like. Preferably the host cells are either a bacterial cell or a plant cell. Methods of 30 transformation may result in transient or stable expression of the DNA under control of the promoter. Preferably, the nucleic acid construct of the present invention is stably inserted into the genome of the recombinant plant cell as a result of the transformation, although transient expression can serve an important purpose,

particularly when the plant under investigation is slow-growing. Plant tissue suitable for transformation include leaf tissue, root tissue, meristems, zygotic and somatic embryos, callus, protoplasts, tassels, pollen, embryos, anthers, and the like. The means of transformation chosen is that most suited to the tissue to be transformed.

5 [0088] An appropriate method of stably introducing the nucleic acid construct into plant cells is to infect a plant cell with *Agrobacterium tumefaciens* or *Agrobacterium rhizogenes* previously transformed with the nucleic acid construct. As described above, the Ti (or R1) plasmid of *Agrobacterium* enables the highly successful transfer of a foreign DNA into plant cells. Another approach to  
10 transforming plant cells with a gene which imparts resistance to pathogens is particle bombardment (also known as biolistic transformation) of the host cell, as disclosed in U.S. Patent Nos. 4,945,050, 5,036,006, and 5,100,792, all to Sanford et al., and in Emerschad et al., "Somatic Embryogenesis and Plant Development from Immature Zygotic Embryos of Seedless Grapes (*Vitis vinifera*)," Plant Cell Reports, 14:6-12  
15 (1995), which are hereby incorporated by reference in their entirety. Yet another method of introduction is fusion of protoplasts with other entities, either minicells, cells, lysosomes or other fusible lipid-surfaced bodies (Fraley et al., Proc. Natl. Acad. Sci. USA, 79:1859-63 (1982), which is hereby incorporated by reference in its entirety). The DNA molecule may also be introduced into the plant cells by  
20 electroporation (Fromm et al., Proc. Natl. Acad. Sci. USA, 82:5824 (1985), which is hereby incorporated by reference in its entirety). In this technique, plant protoplasts are electroporated in the presence of plasmids containing the expression cassette. Electrical impulses of high field strength reversibly permeabilize biomembranes allowing the introduction of the plasmids. Electroporated plant protoplasts reform the  
25 cell wall, divide, and regenerate. The precise method of transformation is not critical to the practice of the present invention. Any method that results in efficient transformation of the host cell of choice is appropriate for practicing the present invention.

[0089] After transformation, the transformed plant cells must be regenerated.  
30 Plant regeneration from cultured protoplasts is described in Evans et al., Handbook of Plant Cell Cultures, Vol. 1, MacMillan Publishing Co., NY (1983); Vasil (ed.), Cell Culture and Somatic Cell Genetics of Plants, Acad. Press, Orlando, Vol. I, 1984, and Vol. III (1986); and Fitch et al., "Somatic Embryogenesis and Plant Regeneration

from Immature Zygotic Embryos of Papaya (*Carica papaya* L.)," Plant Cell Rep., 9:320 (1990), which are hereby incorporated by reference in their entirety.

[0090] Means for regeneration vary from species to species of plants, but generally a suspension of transformed protoplasts or a petri plate containing explants 5 is first provided. Callus tissue is formed and shoots may be induced from callus and subsequently rooted. Alternatively, embryo formation can be induced in the callus tissue. These embryos germinate as natural embryos to form plants. The culture media will generally contain various amino acids and hormones, such as auxin and cytokinins. Efficient regeneration will depend on the medium, on the genotype, and 10 on the history of the culture. If these three variables are controlled, then regeneration is usually reproducible and repeatable.

[0091] Preferably, transformed cells are first identified using a selection marker simultaneously introduced into the host cells along with the nucleic acid construct of the present invention. Suitable selection markers include, without 15 limitation, markers encoding for antibiotic resistance, such as the nptII gene which confers kanamycin resistance (Fraley et al., "Expression of Bacterial Genes in Plant Cells," Proc. Natl. Acad. Sci. USA, 80:4803-4807 (1983), which is hereby incorporated by reference in its entirety), and the genes which confer resistance to gentamycin, G418, hygromycin, streptomycin, spectinomycin, tetracycline, 20 chloramphenicol, and the like. Cells or tissues are grown on a selection medium containing the appropriate antibiotic, whereby generally only those transformants expressing the antibiotic resistance marker continue to grow. Other types of markers are also suitable for inclusion in the expression cassette of the present invention. For example, a gene encoding for herbicide tolerance, such as tolerance to sulfonylurea is 25 useful, or the dhfr gene, which confers resistance to methotrexate (Bourouis et al., EMBO J., 2:1099-1104 (1983), which is hereby incorporated by reference in its entirety). Similarly, "reporter genes," which encode for enzymes providing for production of an identifiable compound are suitable. The most widely used reporter gene for gene fusion experiments has been uidA, a gene from *Escherichia coli* that 30 encodes the  $\beta$ -glucuronidase protein, also known as GUS (Jefferson et al., "GUS Fusions:  $\beta$  Glucuronidase as a Sensitive and Versatile Gene Fusion Marker in Higher Plants," EMBO J., 6:3901-3907 (1987), which is hereby incorporated by reference in its entirety). Similarly, enzymes providing for production of a compound identifiable

by luminescence, such as luciferase, are useful. The selection marker employed will depend on the target species; for certain target species, different antibiotics, herbicide, or biosynthesis selection markers are preferred.

[0092] Plant cells and tissues selected by means of an inhibitory agent or other selection marker are then tested for the acquisition of the transgene by Southern blot hybridization analysis, using a probe specific to the transgenes contained in the given cassette used for transformation (Sambrook et al., "Molecular Cloning: A Laboratory Manual," Cold Spring Harbor, NY, Cold Spring Harbor Laboratory Press (1989), which is hereby incorporated by reference in its entirety).

[0093] After the expression cassette is stably incorporated in transgenic plants, it can be transferred to other plants by sexual crossing. Any of a number of standard breeding techniques can be used, depending upon the species to be crossed. Once transgenic plants of this type are produced, the plants themselves can be cultivated in accordance with conventional procedure so that the nucleic acid construct is present in the resulting plants. Alternatively, transgenic seeds or propagules (e.g., cuttings) are recovered from the transgenic plants. These seeds can then be planted in the soil and cultivated using conventional procedures to produce transgenic plants.

[0094] Thus, in other embodiments, the present invention includes transgenic plants and seeds produced by transformation with the nucleic acid molecule of the present invention which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant. Examples of transgenic plants include crop plants such as alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane. Other examples of transgenic plants include ornamental plants such as *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

[0095] The nucleic acid molecule of the present invention can be utilized to restore fertility to cytoplasmic male sterile plants for a wide variety of crop plants and ornamental plants. Thus, the present invention also relates to a method of restoring fertility to cytoplasmic male sterile plants involving transforming a cytoplasmic male

sterile plant with a nucleic acid molecule of the present invention under conditions effective to restore fertility to the cytoplasmic male sterile plant. The plant can have 2 or more copies of the nucleic acid molecule.

[0096] The nucleic acid molecule of the present invention can be utilized in a 5 method for identifying genes affecting male fertility or mitochondrial gene expression in other species. The first PPR gene family member whose function was characterized is *crp1*, a gene involved in RNA processing in chloroplasts (Fisk et al., “Molecular Cloning of the Maize Gene *crp1* Reveals Similarity Between Regulators of Mitochondrial and Chloroplast Gene Expression,” EMBO J., 18: 2621-2630 10 (1999), which is hereby incorporated by reference in its entirety). *Rf-PPR592* and *crp1* exhibit some sequence similarity, though there are other PPR motif proteins in the databases with greater similarity to *Rf-PPR592*. Both of these genes affect accumulation of particular RNA transcripts.

[0097] Thus, another aspect of the present invention relates to a method of 15 identifying a candidate gene restoring fertility in plants. The method involves analyzing the candidate gene for the presence of the above nucleic acid molecule in accordance with the present invention.

[0098] Identification of the nucleic acid molecules of the present invention suggests new strategies for identification of restorers or nuclear male sterility (ms) 20 alleles in crop species that are more important agriculturally than petunia. Thus, another aspect of the present invention relates to a method of identifying a candidate plant suitable for breeding with a cytoplasmic male sterile plant. The method involves analyzing the candidate plant for the presence, in its genome, of the above nucleic acid molecule of the present invention.

[0099] For example, it is possible that a nucleic acid molecule of the present 25 invention corresponds to a restorer allele in rice. Since the complete genome sequence of rice is publicly available, using the above-described method for identifying PPR motif-containing genes, candidates for rice restorer genes can be identified in the rice chromosomal region which is genetically linked to the rice 30 restoration phenotype. Using standard methods of cloning and rice transformation, the candidate rice restorer gene can be introduced as a transgene into a rice CMS line and the fertility of the transformants can be evaluated to determine whether the PPR gene is actually a restorer gene.

**[0100]** The fact that the nucleic acid molecules of the present invention can be a PPR motif gene can also be used to identify putative genes in other species that might encode male sterility when disrupted. The homolog of a restorer gene in one species could, when mutated, be a male sterility-encoding gene in another species.

5 Creating a male sterile line can be valuable for certain applications. For example, flowers of petunia and some other horticultural species undergo a phenomenon called pollination-induced senescence (Xu et al., "Programmed Cell Death During Pollination-Induced Petal Senescence in Petunia," Plant Phys., 122:1323-1333 (2000), which is hereby incorporated by reference in its entirety). Flowers are triggered to 10 senesce when pollinated. A male sterile flower will last longer when the plant is male sterile, because no self pollen will be available to pollinate it.

**[0101]** Since it is possible to introduce genes into yeast mitochondria, it is likely that methods for introducing genes into plant mitochondria can be developed. When it becomes possible to introduce the *pcf* gene or a toxic homolog containing the 15 sequences on which the nucleic acid molecule of the present invention operates, then a new CMS/restorer system can be created in a different species. In such a system, a male sterile line is created by introducing *pcf* or a *pcf* homolog into the mitochondrial genome. This CMS line can then be crossed with a line containing the nucleic acid 20 molecule of the present invention in the nuclear genome, introduced by standard transformation methods, to create hybrid seed that will give rise to fertile progeny plants.

**[0102]** Thus, the present invention also relates to a method of producing hybrid plant seed. The method first involves providing a cytoplasmic male sterile plant. Next, a second plant containing the above nucleic acid molecule in accordance 25 with the present invention is provided. Finally, the cytoplasmic male sterile plant and the second plant are bred under conditions effective to produce hybrid progeny seed which yield fertile plants.

**[0103]** Another aspect of the present invention relates to a method of producing plant seeds for an inbred line of plants. The method first involves 30 providing a cytoplasmic male sterile plant. Next, a second plant containing the above nucleic acid molecule in accordance with the present invention is provided. Then, the cytoplasmic male sterile plant and the second plant are bred under conditions effective to produce hybrid progeny seed which yield fertile plants. Next, hybrid fertile plants

are produced from the hybrid progeny seeds. Finally, the hybrid fertile plants and the second plant are backcrossed to produce seed which yield inbred progeny plants.

**[0104]** Backcrossing methods can be used with the present invention to improve or introduce a characteristic into the present cultivar. The term backcrossing 5 refers to the repeated crossing of a hybrid progeny back to one of the parental plants for that hybrid. The parental plant which contributes the locus for the desired characteristic is termed the nonrecurrent or donor parent. This terminology refers to the fact that the nonrecurrent parent is used one time in the backcross protocol and, therefore, does not recur. The parental plant to which the locus or loci from the 10 nonrecurrent parent are transferred is known as the recurrent parent as it is used for several rounds in the backcrossing protocol (Poehlman et al., Breeding Field Crops, 4th Ed., Ames, Iowa, Iowa State University Press, (1995); Fehr, ed., Principles of Cultivar Development, Vol. 1: Theory and Technique, NY, NY, Macmillan Publishing Company (1987); and Fehr, ed., Principles of Cultivar Development, Vol. 15 2: Crop Species, NY, NY, Macmillan Publishing Company (1987), which are hereby incorporated by reference in their entirety).

**[0105]** In a typical backcross protocol, the phenotypically and/or commercially appealing cultivar or accession (recurrent parent) is crossed with a second cultivar (nonrecurrent parent) that carries the single locus of interest (e.g., the 20 GSB resistance gene locus) to be transferred. The resulting progeny from this cross are then crossed again to the recurrent parent and the process is repeated until a plant is obtained where essentially all of the desired morphological and physiological characteristics of the recurrent parent are recovered in the converted plant, in addition to the single transferred locus from the nonrecurrent parent.

**[0106]** The selection of a suitable recurrent parent is an important step for a 25 successful backcrossing procedure. The goal of a backcross protocol is to alter or substitute a single trait or characteristic in the original cultivar or accession. To accomplish this, a single locus of the recurrent cultivar is modified or substituted with the desired locus from the nonrecurrent parent, while retaining essentially all of the 30 rest of the desired genetic, and therefore the desired physiological and morphological constitution of the original cultivar. The choice of the particular nonrecurrent parent will depend on the purpose of the backcross; one of the major purposes is to add some commercially desirable, agronomically important trait to the plant. The exact

backcrossing protocol will depend on the characteristic or trait being altered to determine an appropriate testing protocol. Although backcrossing methods are simplified when the characteristic being transferred is a dominant allele, a recessive allele may also be transferred. In this instance it may be necessary to introduce a test 5 of the progeny to determine if the desired characteristic has been successfully transferred.

[0107] A technique, known as modified backcrossing, uses different recurrent parents during the backcrossing. Modified backcrossing may be used to replace the original recurrent parent with a cultivar having certain more desirable characteristics 10 or multiple parents may be used to obtain different desirable characteristics from each.

[0108] Presently, it is possible to introduce genes into chloroplast genomes, which are maternally inherited in many species. Therefore, a CMS/restorer system can also be created by introducing the *pcf* locus, modified for chloroplast expression, 15 into the chloroplast genome. The nucleic acid molecule of the present invention can be modified to replace the mitochondrial transit sequence with a chloroplast transit sequence, using standard methods (Kohler et al., "Exchange of Protein Molecules Through Connections Between Higher Plant Plastids," Science, 276:2039-2042 (1997), which is hereby incorporated by reference in its entirety), so that the protein 20 encoded by the nucleic acid molecule of the present invention will turn off toxic gene expression in the chloroplast. Without the restorer, PCF in the chloroplast is likely to be toxic; PCF is toxic to *E. coli* bacteria. Thus, the present invention also relates to a method of producing plants with a cytoplasmic male sterile plant restoration system. The method first involves transforming a first plant in its chloroplast genome with a 25 nucleic acid which causes the plant to become male sterile. Next, a second plant is transformed with the above nucleic acid molecule in accordance with the present invention whose protein product is targeted to the chloroplast. Finally, the first and second plants are crossed to produce progeny plants possessing a cytoplasmic male sterile plant restoration system.

[0109] Since the nucleic acid molecule of the present invention prevents the expression of an organelle gene, it can be used to control the expression of a chimeric gene introduced into chloroplasts. If there is a useful protein that is desired to be produced from plant chloroplasts by introduction of a gene encoding the valuable 30

protein into the chloroplast genome, production of the valuable protein could deliberately be turned off by expressing the nucleic acid molecule of the present invention from a conditional promoter. When desired, the expression of the nucleic acid molecule of the present invention could be turned off, so that the valuable protein

5 is produced. In further detail, the method of the present invention involves: (1) engineering a chimeric gene including the coding region of a desirable protein and the *pcf* gene sequences that are regulated by the nucleic acid molecule of the present invention; (2) introducing this chimeric gene into chloroplasts of a plant and obtaining a chloroplast transgenic line; (3) engineering the nucleic acid molecule of the present

10 invention or its homologs so that the protein is targeted into chloroplasts; (4) introducing the engineered nucleic acid molecule of the present invention into the nuclear genome of a plant and obtaining a nuclear transgenic line; and (5) crossing plants to set up a regulated system. Alternatively, the plant in (2) can be made first and the gene in (3) introduced by transformation, or the plant in (4) can be made first

15 and the gene in (2) introduced into the plant in (4). When it becomes possible to transform plant mitochondrial genomes, the chimeric gene containing the *pcf* sequence can be introduced into mitochondria and the product of the nucleic acid molecule of the present invention can be targeted to mitochondria to create the analogous system.

20 [0110] The nucleic acid molecule of the present invention must be expressed in most of the plant, because in every tissue examined, the PCF protein is reduced in the presence of the nucleic acid molecule of the present invention (Nivison et al., "Identification of a Mitochondrial Protein Associated With Cytoplasmic Male Sterility in *Petunia*," *Plant Cell*, 1:1121-30 (1989), Nivision et al., "Sequencing, Processing, and Localization of the *Petunia* CMS-Associated Mitochondrial Protein," *Plant J.*, 5:613-623 (1994), which are hereby incorporated by reference in their entirety). The PCF protein does nevertheless vary in abundance (Conley et al., "Tissue-Specific Protein Expression in Plant Mitochondria," *Plant Cell*, 6:85-91 (1994), which is hereby incorporated by reference in its entirety) and is more highly

25 expressed in anthers. Very few promoters have been identified that confer expression in many tissues and in developing microspores at an early stage of pollen development. Thus, the promoter sequence of the nucleic acid molecule of the

present invention could be useful to express any of many different coding regions in a variety of tissues.

[0111] It is also possible to dissect the regulatory sequences of the nucleic acid molecule of the present invention by standard methods to identify those regions 5 that confer expression in particular tissue types. Typically, such regulatory sequences are 5' to the coding region, though the 3' flanking region can also be important. The most novel aspect of the regulatory sequences of the nucleic acid molecule of the present invention is that they confer expression in early microsporogenesis. Most of the published anther-specific promoters are not effective at the early stage of pollen 10 development, when it is critical to restore proper mitochondrial function in plants carrying the CMS cytoplasm. For example, the promoter of a nucleic acid molecule of the present invention could be used with a different coding region to restore fertility to a species with a different CMS-encoding gene. Alternatively, the promoter could be used to control a gene toxic to pollen to confer male sterility, or regulatory 15 elements from this promoter could be combined with those of another promoter to confer expression in early microsporogenesis.

[0112] Thus, another aspect of the present invention relates to a method of directing gene expression to plant mitochondria. The method involves transforming a plant with a chimeric nucleic acid molecule containing a transgene operatively linked 20 to a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant. The promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1. The terminator has a nucleotide sequence of from 25 nucleotide 3761 to 4593 of SEQ ID NO: 1.

[0113] The present invention also relates to a promoter from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant. The 30 promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1.

[0114] Another aspect of the present invention relates to a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to

direct expression of the transgene in the mitochondria of the transformed plant. The terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

[0115] Another aspect of the present invention relates to a nucleic acid construct. The nucleic acid construct includes: (i) a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant and (ii) a nucleic acid heterologous to and operatively coupled to the promoter or the terminator. The promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1. The terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

[0116] Other embodiments of the present invention include isolated expression systems, host cells, transgenic plants, and transgenic plant seeds containing the nucleic acid construct of the present invention.

[0117] Another aspect of the present invention is a method of expressing a gene preferentially in roots of a plant. The method involves transforming a plant with a nucleic acid construct containing a promoter suitable for driving expression preferentially in roots having a nucleotide sequence of from 1 to 1388 of SEQ ID NO: 44; a nucleic acid heterologous to the promoter, where the promoter is operatively coupled 5' to the nucleic acid to induce transcription of the nucleic acid; and a terminator having a nucleotide sequence of from nucleotide 3168 to 4016 of SEQ ID NO: 44, where the terminator is operably coupled 3' to the nucleic acid. This method can be used to express genes in roots of a plant, but not in stems, leaves, or buds.

[0118] The nucleic acid molecule of the present invention or its homologues could also be used to deliberately alter floral morphology to produce novel flowers. Thus, in another embodiment, the present invention is a method of altering plant floral morphology in ornamental plants by transforming an ornamental plant with a nucleic acid molecule of the present invention which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant. Combinations of certain nuclear genes with particular mitochondrial backgrounds have been found to result in altered floral morphology in some genera. For example, in tobacco, combinations of the nuclear genome of one species with the cytoplasm of

another sometimes results in very abnormal flowers, such as flowers in which anthers have been converted to petals. While these particular plants may not be desirable horticulturally, it is possible that the coding region or expression of the nucleic acid molecule of the present invention could be manipulated so that interesting, valuable 5 floral alterations could be obtained. For example, flowers with a second set of petals in place of anthers could be attractive. A similar strategy could be pursued with other species in which novel floral morphology is desirable. Manipulation of the nucleic acid molecule of the present invention could occur, for example, by overexpressing it on a different promoter, changing the coding region, or using standard antisense or 10 gene silencing methods to underexpress homologous genes.

**[0119]** Another aspect of the present invention relates to a method of producing plants with a cytoplasmic male sterile plant restoration system. The method first involves mutagenizing a first plant having a nucleic acid which encodes a protein. The protein has a motif having an amino acid sequence corresponding to any 15 of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input. Next, the mutagenized first plant is crossed with a wild-type plant having mitochondrial 20 DNA polymorphisms compared to mitochondrial DNA in the mutagenized first plant to produce progeny plants. Finally, it is determined if the progeny plants are fertile, whereby fertile progeny plants can be used as a fertile maintainer line, where the mutagenized first plant, the fertile maintainer line, and a wild-type allele present in the first plant before mutagenesis comprises a new cytoplasmic male sterile plant 25 restoration system. Figures 7A-B show this aspect of the present invention in detail.

**[0120]** The *Arabidopsis thaliana* genome sequence contains a PPR gene highly similar to the nucleic acid molecule of the present invention. There are existing, publicly available insertional element collections that can be screened by 30 standard methods to find a mutant in which the homolog of the nucleic acid molecule of the present invention is disrupted. The mutant can be examined to determine whether it encodes male sterility. Because the nucleic acid molecule of the present invention is important for nuclear/organelle interaction to produce male fertility, its

homologs in other species are likely to be essential for proper pollen development in those species.

[0121] By mutating a PPR gene in a plant that does not have a CMS/restorer system, such a system can be created. In this strategy, a PPR gene is mutated and the 5 plant becomes male sterile. The mutated PPR alleles are then crossed with plants carrying other cytoplasms, present in other varieties of the plant or in intercrossable species. If a cytoplasm can be found in which the plant is fertile in the presence of the two mutated PPR alleles, then a new CMS/restorer system will have been created. A line carrying the new cytoplasm plus the mutated alleles becomes the maintainer line. 10 The line carrying the first cytoplasm plus the mutated alleles becomes *rf/rf* CMS. A line carrying an unmutated allele plus a mutated allele in the presence of the CMS cytoplasm becomes *Rf/rf* CMS. These lines can then be exploited just as standard maintainer, sterile, and restored lines are currently used in hybrid seed production (Figure 7).

[0122] For example, tomato does not have a CMS/restorer system. It is known that markers near petunia *Rf* map to a region of the tomato genome where two nuclear male sterility alleles exist. Possibly, the tomato ortholog of the nucleic acid molecule of the present invention, when mutated, results in male sterility. If so, then the cytoplasms of the intercrossable wild tomato species can be tested to determine 20 whether they can confer male fertility to a tomato line homozygous for the mutated PPR gene.

[0123] The nucleic acid molecule of the present invention may not be usable directly to restore fertility to CMS lines of most other species. Current information indicates that different mitochondrial genes are present in different CMS lines. In 25 most cases, restorer genes will have a specific mechanism of action—suppression of expression of the abnormal mitochondrial gene. However, by chance, there may be a few species that carry a CMS cytoplasm whose abnormality can be ameliorated by the nucleic acid molecule of the present invention. This can be determined by introducing the nucleic acid molecule of the present invention into the other species 30 and determining whether the transgenic plants become male fertile. If so, the nucleic acid molecule of the present invention can be used as a fertility restorer for this species.

## EXAMPLES

[0124] The following examples are provided to illustrate embodiments of the present invention but are by no means intended to limit its scope.

5 **Example 1 – Identification of Two PPR-Containing ORFs as Potential Candidates for the Rf Gene**

[0125] Previously, the isolation of a 37.5-kb BIBAC clone, SB5, that cosegregates with the Rf gene has been reported (Bentolila et al., “Identification of a 10 BIBAC Clone That Co-Segregates With the Petunia Restorer of Fertility (Rf) Gene,” *Mol. Genet. Genomics*, 266:223-230 (2001), which is hereby incorporated by reference in its entirety). SB5 is part of a contig that was constructed by screening a Petunia BIBAC library with a marker, EACA/MCTC, tightly linked to Rf. No recombination was identified between EACA/MCTC and Rf after examining 15 1,078 meiotic events. The genetic delimitation of the Rf locus was achieved only partially on the BIBAC contig. One extremity of the contig was separated from Rf by the occurrence of four recombination events, whereas no crossing-over was found between Rf and the other extremity (Bentolila et al., “Identification of a BIBAC Clone That Co-Segregates With the Petunia Restorer of Fertility (Rf) Gene,” *Mol. Genet. Genomics*, 266:223-230 (2001), which is hereby incorporated by reference in 20 its entirety). Because of the possibility that Rf might lie further away in the area not covered by the contig, a walk was initiated by screening the BIBAC library with a probe lying on the extremity that cosegregates with Rf. Unfortunately, the only hits were clones already isolated in the contig, demonstrating the presence of a gap in the 25 Petunia BIBAC library.

[0126] Before increasing the redundancy of the library to find new clones covering the gap, it was determined whether the Rf gene might lie in the SB5 clone. Because the BIBAC vector is a binary vector allowing Agrobacterium-mediated plant transformation (Hamilton, “A Binary-BAC System for Plant Transformation With 30 High-Molecular-Weight DNA,” *Gene*, 200:107-116 (1997), which is hereby incorporated by reference in its entirety), SB5 was used to restore fertility to CMS plants. Unfortunately, although SB5 is stable in *E. coli*, it underwent multiple rearrangements when introduced into *A. tumefaciens*, thus precluding its use in

transgenic experiments. Randomly chosen clones of various sizes did not show this instability in *A. tumefaciens*, pointing to special features in the sequence of the SB5 insert.

[0127] To address whether Rf might lie in the SB5 clone, shotgun sequencing 5 of the entire clone was carried out and the predicted ORFs for candidate Rf genes were examined.

[0128] Because of difficulties in contig assembly caused by the presence of 10 repeated sequences, BamHI subclones rather than the entire BIBAC clone were used as the starting material for shotgun sequencing. DNA was sonicated into 1- to 3-kb fragments, which were gel purified (Geneclean Spin Kit, Bio 101, Vista, CA), end-repaired with T4 DNA polymerase (GIBCO/BRL, Rockfield, MD) in the presence of all four dNTPs, and ligated at a mass ratio of 3 inserts to 1 vector into the SmaI site of the pTrueBlue vector (Genomics One, Buffalo, NY). The ligation product was introduced into Electromax DH10B Escherichia coli cells (GIBCO/BRL), and DNA 15 obtained from the white colonies by minipreparation was sequenced with the T7 primer in the Cornell BioResource Center. The sequences were assembled into contigs with SEQUENCHER (Gene Codes, Ann Arbor, MI).

[0129] ORFs from BIBAC SB5, their promoter region, and poly(A) signals 20 were predicted by using GENSCAN (Burge et al., "Prediction of Complete Gene Structures in Human Genomic DNA," *J. Mol. Biol.*, 268:78-94 (1997), which is hereby incorporated by reference in its entirety) with the Arabidopsis parameter matrix. Duplicated blocks in the Rf locus were determined by aligning the genomic sequence against itself by using the dot-plot feature from the MEGALIGN program (DNAstar, Madison, WI) with a 90% match. The presence of a transit peptide in the 25 ORFs was determined by using PREDOTAR version 0.5 , TARGETP (Emanuelsson et al., "Predicting Subcellular Localization of Proteins Based on Their N-Terminal Amino Acid Sequence," *J. Mol. Biol.*, 300:1005-1016 (2000); Nielsen et al., "Identification of Prokaryotic and Eukaryotic Signal Peptides and Prediction of Their Cleavage Sites," *Prot. Eng.*, 10:1-6 (1997), which are hereby incorporated by 30 reference in their entirety), and MITOPROT (Scharfe et al., *Nucleic Acids Res.*, 28:155-158 (2000), which is hereby incorporated by reference in its entirety). The length of the transit peptide was predicted by TARGETP and MITOPROT.

[0130] PPR motifs were identified in Rf-PPR592 and Rf-PPR591 by the MEME software (Bailey et al., in "Proceedings of the Second International Conference on Intelligent Systems for Molecular Biology," in Altman, eds. (Am. Assoc. Artificial Intelligence Press, Menlo Park, CA), pp. 28-36 (1994), which is hereby incorporated by reference in its entirety). The parameters for motif searching were set as minimum width = 35, maximum width = 35. The PPR consensus motif computed from the comparison of 1,303 motifs has been described previously (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," *Trends Biochem. Sci.*, 25:46-47 (2000), which is hereby incorporated by reference in its entirety).

[0131] Because the Rf gene is expected to be targeted to mitochondria where it can act upon the pcf gene to prevent its expression, an ORF predicted to carry mitochondrial transit sequences was searched. Two ORFs with putative mitochondrial targeting signals were identified. The two ORFs are adjacent to each other and appear to have originated from duplications in the promoter and coding region, but carry divergent 3' flanking regions (Figure 1A). The ORFs were 92% identical at the nucleotide level, and the predicted proteins were 93% similar, with C termini that differ completely in their final 12 aa. Both ORFs carry PPR motifs; one encodes 591 aa and the other encodes 592 aa, and were therefore named Rf-PPR591 and Rf-PPR592. A third PPR-containing ORF might lie in the vicinity of the two PPR-containing ORFs shown in Figure 1A. On the left extremity lies a genomic block that shares high similarity with the end of the coding sequence of Rf-PPR592 and its terminator region.

[0132] According to cleavage prediction programs, both putative proteins exhibited 28-residue mitochondrial transit peptides. Predicted transit peptides of Rf-PPR592 and Rf-PPR591 differed by only one substitution. To determine whether the predicted transit peptide could target a passenger protein to mitochondria, 44 codons from the 5' end of the Rf-PPR592 coding region were inserted 5' to the coding region of an enhanced GFP. DNAs of this construct and of one known to target GFP to mitochondria were bombarded into onion epidermal cells. Both GFPs appeared to be localized to the same type of organelle in the single cells shown in Figures 1B and C. Because the predicted transit peptides of Rf-PPR592 and Rf-PPR591 differed by only

one amino acid, it was expected that not only Rf-PPR592 but also Rf-PPR591 would be mitochondrially localized.

[0133] Most of the predicted mature protein (87%) of Rf-PPR592 consisted of 14 PPRs (Figure 1D). These repeats extended from the amino acid in position 54 to 5 the amino acid in position 544 and are organized in two sets of tandem repeats, one set containing 3 PPRs from amino acid 54 to amino acid 158, the other set containing 11 PPRs from amino acid 160 to amino acid 544. Because the Rf-PPR591 and Rf-PPR592 proteins are 93% similar and differ mainly in the last 12 C-terminal amino acids, their organization with respect to PPRs is identical. There was a very good 10 agreement between the consensus motif derived from the 14 PPRs found in Rf-PPR592 (hereafter designated 14 PPR consensus) and the consensus motif derived from 1,303 PPRs (hereafter designated 1303 PPR consensus) reported previously (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," *Trends Biochem. Sci.*, 25:46-47 (2000), which is hereby incorporated by 15 reference in its entirety) (Figure 1D). Whenever a discrepancy occurred between the consensus motif of the 14 PPRs in Rf-PPR592 and the 1303 PPR consensus, the difference usually was a conservative substitution. For instance, the aspartic acid in the first position of the 14 PPR consensus is replaced by a glutamic acid in the 1303 PPR consensus. Moreover, when the most frequent amino acid in the 14 PPR 20 consensus at a given position differed from the corresponding amino acid found in the 1303 PPR consensus, the amino acid in the 1303 consensus was generally the second most frequent in the 14 PPR consensus (glutamic acid at position 1, asparagine at position 18, alanine at position 28, tyrosine at position 29; Figure 1D).

[0134] It has been demonstrated that *Rf-PPR592*, a gene encoding a 592-aa 25 protein containing 14 PPRs, was able to restore fertility to CMS plants. The PPR motif, a degenerate 35-aa repeat, has been found in a very large gene family in the *Arabidopsis* genome (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," *Trends Biochem. Sci.*, 25:46-47 (2000), which is hereby 30 incorporated by reference in its entirety). The repeats are organized in tandem arrays with the number of motifs per peptide ranging from 2 to 26. About two-thirds of these *Arabidopsis* PPR proteins are predicted to be targeted to either mitochondria or chloroplasts (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," *Trends Biochem. Sci.*, 25:46-47 (2000), which is hereby

incorporated by reference in its entirety). Although distinct from the tetratricopeptide repeat (TPR), a motif that is likely to be involved in protein binding, the PPR motif shares with the former a predicted spatial structure consisting of two  $\alpha$ -helices (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," Trends Biochem. Sci., 25:46-47 (2000); Das et al., "The Structure of the Tetratricopeptide Repeats of Protein Phosphatase 5: Implications for TPR-Mediated Protein-Protein Interactions," EMBO J., 17:1192-1199 (1998), which are hereby incorporated by reference in their entirety). Tandem PPRs are thought to form a superhelix with a central spiral groove that presumably serves as the ligand-binding surface in a similar way as the one predicted for the tandem TPRs (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," Trends Biochem. Sci., 25:46-47 (2000), which is hereby incorporated by reference in its entirety). However, unlike in the TPR motif, the side chains lining the central groove of the PPR are almost exclusively hydrophilic, suggesting that some or all of the PPR motifs are RNA-binding rather than protein-binding motifs. This hypothesis is supported by the involvement in RNA metabolism and/or translation of the very few PPR motif-containing proteins characterized so far: maize chloroplast CRP1, involved in chloroplast *petD* RNA processing and *petD* and *petA* translation (Fisk et al., "Molecular Cloning of the Maize Gene *Crp1* Reveals Similarity Between Regulators of Mitochondrial and Chloroplast Gene Expression," EMBO J., 18:2621-2630 (1999), which is hereby incorporated by reference in its entirety), *Chlamydomonas* MCA1, required for the accumulation of the chloroplast *petA* transcript (Lown et al., "Chlamydomonas Nuclear Mutants That Fail to Assemble Respiratory or Photosynthetic Electron Transfer Complexes," Biochem. Soc. Trans., 29:452-455(2001), which is hereby incorporated by reference in its entirety), yeast PET309, required for the stability and translation of the *coxI* mitochondrial mRNA (Manthey et al., "The Product of the Nuclear Gene PET309 is Required for Translation of Mature mRNA and Stability or Production of Intron-Containing RNAs Derived from the Mitochondrial COX1 Locus of *Saccharomyces cerevisiae*," EMBO J., 14:4031-4043 (1995), which is hereby incorporated by reference in its entirety), and *Drosophila* BSF, which binds to and stabilizes the bicoid mRNA (Mancebo et al., "BSF Binds Specifically to the *bicoid* mRNA 3' Untranslated Region and Contributes to Stabilization of *bicoid* mRNA," Mol. Cell. Biol., 21:3462-3471 (2001), which is

hereby incorporated by reference in its entirety). That *Petunia Rf* belongs to this family is consistent with its similarity of action to *crp1*, *mca1*, and *pet309*. Mutations in these three genes result in lack of accumulation of a particular transcript and reduced abundance of an organelle protein. Likewise, in *Petunia* restored plants, 5 among the population of *pcf* transcripts with different 5' termini, the ones with termini at -121 exhibit reduced abundance and the amount of the PCF protein is greatly reduced (Pruitt et al., "Transcription of the Petunia mitochondrial CMS-Associated Pcf Locus in Male Serile and Fertility-Restored Lines," Mol. Gen. Genet., 227:348-355 (1991); Nivison et al., "Identification of a Mitochondrial Protein Associated with 10 Cytoplasmic Male Sterility in Petunia," Plant Cell, 1:1121-1130 (1989), which are hereby incorporated by reference in their entirety). However, the alleles of the other PPR genes that are known to reduce RNA and/or protein accumulation are recessive, whereas the *Petunia Rf* allele is dominant. *Rf* genes from other species have been shown to alter the RNA transcript profile of the CMS-associated genes (Wise et al., 15 "Mutator-Induced Mutations of the *rfl* Nuclear Fertility Restorer of T-Cytoplasm Maize Alter the Accumulation of T-*urf13* Mitochondrial Transcripts," Genetics, 143:1383-1394 (1996); Singh et al., "Suppression of Cytoplasmic Male Sterility by Nuclear Genes Alters Expression of a Novel Mitochondrial Gene Region," Plant Cell, 3:1349-1362 (1991); Tang et al., "Transcript Processing Internal to a Mitochondrial 20 Open Reading Frame is Correlated with Fertility Restoration in Male-Sterile Sorghum," Plant J., 10:123-133 (1996); Moneger et al., "Nuclear Restoration of Cytoplasmic Male Sterility in Sunflower is Associated with the Tissue-Specific Regulation of a Novel Mitochondrial Gene," EMBO J., 13:8-17 (1994), which are hereby incorporated by reference in their entirety). In some cases, restoration has 25 been shown to result from enhanced processing of the CMS-associated transcripts (Tang et al., "Transcript Processing Internal to a Mitochondrial Open Reading Frame is Correlated with Fertility Restoration in Male-Sterile Sorghum," Plant J., 10:123-133 (1996); Menassa et al., "Post-Transcriptional and Developmental Regulation of a CMS-Associated Mitochondrial Gene Region by a Nuclear Restorer Gene," Plant J., 30 17:491-499 (1999), which are hereby incorporated by reference in their entirety). Taken together, these observations suggest that *Rf*s in other species could also be PPR-containing genes like the *Petunia Rf*.

[0135] The data presented here show that a pair of duplicated PPR-containing genes, denoted *Rf-PPR591* and *Rf-PPR592*, lie in the *Petunia Rf* locus. A third related PPR gene might lie in the area not covered by the SB5 BIBAC clone as suggested by the high similarity between the sequence available at the end of the clone and the sequence present at the end of the coding sequence of *Rf-PPR592* and in its terminator region.

[0136] In *Brassica napus*, the restorer locus has been shown to affect the transcripts of several mitochondrial genes, two of them being associated with the *nap* and *pol* CMS (Singh et al., "Nuclear Genes Associated With a Single Brassica CMS Restorer Locus Influence Transcripts of Three Different Mitochondrial Gene Regions," *Genetics*, 143:505-516 (1996); Li et al., "Restorer Genes for Different Forms of *Brassica* Cytoplasmic Male Sterility Map to a Single Nuclear Locus That Modifies Transcripts of Several Mitochondrial Genes," *Proc. Natl. Acad. Sci. USA*, 95:10032-10037 (1998), which are hereby incorporated by reference in their entirety).

At the same locus have been mapped *Rfp*, the restorer gene to the *pol* CMS, that modifies the transcripts of the *pol* CMS-associated *orf224/atp6* mitochondrial DNA region, *Rfn*, the restorer gene to the *nap* CMS that modifies the transcripts of the *nap* CMS-associated *orf222/nad5c/orf139* mitochondrial DNA region, and *Mmt* (modifier of mitochondrial transcripts), a gene that modifies the transcripts of the *nad4* gene and another gene possibly involved in cytochrome *c* biogenesis (Li et al., "Restorer Genes for Different Forms of *Brassica* Cytoplasmic Male Sterility Map to a Single Nuclear Locus That Modifies Transcripts of Several Mitochondrial Genes," *Proc. Natl. Acad. Sci. USA*, 95:10032-10037 (1998), which is hereby incorporated by reference in its entirety). The resolution of the genetic mapping in these studies did not allow the authors to address whether the three genes represent different alleles of a single gene or whether the restorer locus might contain multiple, related, tightly linked genes. A similar situation occurs in *Sorghum*, where at the *Rf3* locus, one of the two restorers to A3 CMS, has been mapped a gene that regulates the transcript-processing activity of A3 CMS-associated *orf107* and the *Mmt1* gene that enhances the transcript processing of *urf209* (Tang et al., "Cosegregation of Single Genes Associated with Fertility Restoration and Transcript Processing of Sorghum Mitochondrial *orf107* and *urf209*," *Genetics*, 150:383-391 (1998), which is hereby incorporated by reference in its

entirety). As in *Brassica napus*, either a multiallelic model or tightly linked genes could account for this result.

[0137] It will be worthwhile to determine whether *Rf-PPR591* affects the profile of mitochondrial transcripts other than *pcf* in transgenic plants. If so, it would 5 strengthen the hypothesis that *Rf* alleles arise as modifications, perhaps through duplication, of existing alleles that control mitochondrial gene expression. According to this theory, once CMS occurs in a plant species, there may be strong selective pressure for the plant to overcome it by recruiting preexisting activities and redirecting them to down-regulate the expression of CMS-encoding genes.

10 Conceivably, recombination among closely related PPR-containing genes could have led to the appearance of the *Rf-PPR592* gene.

**Example 2 - A Deletion in the Promoter of *rf-PPR592* Prevents Its Expression in CMS Floral Buds**

[0138] If one of the candidate ORFs, *Rf-PPR591* or *Rf-PPR592*, is the *Rf* gene, some sequence polymorphism between the allele of these ORFs found in a restorer line (*Rf/Rf*) and the allele found in a CMS plant (*rf/rf*) might be expected. Presumably some difference in the sequences of the dominant *Rf* allele vs. the 15 recessive nonrestoring allele *rf* must reflect their opposite restoring ability. The sequence of *rf-PPR592* was obtained by amplifying genomic DNA of a Petunia hybrida *rf/rf* plant, where *rf* was inherited from a *P. hybrida* line called 2423, with the PfuTurbo Hotstart DNA polymerase (Stratagene, La Jolla, CA) and PCR primers flanking *Rf-PPR591* (5'-TGCACAGTGTATATTACATACCC-3'; SEQ ID NO: 20 46) and *Rf-PPR592* (5'-TTTATGATACATGGATTCAACGAC-3'; SEQ ID NO: 47). A PCR product was obtained only with a primer specific to the 3' flanking region of *Rf-PPR592*, not with a primer specific to the 3' flanking region of *Rf-PPR591*. The *rf-PPR592* PCR product showed a reduction in size of about 500nt compared with the 25 *Rf-PPR592* PCR product amplified from the genomic DNA of an *Rf/Rf* line (Figure 2A). Using the same primers, a PCR product similar in size to *rf-PPR592* was 30 amplified from another nonrestoring *P. hybrida* line as well as from a nonrestoring *Petunia parodii* line. The *rf-PPR592* PCR product amplified from the *P. hybrida* 2423 sequence was cloned into the pCR-Blunt II-TOPO vector (Invitrogen, Carlsbad, CA) and sequenced, revealing a gene 97% identical to *Rf-PPR591* and 94% identical

to Rf-PPR592 in the coding region, with the predicted proteins 98% and 94% similar, respectively. Because of the primer design, the rf-PPR592 sequence lacks 35 nt available for Rf-PPR592. Similarity blocks between rf-PPR592, Rf-PPR592, and Rf-PPR591 were determined by comparing the aligned sequences with SEQUENCHER.

5 Percent similarity was computed by using the MEGALIGN program. Comparison of the similarities of regions of the three different PPR genes revealed that the 5' promoter region of rf-PPR592 is most similar to Rf-PPR591, whereas the 3' flanking region of rf-PPR592 is most similar to Rf-PPR592. The genomic structure of rf-PPR592 was consistent with the past occurrence of recombination between two genes 10 similar to Rf-PPR591 and Rf-PPR592 (Figure 2B). Because PCR amplification could have resulted in an artificial recombination between Rf-PPR591 and Rf-PPR592 due to their high similarity, the Rf-PPR592 PCR product was resequenced as a control experiment. The sequences of three rf-PPR592 and Rf-PPR592 clones were determined. No evidence of recombination was found in any of the sequenced Rf- 15 PPR592 clones, thus precluding PCR amplification as the source of the genetic mosaic found in the rf-PPR592 ORF.

20 [0139] rf-PPR592 carries a 530-nt deletion from -556 to -27 relative to the start codon of Rf-PPR592. This deletion is responsible for the observed difference in the sizes of the respective amplicons. Rf-PPR591 has a 49-nt gap within the same region, from -273 to -224 relative to the start codon of Rf-PPR592 (Figure 2B).

25 [0140] RT-PCR experiments were performed to determine whether both Rf-PPR592 and rf-PPR592 are expressed in Petunia floral buds. The RT reaction was performed with Superscript II RNase H- reverse transcriptase (GIBCO), and the PCR was performed with the PfuTurbo Hotstart DNA polymerase. The reverse primer R3 used for reverse transcription (RT)-PCR lies in the 3' untranslated region of the Rf-PPR592 gene at position +430 to +454. The forward primer used for the PCR lies in the coding sequence and is specific to the rf or Rf allele, F2S or F2, respectively, because of DNA polymorphisms between rf and Rf in this area. Primer pairs F2SR3 amplified a 1333-bp product and F2R3 amplified a 1507-bp product. R3, 5'-TGAAAATGACAATCGAACAGAAAA-3' (SEQ ID NO: 48); F2, 5'-AACATTCCCTCCAGACATTATTACA-3' (SEQ ID NO: 49); F2S, 5'-GACGCTGAGGAAATAATGAGATAC-3' (SEQ ID NO: 50).

5 [0141] An Rf-PPR592 transcript was detected in floral buds in lines carrying the Rf allele, but no transcripts of rf-PPR592 were detected in a homozygous nonrestoring rf/rf line (Figure 3A). The absence of the upstream 530-nt region in rf-PPR592 is likely to prevent the expression of PPR592 in the floral buds of nonrestoring lines.

10 [0142] Since rf-PPR592 encodes a protein that is very similar to the one encoded by Rf-PPR592, a survey of its expression was conducted in tissues other than the floral buds. From all of the tissues analyzed, an rf-PPR592 transcript was detected only in roots of a nonrestoring rf/rf line (Figure 3B).

15 [0143] A deletion of 530 nt in the promoter area of the *rf-PPR592* gene is the likely cause of its nonexpression in the floral buds of CMS plants. That the *rf-PPR592* gene, which encodes a protein 98% similar to Rf-PPR591 and 94% similar to Rf-PPR592, has not yet accumulated missense mutations suggests either a recent deletion in the promoter or a functional expression in plant organs other than the floral buds. This latter possibility was supported by the finding of an *rf-PPR592* transcript in the roots of homozygous nonrestoring *rf/rf* line.

20 [0144] Sequence inspection demonstrated that a recombination event between two genes similar to *Rf-PPR591* and *Rf-PPR592* can explain the formation of *rf-PPR592*. Perhaps once *Rf-PPR592* was generated and happened to prevent the expression of *pcf*, its maintenance required the presence of the CMS-associated gene. The absence of the CMS-associated gene in new nucleocytoplasmic combinations might have resulted in recombination between *Rf-PPR591* and *Rf-PPR592* because of their high similarity. In *Brassica* and related genera, *Rfn* is found only in association with the *nap* cytoplasm, suggesting that the evolutionary appearance of the *nap* cytoplasm and the attending male sterility may have provided the selective pressure for the origin, and possibly the continued presence, of *Rfn* in *B. napus* (Li et al., “Restorer Genes for Different Forms of *Brassica* Cytoplasmic Male Sterility Map to a Single Nuclear Locus That Modifies Transcripts of Several Mitochondrial Genes,” *Proc. Natl. Acad. Sci. USA*, 95:10032-10037 (1998), which is hereby incorporated by reference in its entirety). Sampling of more *rf-PPR592* genes from different *Petunia* species should help in understanding the evolution of CMS and fertility restoration in this genus.

**Example 3 – Rf-PPR592 Is Able to Restore Fertility to CMS Plants**

[0145] A sequence encoding the N-terminal 44 aa of Rf-PPR592 was inserted 5' to the green fluorescent protein (GFP) sequence in the pOL vector (Peeters et al., 5 “Duplication and Quadruplication of *Arabidopsis Thaliana* Cysteinyl- and Asparaginyl-tRNA Synthetase Genes of Organellar Origin,” *J. Mol. Evol.*, 50:413-423 (2000), which is hereby incorporated by reference in its entirety) to use in transient assay of protein localization. As a control, a vector carrying GFP fused with a known mitochondrial coxIV transit peptide (Akashi et al., “Potential Dual Targeting 10 of an *Arabidopsis* Archaeabacterial-Like Histidyl-Trna Synthetase to Mitochondria and Chloroplasts,” *FEBS Lett.*, 431:39-44 (1998), which is hereby incorporated by reference in its entirety) was also used in the transient assays. DNAs of GFP constructs were bombarded into onion epidermal cells as described in Scott et al., “Model System For Plant Cell Biology: GFP Imaging In Living Onion Epidermal 15 Cells,” *BioTechniques*, 26:1125, 1128-1132 (1999), which is hereby incorporated by reference in its entirety.

[0146] For the stable transformation experiments, genomic DNA from the Rf-PPR592 gene was amplified from the SB5 BIBAC clone with the PfuTurbo Hotstart DNA polymerase and the primers F11-XbaI (5'-  
20 TCTAGAAAAAAATGAAGGGGGATCAAT-3'; SEQ ID NO: 51) and R11-EcoRI (5'-GAATTCACTTGCTCTCACGATAAACTAAGA-3'; SEQ ID NO: 52) (underlined are the restriction sites added to the 5' end of the primers for further use in the cloning of the PCR product). The PCR product was first cloned into the pCR-Blunt II-TOPO vector, and its sequence was checked to be free of possible mutations 25 generated by the polymerase. The PCR product was then released from the pCR-Blunt II-TOPO vector by digestion with XbaI and EcoRI, gel purified, and cloned into XbaI/EcoRI-digested binary vector pGPTVKan (Becker et al., “New Plant Binary Vectors With Selectable Markers Located Proximal to the Left T-DNA Border,” *Plant Mol. Biol.*, 20:1195-1197 29 (1992), which is hereby incorporated by reference in its entirety). Petunia transformation and regeneration were performed as described in Horsch et al., “A Simple and General-Method for Transferring Genes Into Plants,” *Science*, 227:1229-1231 30 (1985), which is hereby incorporated by reference in its 30 entirety. Transformants were selected on 300 mg/liter kanamycin, 100 mg/liter

ticarcillin/clavulanic acid (15:1, Duchefa Biochemie, Harlem, The Netherlands).

Shoots were rooted on N13 medium (O'Connell et al., "Somatic Hybridization Between *Lycopersicon-Esculentum* and *Lycopersicon-Pennellii*," Theor. Appl. Genet., 70:1-12 (1985), which is hereby incorporated by reference in its entirety)

5 before transfer to soil.

[0147] To determine whether Rf-PPR592 could restore fertility to rf/rf CMS lines, a 4.6-kb fragment carrying the entire coding region was introduced into the binary vector pGPTVKan. This fragment carries 2007 nt upstream of the start codon and 861 nt downstream of the stop codon. The pGPTVKan-4.6 kb Rf-PPR592 vector 10 was transferred into *A. tumefaciens* strain LBA4404, which was used to transform a *P. parodii* rf/rf CMS line (Figure 4A) and a *P. hybrida* rf/rf CMS line (Figure 4C). More than two dozen independent transformants were obtained and grown to 15 flowering. Fertile transformants were observed after transformation of both lines (Figures 4B and D). Among these were several fertile transformants carrying a single copy of the introduced Rf-PPR592 genomic DNA. Flowers of one of the *P. parodii* primary transformant plants were selfed, and a population of 40 T1 progeny was grown to flowering.

[0148] DNA extractions and Southern blotting were performed as described in Bentolila et al., "Locating the Petunia Rf Gene on a 650 kb DNA Fragment," Theor. Appl. Genet., 96:980-988 (1998), which is hereby incorporated by reference in its entirety. Floral bud protein was prepared for cell culture protein as described in Kohler et al., "The Green Fluorescent Protein as a Marker to Visualize Plant - Mitochondria *in vivo*," Plant Journal, 11:613-621 (1997), which is hereby incorporated by reference in its entirety. After separation by SDS/PAGE (15%), 25 immunoblots on Hybond-P poly(vinylidene difluoride) membranes (PVDF; Amersham Pharmacia, Picataway, NJ) were prepared as previously described (Reed et al., "High-Level Expression of a Synthetic Red-Shifted GFP Coding Region Incorporated into Transgenic Chloroplasts," Plant J., 27:257-2653 (2001), which is hereby incorporated by reference in its entirety) and probed with a 1:5000 dilution of 30 the anti-PCF antibody (Nivison et al., "Sequencing, Processing, and Localization of the Petunia CMS-Associated Mitochondrial Protein," Plant J., 5:613-623 (1994), which is hereby incorporated by reference in its entirety).

[0149] DNA blot hybridization revealed that the fertile phenotype cosegregated with the Rf-PPR592 transgene (Figure 5A). The T1 progeny were also surveyed for the presence of the CMS-associated 19.5-kDa PCF protein. The 19.5-kDa protein was found to be decreased about 10-fold in fertile progeny restored by 5 Rf-PPR592 relative to sterile progeny and the parental CMS line (Figure 5B). Thus, Rf-PPR592 was capable of restoring fertility by decreasing the amount of the PCF protein.

[0150] The cloning of a gene that can restore fertility to male-sterile *Petunia* lines will facilitate elucidation of the mechanism by which expression of the CMS-10 associated mitochondrial gene is suppressed. The reduced amount of the PCF protein could be due to a reduction in the abundance of one of the *Petunia* CMS-associated transcripts, which was reported previously (Pruitt et al., "Transcription of the *Petunia* Mitochondrial CMS-Associated *pcf* Locus in Male Sterile and Fertility-Restored Lines," Mol. Gen. Genet., 227:348-355 (1991), which is hereby incorporated by 15 reference in its entirety), or to a translation defect that destabilizes the transcript. In yeast, mutation in a transcript-specific translation factor destabilizes the particular transcript with which the factor normally interacts (Poutre et al., "PET111, a *Saccharomyces cerevisiae* Nuclear Gene Required for Translation of the Mitochondrial mRNA Encoding Cytochrome C Oxidase Subunit II," Genetics, 20: 115:637-647 (1987), which is hereby incorporated by reference in its entirety).

[0151] A number of fertility restorer genes in other species are known to alter transcript profiles and mitochondrial gene product accumulation (Moneger et al., "Nuclear Restoration of Cytoplasmic Male Sterility in Sunflower is Associated with the Tissue-Specific Regulation of a Novel Mitochondrial Gene," EMBO J., 13:8-17 25 (1994); Singh et al., "Nuclear Genes Associated With a Single Brassica CMS Restorer Locus Influence Transcripts of Three Different Mitochondrial Gene Regions," Genetics, 143:505-516 (1996); Dewey et al., "Novel Recombinations in the Maize Mitochondrial Genome Produce a Unique Transcriptional Unit in the Texas Male-Sterile Cytoplasm," Cell, 44:439-49 (1986); Wise et al., "Mitochondrial Transcript 30 Processing and Restoration of Male Fertility in T-Cytoplasm Maize," J Hered, 90:380-385 (1999), which are hereby incorporated by reference in their entirety). In addition to the molecular phenotype of restoration, the *Petunia* Rf locus and Rf loci from other species may be similar in genomic organization (Li et al., "Restorer Genes

for Different Forms of *Brassica* Cytoplasmic Male Sterility Map to a Single Nuclear Locus That Modifies Transcripts of Several Mitochondrial Genes," Proc. Natl. Acad. Sci. USA, 95:10032-10037 (1998); Tang et al., "Cosegregation of Single Genes

Associated with Fertility Restoration and Transcript Processing of Sorghum

5 Mitochondrial *orf107* and *urf209*," Genetics, 150:383-391 (1998), which are hereby incorporated by reference in their entirety). The identification of *Petunia Rf* as a PPR family member suggests that searching for PPR motif genes near known restorer loci should be a useful strategy to identify candidate restorer genes in other species. Further studies of *Rf-PPR592* and other PPR motif-containing genes in plants, fungi, 10 and animals will be required to determine whether the motif has a direct role in RNA-protein and/or protein-protein interactions.

**Example 4 – Use of *Rf-PPR592* or its Homologs/Derivatives to Create Novel Floral Structures**

15 [0152] In Petunia, recombination events near the *Rf* locus in standard sexual crosses resulted in plants with abnormal floral appearance. Moreover, a few of the initial transgenic plants transformed by *Rf-PPR592* produced flowers with abnormal appearance. Furthermore, a number of transgenic plants transformed by *Rf-PPR592* 20 and *Rf-PPR591* exhibit abnormalities in floral and vegetative structures. An example of abnormal flowers seen in some transgenic plants are shown in Figure 6.

**Example 5 - Identification of a Rice Fertility Restorer Gene**

25 [0153] The complete rice genome sequence, which has been deposited in EMBL/GenBank/DDBJ, was examined for genes similar to the petunia *Rf* gene, using BLASTP. The gene most similar to the petunia *Rf* locus was termed as *Rice homolog of Petunia restorer 1 (Rhpr1)*. This gene is located very close to the rice *Rf4* marker C1261. There were a total of 10 PPR genes in the vicinity of this marker on rice 30 chromosome 10, which were termed as *Rhpr1* to *Rhpr10* (SEQ ID NOS: 20, 22, 24, 26, 28, 30, 32, 34, 36, and 38).

[0154] The most immediate usefulness of identification of the rice restorer gene is in marker-assisted selection. This facilitates introduction of the natural wild abortive-restorer *Rf4* gene, which can restore fertility to the wild abortive cytoplasm

by traditional crosses into elite breeding lines for use as a parent in a three-line breeding scheme. This use of the information does not involve genetically modified organisms and, therefore, can proceed without any of the attendant issues. Random screening has identified some molecular markers that are already being used to 5 transfer *Rf* genes in certain nuclear backgrounds (Ichikawa et al., "A Rapid PCR-Aided Selection of a Rice Line Containing the *Rf-I* Gene Which is Involved in Restoration of the Cytoplasmic Male Sterility," Molecular Breeding, 3:195-202 (1997); Jing et al., "Mapping Fertility-Restoring Genes of Rice WA Cytoplasmic 10 Male Sterility Using SSLP Markers," Bot. Bull. Acad. Sin., 42:167-171 (2001), which are hereby incorporated by reference in their entirety). Knowing the actual *Rf* gene sequence makes laborious screening for markers suitable between different breeding 15 lines unnecessary.

[0155] The next possibility is to more rapidly transfer the *Rf4* gene into existing elite breeding lines by transformation rather than by sexual crosses. In such a 15 strategy, the entire natural *Rf4* gene would be used to transform a rice line for the three-line hybrid rice production method.

[0156] Because the three-line method for hybrid rice production requires time-consuming breeding and labor, presently there are attempts to exploit temperature-sensitive male sterility mutants for a two-line method of hybrid seed production. The 20 three-line method for hybrid rice production involves construction of three lines. Two lines are backcrossed repeatedly so that they contain the same nuclear genome. One contains the CMS cytoplasm ("CMS parent") and is male sterile while the other ("Fertile Maintainer") contains the normal cytoplasm but no restorer of fertility (*Rf*) alleles. By crossing the maintainer as male and the CMS line as female, seeds of the 25 CMS line with a known nuclear background can be produced in large quantity. The third line is homozygous for one or more fertility restoration loci. Hybrid seed is produced by crossing the third line with the CMS parent. The nuclear genomes of the third line and the CMS line are selected by breeders to optimize heterosis and desirable characteristics for the region in which the hybrid rice will be grown.

[0157] Rice plants have been found that contain a mutant allele that encodes 30 male sterility at high temperatures but fertility at low temperatures (Dong et al., "Molecular Mapping of a Rice Gene Conditioning Thermosensitive Genic Male Sterility Using AFLP, RFLP and SSR Techniques," Theor Appl Genet., 100:727-734

(2000), which is hereby incorporated by reference in its entirety). By growing the rice at high temperatures, it can be used as the sterile parent in a cross with an elite breeding line. The mutant rice can be propagated by selfing when grown at low temperatures. The use of this method in the field on large-scale has not been reported 5 in the literature, so the feasibility of using a natural temperature-sensitive mutant is not known.

[0158] A cloned *Rf4* gene could also be used in a two-line method. In this scheme, the *Rf4* gene regulatory sequences would be engineered so that it could be turned on when desired. Then, both a CMS line and a maintainer line, which requires 10 multiple crosses over a number of years to produce, are not needed. A single line, the CMS line containing the engineered *Rf4* gene, would serve both as CMS parent and as its own maintainer line (Figure 8). The CMS line would be propagated by selfing by turning on the *Rf4* gene. Without induction of the engineered *Rf4* gene, however, the line would be sterile and therefore could be used as a CMS parent.

[0159] Although the invention has been described in detail for the purpose of illustration, it is understood that such detail is solely for that purpose, and variations 15 can be made therein by those skilled in the art without departing from the spirit and scope of the invention which is defined by the following claims.

**WHAT IS CLAIMED:**

1. An isolated nucleic acid molecule which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant, wherein the nucleic acid molecule: (1) encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41; (2) encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified by a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input; (3) hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C; or (4) has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

2. The isolated nucleic acid molecule according to claim 1, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

3. The isolated nucleic acid molecule according to claim 1, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ

ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

4. The isolated nucleic acid molecule according to claim 1, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

5. The isolated nucleic acid molecule according to claim 1, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

6. The isolated nucleic acid molecule according to claim 1, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

7. A isolated protein encoded by the nucleic acid molecule according to claim 1.

8. The isolated protein according to claim 7, wherein the protein has an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

9. The isolated protein according to claim 7, wherein the protein encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or

an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

10. The isolated protein according to claim 7, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

11. The isolated protein according to claim 7, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

12. The isolated protein according to claim 7, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

13. An isolated expression system comprising the nucleic acid molecule of claim 1.

14. The isolated expression system according to claim 13, wherein the nucleic acid molecule is in proper sense orientation.

15. An isolated host cell comprising the nucleic acid molecule of claim 1.

16. The isolated host cell according to claim 15, wherein the nucleic acid molecule is in an expression system.

17. The isolated host cell according to claim 15, wherein the host cell is a plant cell.

18. The isolated host cell according to claim 15, wherein the host cell is a bacterial cell.

19. A transgenic plant transformed with the nucleic acid molecule according to claim 1.

20. The transgenic plant according to claim 19, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

21. The transgenic plant according to claim 19, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

22. The transgenic plant according to claim 19, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

23. The transgenic plant according to claim 19, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID

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NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

24. The transgenic plant according to claim 19, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

25. The transgenic plant according to claim 19, wherein the transgenic plant is a crop plant.

26. The transgenic plant according to claim 25, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

27. The transgenic plant according to claim 19, wherein the transgenic plant is an ornamental plant.

28. The transgenic plant according to claim 27, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

29. A transgenic plant seed transformed with the nucleic acid molecule according to claim 1.

30. The transgenic plant seed according to claim 29, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

31. The transgenic plant seed according to claim 29, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence

corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

32. The transgenic plant seed according to claim 29, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

33. The transgenic plant seed according to claim 29, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

34. The transgenic plant seed according to claim 29, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

35. The transgenic plant seed according to claim 29, wherein the transgenic plant is a crop plant.

36. The transgenic plant seed according to claim 35, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini,

cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

37. The transgenic plant seed according to claim 29, wherein the transgenic plant is an ornamental plant.

38. The transgenic plant seed according to claim 37, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

39. A method of restoring fertility to cytoplasmic male sterile plants comprising:

transforming a cytoplasmic male sterile plant with a nucleic acid molecule according to claim 1 under conditions effective to restore fertility to the cytoplasmic male sterile plant.

40. The method according to claim 39, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

41. The method according to claim 39, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

42. The method according to claim 39, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO:

40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

43. The method according to claim 39, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

44. The method according to claim 39, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

45. The method according to claim 39, wherein the plant is a crop plant.

46. The method according to claim 45, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

47. The method according to claim 39, wherein the plant is an ornamental plant.

48. The method according to claim 47, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

49. The method according to claim 39, wherein the plant has 2 or more copies of the nucleic acid molecule.

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50. A method of identifying a candidate plant suitable for breeding with a cytoplasmic male sterile plant comprising:

analyzing the candidate plant for the presence, in its genome, of a nucleic acid molecule according to claim 1.

51. The method according to claim 50, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

52. The method according to claim 50, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

53. The method according to claim 50, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

54. The method according to claim 50, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

55. The method according to claim 50, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

56. The method according to claim 50, wherein the plant is a crop plant.

57. The method according to claim 56, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

58. The method according to claim 50, wherein the plant is an ornamental plant.

59. The method according to claim 58, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

60. A method of identifying a candidate gene restoring fertility in plants, said method comprising:

analyzing the candidate gene for the presence of a nucleic acid molecule according to claim 1.

61. The method according to claim 60, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

62. The method according to claim 60, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a

NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

63. The method according to claim 60, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

64. The method according to claim 60, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

65. The method according to claim 60, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

66. The method according to claim 60, wherein the plant is a crop plant.

67. The method according to claim 66, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

68. The method according to claim 60, wherein the plant is an ornamental plant.

69. The method according to claim 68, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

70. A method of producing hybrid plant seed comprising:  
providing a cytoplasmic male sterile plant;  
providing a second plant comprising a nucleic acid molecule according to claim 1; and

breeding the cytoplasmic male sterile plant and the second plant under conditions effective to produce hybrid progeny seed which yield fertile plants.

71. The method according to claim 70, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

72. The method according to claim 70, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

73. The method according to claim 70, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

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74. The method according to claim 70, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

75. The method according to claim 70, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

76. The method according to claim 70, wherein the plants are crop plants.

77. The method according to claim 76, wherein the crop plants are selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

78. The method according to claim 70, wherein the plants are ornamental plants.

79. The method according to claim 78, wherein the ornamental plants are selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

80. The hybrid progeny seed produced by the method according to claim 70.

81. The hybrid progeny plants grown from the hybrid progeny seed produced by the method according to claim 70.

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82. A method of producing plant seeds for an inbred line of plants comprising:

providing a cytoplasmic male sterile plant;

providing a second plant comprising a nucleic acid molecule according to claim 1;

breeding the cytoplasmic male sterile plant and the second plant under conditions effective to produce hybrid progeny seed which yield fertile plants;

producing hybrid fertile plants from the hybrid progeny seeds;

and

backcrossing the hybrid fertile plants and the second plant to produce seed which yield inbred progeny plants.

83. The method according to claim 82, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

84. The method according to claim 82, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

85. The method according to claim 82, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

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86. The method according to claim 82, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

87. The method according to claim 82, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

88. The method according to claim 82, wherein the plants are crop plants.

89. The method according to claim 88, wherein the crop plants are selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

90. The method according to claim 82, wherein the plants are ornamental plants.

91. The method according to claim 90, wherein the ornamental plants are selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

92. The seed which yield inbred progeny plants produced by the method according to claim 82.

93. The inbred progeny plants grown from the seed which yield inbred progeny plants produced by the method of claim 82.

94. A method of directing gene expression to plant mitochondria comprising:

transforming a plant with a chimeric nucleic acid molecule comprising a transgene operatively linked to a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant, wherein the promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1 and the terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

95. The method according to claim 94, wherein the method is carried out with a promoter having a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1.

96. The method according to claim 94, wherein the method is carried out with a terminator having a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

97. The method according to claim 94, wherein the plant is a crop plant.

98. The method according to claim 97, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

99. The method according to claim 94, wherein the plant is an ornamental plant.

100. The method according to claim 99, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

101. A promoter from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant, wherein the promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1.

102. A terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant, wherein the terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

103. A nucleic acid construct comprising:  
a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant and  
a nucleic acid heterologous to and operatively coupled to the promoter or the terminator, wherein the promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1 and the terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

104. A nucleic acid construct according to claim 103, wherein the construct is provided with a promoter from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant, said promoter having a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1.

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105. A nucleic acid construct according to claim 103, wherein the construct is provided with a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant, said terminator having a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

106. An isolated expression system comprising the nucleic acid construct according to claim 103.

107. An isolated host cell comprising the nucleic acid construct according to claim 103.

108. A transgenic plant comprising the nucleic acid construct according to claim 103.

109. A transgenic plant seed comprising the nucleic acid construct according to claim 103.

110. A method of expressing a gene preferentially in roots of a plant comprising:

transforming a plant with a nucleic acid construct comprising: a promoter suitable for driving expression preferentially in roots having a nucleotide sequence of from 1 to 1388 of SEQ ID NO: 44; a nucleic acid heterologous to the promoter, wherein the promoter is operatively coupled 5' to the nucleic acid to induce transcription of the nucleic acid; and a terminator having a nucleotide sequence of from nucleotide 3168 to 4016 of SEQ ID NO: 44, wherein the terminator is operably coupled 3' to the nucleic acid.

111. The method according to claim 110, wherein the plant is a crop plant.

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112. The method according to claim 111, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

113. The method according to claim 111, wherein the plant is an ornamental plant.

114. The method according to claim 113, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

115. A method of altering plant floral morphology in ornamental plants comprising:

transforming an ornamental plant with a nucleic acid molecule according to claim 1.

116. The method according to claim 115, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

117. The method according to claim 115, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

118. The method according to claim 115, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

119. The method according to claim 115, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

120. The method according to claim 115, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

121. The method according to claim 115, wherein the plant is an ornamental plant.

122. The method according to claim 121, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

123. A method of producing plants with a cytoplasmic male sterile plant restoration system comprising:

transforming a first plant in its chloroplast genome with a nucleic acid which causes the plant to become male sterile;

transforming a second plant with a nucleic acid molecule according to claim 1 whose protein product is targeted to the chloroplast; and crossing the first and second plants to produce progeny plants possessing a cytoplasmic male sterile plant restoration system.

124. The method according to claim 123, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

125. The method according to claim 123, wherein the nucleic acid molecule encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

126. The method according to claim 123, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

127. The method according to claim 123, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

128. The method according to claim 123, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

129. A method of producing plants with a cytoplasmic male sterile plant restoration system comprising:

mutagenizing a first plant having a nucleic acid which encodes a protein comprising a motif having an amino acid sequence corresponding to any of

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SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input;

crossing the mutagenized first plant with a wild-type plant having mitochondrial DNA polymorphisms compared to mitochondrial DNA in the mutagenized first plant to produce progeny plants; and

determining if the progeny plants are fertile, whereby fertile progeny plants can be used as a fertile maintainer line, wherein the mutagenized first plant, the fertile maintainer line, and a wild-type allele present in the first plant before mutagenesis comprises a new cytoplasmic male sterile plant restoration system.

130. An isolated nucleic acid sequence corresponding to SEQ ID NO: 42 or SEQ ID NO: 44.

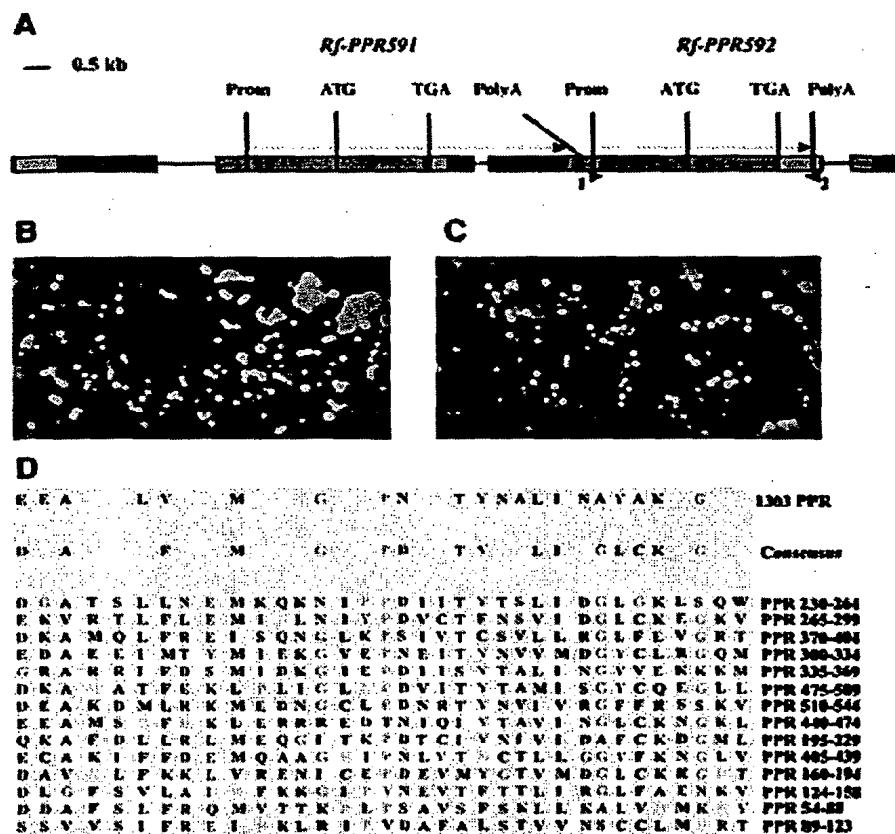
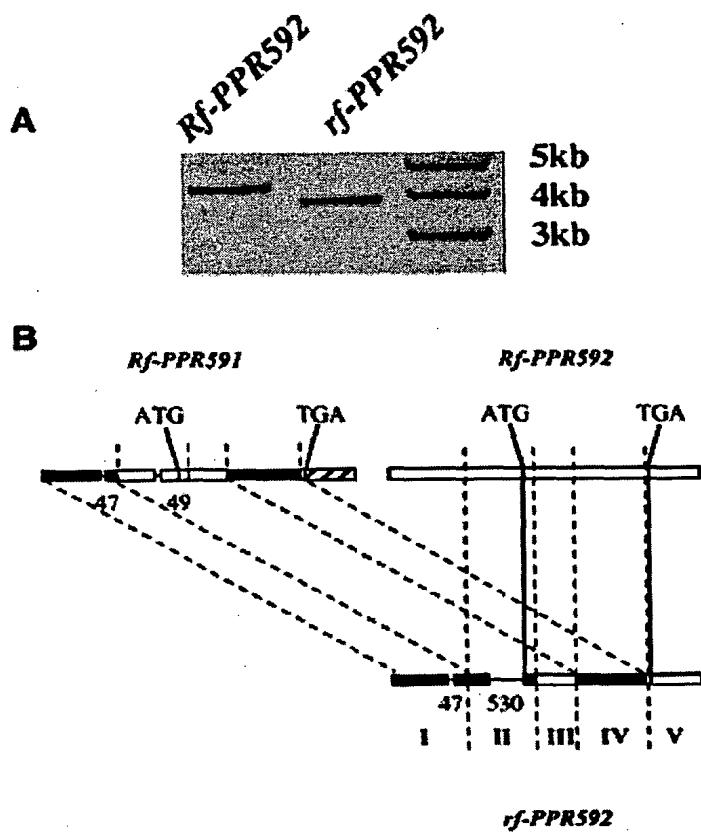
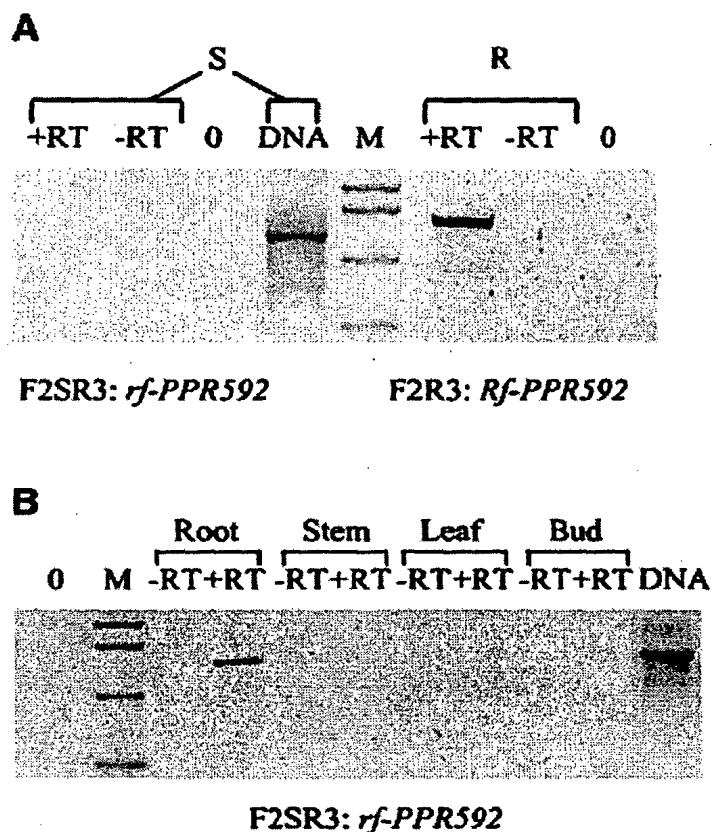


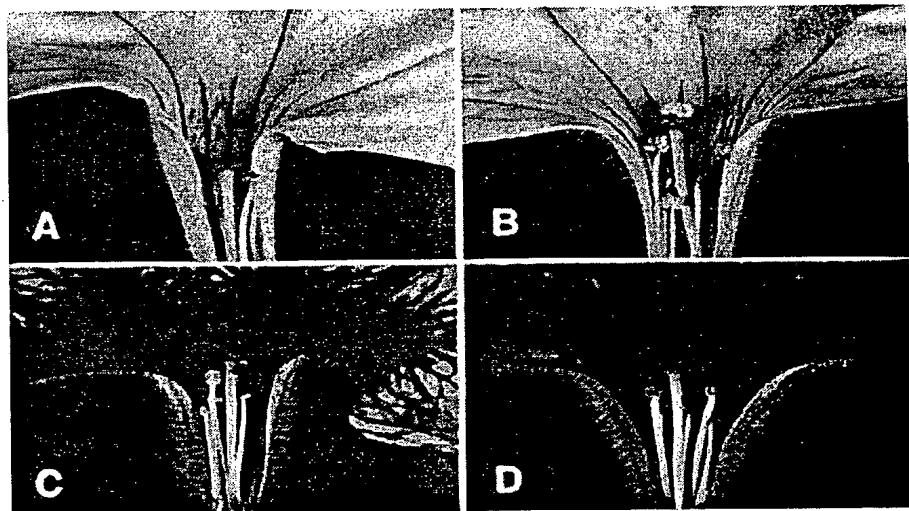
Figure 1



**Figure 2**



**Figure 3**

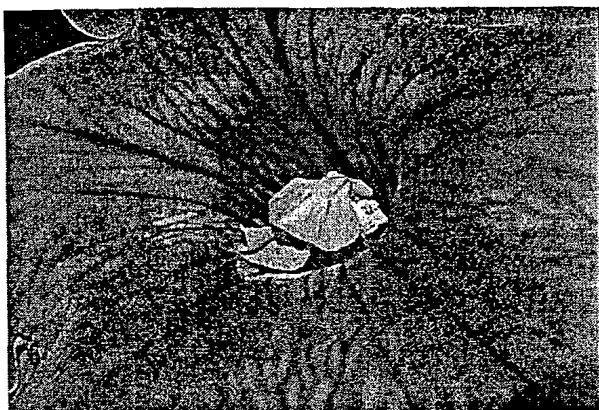


**Figure 4**



**Figure 5**

A.

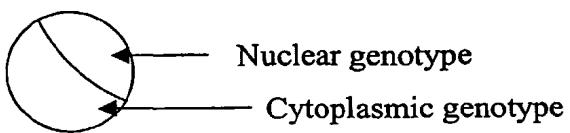


B.



**Figure 6**

## Diagram of a plant genotype



Wild-type plant with wild-type nuclear alleles of gene Ms, a PPR motif gene needed for male fertility



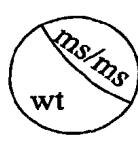
Mutagenesis of the Ms gene (e.g. by an insertional element, radiation, chemicals, etc.)



Heterozygous mutant at Ms locus



Self-cross



Mutant male sterile plant



By sexual crosses or cybridization with genotypes carrying different organelle genomes than the initial wt genome, or by mutagenesis of genome(s) in the wt cytoplasm, create a genotype with mutant ms/ms nuclear alleles that is male fertile



Male fertile plant

These genotypes are then utilized as a CMS/restorer system for hybrid seed production and breeding as follows:



CMS line



Fertile maintainer line



Fertility restorer lines

**Figure 7A**

Examples illustrating how new lines can be used as a CMS/restorer system

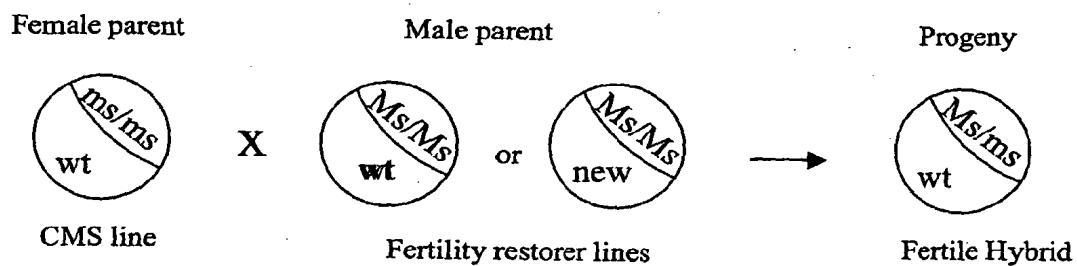
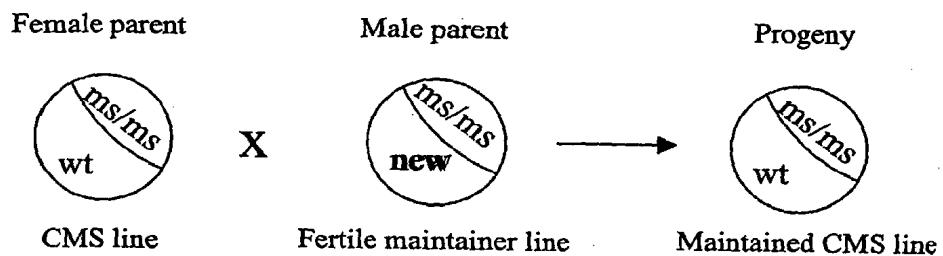
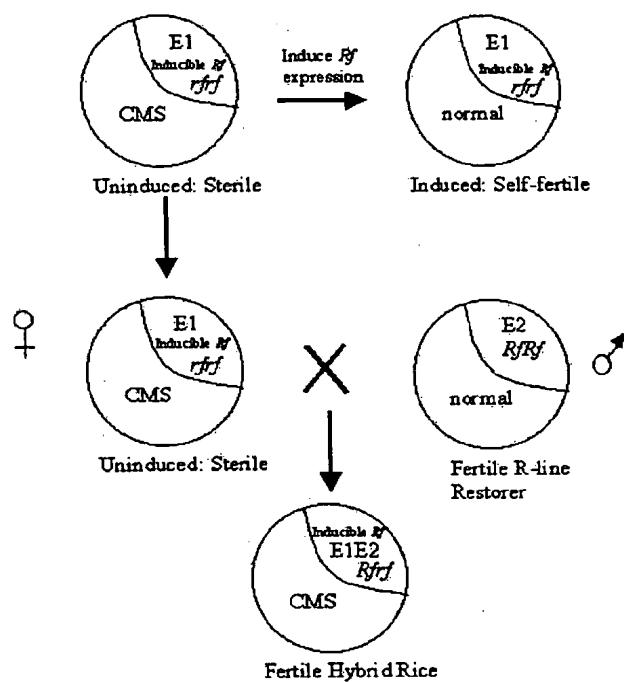


Figure 7B



**Figure 8**

## SEQUENCE LISTING

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<120> GENES FOR ALTERING MITOCHONDRIAL FUNCTION AND FOR  
HYBRID SEED PRODUCTION

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<170> PatentIn Ver. 2.1

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 <211> 592  
 <212> PRT  
 <213> Petunia sp.

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 Cys Ser Ile Ser Val Lys Gly Asn Phe Gly Val Ser Asn Glu Phe Glu  
 35 40 45  
 Asn Val Lys Cys Leu Asp Asp Ala Phe Ser Leu Phe Arg Gln Met Val  
 50 55 60  
 Thr Thr Lys Pro Leu Pro Ser Ala Val Ser Phe Ser Lys Leu Leu Lys  
 65 70 75 80  
 Ala Leu Val His Met Lys His Tyr Ser Ser Val Val Ser Ile Phe Arg  
 85 90 95  
 Glu Ile His Lys Leu Arg Ile Pro Val Asp Ala Phe Ala Leu Ser Thr  
 100 105 110  
 Val Val Asn Ser Cys Cys Leu Met His Arg Thr Asp Leu Gly Phe Ser  
 115 120 125  
 Val Leu Ala Ile His Phe Lys Lys Gly Ile Pro Tyr Asn Glu Val Thr  
 130 135 140  
 Phe Thr Thr Leu Ile Arg Gly Leu Phe Ala Glu Asn Lys Val Lys Asp  
 145 150 155 160  
 Ala Val His Leu Phe Lys Lys Leu Val Arg Glu Asn Ile Cys Glu Pro  
 165 170 175  
 Asp Glu Val Met Tyr Gly Thr Val Met Asp Gly Leu Cys Lys Lys Gly

180	185	190
His Thr Gln Lys Ala Phe Asp Leu Leu Arg Leu Met Glu Gln Gly Ile		
195	200	205
Thr Lys Pro Asp Thr Cys Ile Tyr Asn Ile Val Ile Asp Ala Phe Cys		
210	215	220
Lys Asp Gly Met Leu Asp Gly Ala Thr Ser Leu Leu Asn Glu Met Lys		
225	230	235
Gln Lys Asn Ile Pro Pro Asp Ile Ile Thr Tyr Thr Ser Leu Ile Asp		
245	250	255
Gly Leu Gly Lys Leu Ser Gln Trp Glu Lys Val Arg Thr Leu Phe Leu		
260	265	270
Glu Met Ile His Leu Asn Ile Tyr Pro Asp Val Cys Thr Phe Asn Ser		
275	280	285
Val Ile Asp Gly Leu Cys Lys Glu Gly Lys Val Glu Asp Ala Glu Glu		
290	295	300
Ile Met Thr Tyr Met Ile Glu Lys Gly Val Glu Pro Asn Glu Ile Thr		
305	310	315
320		
Tyr Asn Val Val Met Asp Gly Tyr Cys Leu Arg Gly Gln Met Gly Arg		
325	330	335
Ala Arg Arg Ile Phe Asp Ser Met Ile Asp Lys Gly Ile Glu Pro Asp		
340	345	350
Ile Ile Ser Tyr Thr Ala Leu Ile Asn Gly Tyr Val Glu Lys Lys Lys		
355	360	365
Met Asp Lys Ala Met Gln Leu Phe Arg Glu Ile Ser Gln Asn Gly Leu		
370	375	380
Lys Pro Ser Ile Val Thr Cys Ser Val Leu Leu Arg Gly Leu Phe Glu		
385	390	395
400		
Val Gly Arg Thr Glu Cys Ala Lys Ile Phe Phe Asp Glu Met Gln Ala		
405	410	415
Ala Gly His Ile Pro Asn Leu Tyr Thr His Cys Thr Leu Leu Gly Gly		
420	425	430
Tyr Phe Lys Asn Gly Leu Val Glu Glu Ala Met Ser His Phe His Lys		

435 440 445

Leu Glu Arg Arg Arg Glu Asp Thr Asn Ile Gln Ile Tyr Thr Ala Val  
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Ile Asn Gly Leu Cys Lys Asn Gly Lys Leu Asp Lys Ala His Ala Thr  
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Phe Glu Lys Leu Pro Leu Ile Gly Leu His Pro Asp Val Ile Thr Tyr  
485 490 495

Thr Ala Met Ile Ser Gly Tyr Cys Gln Glu Gly Leu Leu Asp Glu Ala  
500 505 510

Lys Asp Met Leu Arg Lys Met Glu Asp Asn Gly Cys Leu Pro Asp Asn  
515 520 525

Arg Thr Tyr Asn Val Ile Val Arg Gly Phe Phe Arg Ser Ser Lys Val  
530 535 540

Ser Glu Met Lys Ala Phe Leu Lys Glu Ile Ala Gly Lys Ser Phe Ser  
545 550 555 560

Phe Glu Ala Ala Thr Val Glu Leu Leu Met Asp Ile Ile Ala Glu Asp  
565 570 575

Pro Ser Leu Leu Asn Met Ile Pro Glu Phe His Arg Asp Asn Lys Lys  
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<220>  
<223> Description of Artificial Sequence: Consensus  
motif derived from 1,303 PPRs reported by Small &  
Peeters

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Glu Glu Ala Leu Tyr Met Gly Pro Asn Thr Tyr Asn Ala Leu Ile Asn  
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Ala Tyr Ala Lys Gly

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<212> PRT  
<213> Artificial Sequence

<220>  
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motif derived from the 14 PPRs found in Rf-PPR592

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Asp Asp Ala Phe Ser Leu Phe Arg Gln Met Val Thr Thr Lys Pro Leu  
1 5 10 15

Pro Ser Ala Val Ser Phe Ser Lys Leu Leu Lys Ala Leu Val His Met  
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Lys His Tyr  
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<211> 35  
<212> PRT  
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Val Asp Ala Phe Ala Leu Ser Thr Val Val Asn Ser Cys Cys Leu Met  
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His Arg Thr  
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<210> 7  
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<212> PRT  
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Asp Leu Gly Phe Ser Val Leu Ala Ile His Phe Lys Lys Gly Ile Pro  
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Tyr Asn Glu Val Thr Phe Thr Thr Leu Ile Arg Gly Leu Phe Ala Glu  
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Asn Lys Val  
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<210> 8  
<211> 35  
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Pro Asp Glu Val Met Tyr Gly Thr Val Met Asp Gly Leu Cys Lys Lys  
20 25 30

Gly His Thr  
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Gln Lys Ala Phe Asp Leu Leu Arg Leu Met Glu Gln Gly Ile Thr Lys  
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Pro Asp Thr Cys Ile Tyr Asn Ile Val Ile Asp Ala Phe Cys Lys Asp  
20 25 30

Gly Met Leu  
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<210> 10  
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20 25 30

Ser Gln Trp  
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Pro Asp Val Cys Thr Phe Asn Ser Val Ile Asp Gly Leu Cys Lys Glu  
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Gly Lys Val  
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1 5 10 15

Pro Asn Glu Ile Thr Tyr Asn Val Val Met Asp Gly Tyr Cys Leu Arg  
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Gly Gln Met  
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<210> 13  
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<400> 13  
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1 5 10 15

Pro Asp Ile Ile Ser Tyr Thr Ala Leu Ile Asn Gly Tyr Val Glu Lys  
20 25 30

Lys Lys Met  
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<210> 14  
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<212> PRT  
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<400> 14  
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20 25 30

Gly Arg Thr  
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<210> 15  
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<213> Petunia sp.

<400> 15  
Glu Cys Ala Lys Ile Phe Phe Asp Glu Met Gln Ala Ala Gly His Ile  
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Gly Leu Val  
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<210> 16  
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<400> 16  
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Thr Asn Ile Gln Ile Tyr Thr Ala Val Ile Asn Gly Leu Cys Lys Asn  
20 25 30

Gly Lys Leu  
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<210> 17  
<211> 35  
<212> PRT  
<213> Petunia sp.

<400> 17  
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Pro Asp Val Ile Thr Tyr Thr Ala Met Ile Ser Gly Tyr Cys Gln Glu  
20 25 30

Gly Leu Leu  
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<210> 18  
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<212> PRT  
<213> Petunia sp.

<400> 18  
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Ser Lys Val  
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 <213> Oryza sativa

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<212> PRT

<213> Oryza sativa

<400> 21

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 35 40 45

Arg Gly Arg Gly Ala Ser Ile Tyr Gly Leu Asn Arg Ala Leu Ala Asp  
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Val Ala Arg His Ser Pro Ala Ala Ala Val Ser Arg Tyr Asn Arg Met  
 65 70 75 80

Ala Arg Ala Gly Ala Gly Lys Val Thr Pro Thr Val His Thr Tyr Ala  
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Ile Leu Ile Gly Cys Cys Cys Arg Ala Gly Arg Leu Asp Leu Gly Phe  
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Ala Ala Leu Gly Asn Val Val Lys Lys Gly Phe Arg Val Asp Ala Ile  
 115 120 125

Thr Phe Thr Pro Leu Leu Lys Gly Leu Cys Ala Asp Lys Arg Thr Ser  
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Asp Ala Met Asp Ile Val Leu Arg Arg Met Thr Glu Leu Gly Cys Ile  
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Pro Asp Val Phe Ser Tyr Asn Asn Leu Leu Lys Gly Leu Cys Asp Glu  
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Asn Arg Ser Gln Glu Ala Leu Glu Leu Leu His Met Met Ala Asp Asp  
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Arg Gly Gly Ser Pro Pro Asp Val Val Ser Tyr Asn Thr Val Leu  
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Asn Gly Phe Phe Lys Glu Gly Asp Ser Asp Lys Ala Tyr Ser Thr Tyr  
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Ser Ile Ile Ala Ala Leu Cys Lys Ala Gln Ala Met Asp Lys Ala Met  
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Glu Val Leu Asn Thr Met Val Lys Asn Gly Val Met Pro Asp Cys Met  
 260 265 270

Thr Tyr Asn Ser Ile Leu His Gly Tyr Cys Ser Ser Gly Gln Pro Lys  
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Glu Ala Ile Gly Thr Leu Lys Lys Met Arg Ser Asp Gly Val Glu Pro  
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Asn Val Val Thr Tyr Ser Ser Leu Met Asn Tyr Leu Cys Lys Asn Gly  
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Arg Ser Thr Glu Ala Arg Lys Ile Phe Asp Ser Met Thr Lys Arg Gly  
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Thr Lys Gly Ala Leu Val Glu Met His Ala Leu Leu Asp Leu Met Val  
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Val Ile Asp Val Leu Cys Lys Ser Gly Ser Val Asp Asp Ala Met Leu  
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Tyr Thr Ser Leu Ile His Gly Leu Cys Thr Cys Asp Lys Trp Asp Lys  
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Val Ile Glu Ser Glu Lys Leu Phe Asp Leu Met Val Arg Ile Gly Val  
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Ala Gly Lys Met Asp Glu Ala Thr Lys Leu Leu Ala Ser Met Val Ser  
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Tyr Cys Arg Val Ser Arg Met Asp Asp Ala Leu Ala Leu Phe Lys Glu  
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Leu Gln Gly Leu Phe His Thr Arg Arg Thr Ala Ala Ala Lys Glu Leu  
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Tyr Val Ser Ile Thr Lys Ser Gly Thr Gln Leu Glu Leu Ser Thr Tyr  
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Leu Arg Met Phe Gln Asn Leu Cys Leu Thr Asp Leu Gln Leu Glu Thr  
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Asp Glu Ala Lys Asp Leu Phe Ala Ala His Ser Ala Asn Gly Leu Val  
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Leu Gln Arg Gly Asp Ile Thr Arg Ala Gly Thr Tyr Leu Phe Met Ile  
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Asp Glu Lys His Phe Ser Leu Glu Ala Ser Thr Ala Ser Phe Leu Leu  
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Glu Ser Ser Pro Ile Val Trp Glu Gln Ile Ser Arg Ile Ser His Leu  
 770 775 780

Ser Val Asn Leu Lys Leu Ile Lys Gln Pro Lys Cys Thr Cys Glu Leu  
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Gly Pro Lys Trp Ser Gln Asn Leu Pro Lys Pro Gly Thr Asn Ser Val  
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Gly Ser Val Ala Gln Phe His Leu Ser Arg Gly Gly Tyr Arg Ala Tyr  
 820 825 830

Arg Gly Gly Thr Thr Val Thr Ala Leu Pro Gln Gly Asp Gly Asn Pro  
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Gly Pro Asn Asp Lys Val Asn Pro Gly Arg Thr Asn Leu Ala Gln Asn  
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<211> 3660

<212> DNA

<213> Oryza sativa

<400> 22

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<211> 1219

<212> PRT

<213> Oryza sativa

<400> 23

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 35 40 45

Gly Arg Gly Ala Ser Ile Tyr Gly Leu Asn Cys Ala Leu Ala Asp Val  
 50 55 60

Ala Arg His Ser Pro Ala Ala Val Ser Arg Tyr Asn Arg Met Ala  
 65 70 75 80

Arg Ala Gly Ala Asp Glu Val Thr Pro Asn Leu Cys Thr Tyr Gly Ile  
 85 90 95

Leu Ile Gly Ser Cys Cys Cys Ala Gly Arg Leu Asp Leu Gly Phe Ala  
 100 105 110

Ala Leu Gly Asn Val Ile Lys Lys Gly Phe Arg Val Asp Ala Ile Ala  
 115 120 125

Phe Thr Pro Leu Leu Lys Gly Leu Cys Ala Asp Lys Arg Thr Ser Asp  
 130 135 140

Ala Met Asp Ile Val Leu Arg Arg Met Thr Gln Leu Gly Cys Ile Pro  
 145 150 155 160

Asn Val Phe Ser Tyr Asn Ile Leu Leu Lys Gly Leu Cys Asp Glu Asn

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195	200	205
Phe Phe Lys Glu Gly Asp Leu Asp Lys Ala Tyr Gly Thr Tyr His Glu		
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Ile Ala Ala Leu Cys Lys Ala Gln Ala Met Asp Lys Ala Met Glu Val		
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Thr Glu Ala Arg Lys Met Phe Asp Ser Met Thr Lys Arg Gly Leu Lys		
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Pro Glu Ile Thr Thr Tyr Gly Thr Leu Leu Gln Gly Tyr Ala Thr Lys		
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Gly Ala Leu Val Glu Met His Gly Leu Leu Asp Leu Met Val Arg Asn		
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Gly Ile His Pro Asn His Tyr Val Phe Ser Ile Leu Ile Cys Ala Tyr		
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Ala Lys Gln Gly Lys Val Asp Gln Ala Met Leu Val Phe Ser Lys Met		
385	390	395
Arg Gln Gln Gly Leu Asn Pro Asp Thr Val Thr Tyr Gly Thr Val Ile		
405	410	415
Gly Ile Leu Cys Lys Ser Gly Arg Val Glu Asp Ala Met Arg Tyr Phe		

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Glu Gln Met Ile Asp Glu Arg Leu Ser Pro Gly Asn Ile Val Tyr Asn		
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Ser Leu Ile His Ser Leu Cys Ile Phe Asp Lys Trp Asp Lys Ala Lys		
450	455	460
Glu Leu Ile Leu Glu Met Leu Asp Arg Gly Ile Cys Leu Asp Thr Ile		
465	470	475
Phe Phe Asn Ser Ile Ile Asp Ser His Cys Lys Glu Gly Arg Val Ile		
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Glu Ser Glu Lys Leu Phe Asp Leu Met Val Arg Ile Gly Val Lys Pro		
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Lys Met Asp Glu Ala Thr Lys Leu Leu Ala Ser Met Val Ser Val Gly		
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Met Lys Pro Asp Cys Val Thr Tyr Asn Thr Leu Ile Asn Gly Tyr Cys		
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Lys Ile Ser Arg Met Glu Asp Ala Leu Val Leu Phe Arg Glu Met Glu		
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Gly Ile Thr Glu Ser Gly Thr Gln Leu Glu Leu Ser Thr Tyr Asn Ile		
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Ile Leu His Gly Leu Cys Lys Asn Asn Leu Thr Asp Glu Ala Leu Arg		
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Met Phe Gln Asn Leu Cys Leu Thr Asp Leu Gln Leu Glu Thr Arg Thr		
645	650	655
Phe Asn Ile Met Ile Gly Ala Leu Leu Lys Val Gly Arg Asn Asp Glu		
660	665	670
Ala Lys Asp Leu Phe Ala Ala Leu Ser Ala Asn Gly Leu Val Pro Asp		

675	680	685
Val Arg Thr Tyr Ser Leu Met Ala Glu Asn Leu Ile Glu Gln Gly Leu		
690	695	700
Leu Glu Glu Leu Asp Asp Leu Phe Leu Ser Met Glu Glu Asn Gly Cys		
705	710	715
Thr Ala Asn Ser Arg Met Leu Asn Ser Ile Val Arg Lys Leu Leu Gln		
725	730	735
Arg Gly Asp Ile Thr Arg Ala Gly Thr Tyr Leu Phe Met Ile Asp Glu		
740	745	750
Lys His Phe Ser Leu Glu Ala Ser Thr Ala Scr Leu Phe Leu Asp Leu		
755	760	765
Leu Ser Gly Gly Lys Tyr Gln Glu Tyr His Ser Cys Ile Arg Gly Gly		
770	775	780
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785	790	795
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Val Asn Leu Val Asp Ser Lys Ala Pro Ser Ile Gly Ser Lys Leu Leu		
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Gly Ile Ser Lys Val Gln Met Leu Asn Gly Ser Asn Lys Asp Ser Asp		
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Cys Ile Ser Glu Glu Ile Leu Ser Lys Val Glu Glu Ile Leu Leu Ser		
850	855	860
Cys Gln Val Ile Lys Ser Leu Asp Lys Asp Asp Lys Lys Thr Thr Arg		
865	870	875
Pro Glu Leu Cys Pro Lys Trp Leu Ala Leu Leu Thr Met Glu Asn Ala		
885	890	895
Cys Leu Ser Ala Val Ser Val Glu Glu Thr Ser Asp Thr Val Ser Arg		
900	905	910
Val Gly Gly Asn Phe Lys Glu Thr Leu Arg Glu Met Gly Gly Leu Asp		
915	920	925
Ser Ile Phe Asp Val Met Val Asp Phe His Ser Thr Leu Glu Asn Leu		

930	935	940
Ile Lys Asp Thr Ser Thr Ser Ala Leu Asp Arg Asn Glu Gly Thr Ser		
945	950	955
960		
Leu Gln Ser Ala Ala Leu Leu Leu Lys Cys Leu Lys Ile Leu Glu Asn		
965	970	975
Ala Ile Phe Leu Ser Asp Asp Asn Lys Thr His Leu Leu Asn Met Ser		
980	985	990
Arg Lys Leu Asn Pro Lys Arg Ser Leu Leu Ser Phe Val Gly Val Ile		
995	1000	1005
Ile Asn Thr Ile Glu Leu Leu Ser Ala Leu Ser Ile Leu Gln Asn Ser		
1010	1015	1020
Ser Val Val Ser Ser Ser Thr Tyr Pro Lys Ser Ser Lys Val Ser Gln		
1025	1030	1035
1040		
Gln Ser Tyr Ser Val Val Met Ala Gly Gly Asp Arg Gly Arg Gly Val		
1045	1050	1055
Glu Cys His Pro His Gln Gly Val Ser Ala Ala Leu Leu Arg Pro Gly		
1060	1065	1070
Pro Gln Ala Leu Ala Ala Ser Trp Arg Arg Arg Glu Thr Val Val Arg		
1075	1080	1085
Ser Asp Phe Ala Ala Gly Gly Val Ala Thr Met Gly Asp Ser Pro Gln		
1090	1095	1100
Ala Leu Ser Asp Arg Leu Cys Gly Ser Ala Thr Lys Val Trp Arg Gly		
1105	1110	1115
1120		
Gly Ala Glu Trp Thr Ala Glu Ala Phe Ala Arg Asn Gly Ala Ala Gly		
1125	1130	1135
Pro Ser Gln Ser Arg Leu Pro Val Thr Arg Asn Arg Ala Gln His Tyr		
1140	1145	1150
Ile Ala Lys Ile Trp Ala Thr Leu Thr Ile Ser Met Phe Tyr Lys Leu		
1155	1160	1165
Asp Val Glu Gly Met Glu Asn Leu Pro Pro Asn Ser Ser Pro Ala Ile		
1170	1175	1180
Tyr Val Ala Asn His Gln Ser Phe Leu Asp Ile Tyr Thr Leu Leu Thr		

1185 1190 1195 1200  
Leu Gly Arg Cys Phe Lys Phe Ile Ser Lys Thr Ser Ile Phe Met Phe  
1205 1210 1215  
Arg Ile Ile

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 actgctgcct gctgggctct agccgcattc agttataacc gtacaaactt cagtgatttg 2160  
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 aatctcagga ccggccctgc tcatgatcct tacaccgtgt atcctgtaga gtacttctct 2280  
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<210> 25

<211> 952

<212> PRT

<213> Oryza sativa

<400> 25

Met Ala Arg Arg Ala Ala Ser Arg Ala Val Gly Ala Leu Arg Ser Asp  
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Gly Ser Ile Gln Gly Arg Gly Gly Arg Ala Gly Gly Ser Gly Ala Glu  
 20 25 30

Asp Ala Arg His Val Phe Asp Glu Leu Leu Arg Arg Gly Arg Gly Ala  
 35 40 45

Ser Ile Tyr Gly Leu Asn Arg Ala Leu Ala Asp Val Ala Arg His Ser  
 50 55 60

Pro Ala Ala Ala Val Ser Arg Tyr Asn Arg Met Ala Arg Ala Gly Ala  
 65 70 75 80

Asp Glu Val Thr Pro Asp Leu Cys Thr Tyr Gly Ile Leu Ile Gly Cys  
 85 90 95

Cys Cys Arg Ala Gly Arg Leu Asp Leu Gly Phe Ala Ala Leu Gly Asn  
 100 105 110

Val Ile Lys Lys Gly Phe Arg Val Glu Ala Ile Thr Phe Thr Pro Leu  
 115 120 125

Leu Lys Gly Leu Cys Ala Asp Lys Arg Thr Ser Asp Ala Met Asp Ile

130	135	140
Val Leu Arg Arg Met Thr Glu Leu Gly Cys Ile Pro Asn Val Phe Ser		
145	150	155
Tyr Asn Asn Leu Leu Asn Gly Leu Cys Asp Glu Asn Arg Ser Gln Glu		
165	170	175
Ala Leu Glu Leu Leu His Met Met Ala Asp Asp Arg Gly Gly Ser		
180	185	190
Pro Pro Asp Val Val Ser Tyr Thr Thr Val Ile Asn Gly Phe Phe Lys		
195	200	205
Glu Gly Asp Ser Asp Lys Ala Tyr Ser Thr Tyr His Glu Met Leu Asp		
210	215	220
Arg Gly Ile Leu Pro Asp Val Val Thr Tyr Ser Ser Ile Ile Ala Ala		
225	230	235
Leu Cys Lys Gly Gln Ala Met Asp Lys Pro Trp Ser His Cys Lys Glu		
245	250	255
Gly Arg Val Ile Glu Ser Glu Lys Leu Phe Asp Leu Met Val Arg Ile		
260	265	270
Gly Val Lys Pro Asp Ile Ile Thr Tyr Ser Thr Leu Ile Asp Gly Tyr		
275	280	285
Cys Leu Ala Gly Lys Met Asp Glu Ala Met Lys Leu Leu Ser Gly Met		
290	295	300
Val Ser Val Gly Leu Lys Pro Asn Thr Val Thr Tyr Ser Thr Leu Ile		
305	310	315
Asn Gly Tyr Cys Lys Ile Ser Arg Met Glu Asp Ala Leu Val Leu Phe		
325	330	335
Lys Glu Met Glu Ser Ser Gly Val Ser Pro Asp Ile Ile Thr Tyr Asn		
340	345	350
Ile Ile Leu Gln Gly Leu Phe Gln Thr Arg Arg Thr Ala Ala Ala Lys		
355	360	365
Glu Leu Tyr Val Arg Ile Thr Glu Ser Gly Thr Gln Ile Glu Leu Ser		
370	375	380
Thr Tyr Asn Ile Ile Leu His Gly Leu Cys Lys Asn Lys Leu Thr Asp		

385	390	395	400
Asp Ala Leu Gln Met Phe Gln Asn Leu Cys Leu Met Asp Leu Lys Leu			
405	410	415	
Glu Ala Arg Thr Phe Asn Ile Met Ile Asp Ala Leu Leu Lys Val Gly			
420	425	430	
Arg Asn Asp Glu Ala Lys Asp Leu Phe Val Ala Phe Ser Ser Asn Gly			
435	440	445	
Leu Val Pro Asn Tyr Trp Thr Tyr Arg Leu Met Ala Glu Asn Ile Ile			
450	455	460	
Gly Gln Gly Leu Leu Glu Glu Leu Asp Gln Leu Phe Leu Ser Met Glu			
465	470	475	480
Asp Asn Gly Cys Thr Val Asp Ser Gly Met Leu Asn Phe Ile Val Arg			
485	490	495	
Glu Leu Leu Gln Arg Gly Val Val Val Val Val Ser Gly Glu Ser Ala			
500	505	510	
Thr Thr Pro Pro Pro Thr Leu Lys Ile Leu Thr Cys Gly Ile Thr Val			
515	520	525	
Asn Pro Phe Ser Lys Thr Cys Gly Ile Thr Val Asn Pro Phe Ser Lys			
530	535	540	
Pro Ile Val Gln Thr Gly Ala Cys Gly Gln Val Lys Glu Val Gly Lys			
545	550	555	560
Asn Ala Ser Glu Glu Arg Leu Ile Val Val Ser Ser Gln Glu Ile Pro			
565	570	575	
Asp Asp Pro Val Ser Pro Thr Ile Glu Ala Leu Ile Leu Leu His Ser			
580	585	590	
Lys Ala Ser Thr Leu Ala Glu Asn His Gln Leu Thr Thr Arg Leu Val			
595	600	605	
Val Pro Ser Asn Lys Val Gly Cys Ile Leu Gly Glu Gly Lys Val			
610	615	620	
Ile Thr Glu Met Arg Arg Arg Thr Gly Ala Glu Ile Arg Val Tyr Ser			
625	630	635	640
Lys Ala Asp Lys Pro Lys Tyr Leu Ser Phe Asp Glu Glu Leu Val Gln			

645	650	655
His Ile Ser Leu Ile Leu Val Asp Arg His Ala Gly Arg Ala His Leu		
660	665	670
Leu Ser His Gln Leu Leu Thr Ala Ile Tyr Val Leu Val Leu Asn Arg		
675	680	685
Ser Ile Val Val Ala Glu Val Lys Asn Asn His Gly Thr Ala Ala Cys		
690	695	700
Trp Ala Leu Ala Ala Ile Ser Tyr Asn Arg Thr Asn Phe Ser Asp Leu		
705	710	715
Leu Val Ser His Trp Phe Ile Ile Lys Ala Ser Val Phe Ser Phe Thr		
725	730	735
Leu Gly Leu Gln Asn Leu Arg Thr Gly Pro Ala His Asp Pro Tyr Thr		
740	745	750
Val Tyr Pro Val Glu Tyr Phe Ser Lys Arg Glu Tyr Pro Ser Gly Ser		
755	760	765
Ser Lys Val Ala Pro Ser Ala Ser Tyr Glu Arg Tyr Ala Ala Thr Thr		
770	775	780
Arg Leu Pro Asn Gly Glu Leu Pro Ser Ser Ile Ser Pro Gly Ala Asp		
785	790	795
Tyr Met Ser Cys Arg Ser Tyr Leu Asp Gln Val Pro Thr Asp Arg Tyr		
805	810	815
Ser Asn Arg Val Thr Leu Gln Leu Gly Leu Ser Arg Ala Gly Asn Ser		
820	825	830
Asn Val Gln Gln Leu Gly Ile Thr Arg Ala Gly Asn Ser Asn Ala Tyr		
835	840	845
Asp Tyr Thr Glu Ala Ala Glu Gln Ile His Gly Arg Glu Asp Tyr Arg		
850	855	860
Arg Leu Ser Gly Leu Thr Gly Tyr Pro Gly Gly Ser Ser Asn Cys Gly		
865	870	875
Phe Gln Ile Val Asn Trp Ser Leu Ser Leu Val Leu Val Ile Ser Gly		
885	890	895
Ala Arg Val Lys Leu His Glu Ala His Pro Gly Ser Ser Glu Ser Ile		

900	905	910
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Val Glu Ile Gln Gly Ile Pro Asp Gln Val Lys Ala Ala Gln Ser Leu		
915	920	925

Leu Gln Gly Phe Ile Gly Ala Ser Ser Asn Ser Arg Gln Ala Pro Gln		
930	935	940

Ser Ser Arg Met Ala His Tyr Phe		
945	950	

<210> 26

<211> 1737

<212> DNA

<213> Oryza sativa

<400> 26

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<210> 27  
<211> 578  
<212> PRT  
<213> Oryza sativa

<400> 27  
Met Pro Leu Ala Thr Leu Leu Gly His Leu Ala Ala Gly Arg Phe Gly  
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Leu Val Gln Ala Leu Thr Gly Ala Ala Thr Ala Ala Ala His Arg  
20 25 30

Leu Leu His Leu Leu Leu Arg Thr Ala Pro Pro Pro Pro Leu Pro Asp  
35 40 45

Leu Val Ser Leu Ala Arg Trp Ser Arg Ala His Phe Arg Ala Pro Leu  
50 55 60

Pro Leu Arg Leu His Gly Leu Leu Leu Ala Arg Leu Ala Ser Lys Gly  
65 70 75 80

Leu Tyr Pro Leu Leu Arg Ser Glu Leu His Val Leu Ala Ala Ala Arg  
85 90 95

Leu His Ser Pro Ala Ser Ile Leu Arg Ala Leu Pro Ser Pro Ser Ala  
100 105 110

Ser Ala Ser Ala Ser Thr Pro Leu Ile Ala Asp Met Leu Val Leu Ala  
115 120 125

Leu Ala Arg Ala Ser Gln Pro Leu Arg Ala Tyr Asp Ala Phe Leu Leu  
130 135 140

Ala Gly Glu Ser His Pro Arg His Arg Pro Ser Thr Ser Ser Val Asn  
145 150 155 160

Ala Leu Leu Ala Gly Leu Val Gly Ala Lys Arg Val Asp Leu Ala Glu  
165 170 175

Lys Ala Phe Arg Ser Ala Leu Arg Arg Arg Val Ser Pro Asp Ile Tyr  
180 185 190

Thr Phe Asn Thr Val Ile Ser Gly Leu Cys Arg Ile Gly Gln Leu Arg  
195 200 205

Lys Ala Gly Asp Val Ala Lys Asp Ile Lys Ala Trp Gly Leu Ala Pro  
210 215 220

Ser Val Ala Thr Tyr Asn Ser Leu Ile Asp Gly Tyr Cys Lys Lys Gly  
225 230 235 240

Gly Ala Gly Asn Met Tyr His Val Asp Met Leu Leu Lys Glu Met Val  
245 250 255

Glu Ala Gly Ile Ser Pro Thr Ala Val Thr Phe Gly Val Leu Ile Asn  
260 265 270

Gly Tyr Cys Lys Asn Ser Asn Thr Ala Ala Ala Val Arg Val Phe Glu  
275 280 285

Glu Met Lys Gln Gln Gly Ile Ala Ala Ser Val Val Thr Tyr Asn Ser  
290 295 300

Leu Ile Ser Gly Leu Cys Ser Glu Gly Lys Val Glu Glu Gly Val Lys  
305 310 315 320

Leu Met Glu Glu Met Glu Asp Leu Gly Leu Ser Pro Asn Glu Ile Thr  
325 330 335

Phe Gly Cys Val Leu Lys Gly Phe Cys Lys Lys Gly Met Met Ala Asp  
340 345 350

Ala Asn Asp Trp Ile Asp Gly Met Thr Glu Arg Asn Val Glu Pro Asp  
355 360 365

Val Val Ile Tyr Asn Ile Leu Ile Asp Val Tyr Arg Arg Leu Gly Lys  
370 375 380

Met Glu Asp Ala Met Ala Val Lys Glu Ala Met Ala Lys Lys Gly Ile  
385 390 395 400

Ser Pro Asn Val Thr Thr Tyr Asn Cys Leu Ile Thr Gly Phe Ser Arg  
405 410 415

Ser Gly Asp Trp Arg Ser Ala Ser Gly Leu Leu Asp Glu Met Lys Glu  
420 425 430

Lys Gly Ile Glu Ala Asp Val Val Thr Tyr Asn Val Leu Ile Gly Ala  
435 440 445

Leu Cys Cys Lys Gly Glu Val Arg Lys Ala Val Lys Leu Leu Asp Glu  
450 455 460

Met Ser Glu Val Gly Leu Glu Pro Asn His Leu Thr Tyr Asn Thr Ile  
465 470 475 480

Ile Gln Gly Phe Cys Asp Lys Gly Asn Ile Lys Ser Ala Tyr Glu Ile  
 485 490 495

Arg Thr Arg Met Glu Lys Cys Arg Lys Arg Ala Asn Val Val Thr Tyr  
 500 505 510

Asn Val Phe Ile Lys Tyr Phe Cys Gln Ile Gly Lys Met Asp Glu Ala  
 515 520 525

Asn Asp Leu Leu Asn Glu Met Leu Asp Lys Cys Leu Val Pro Asn Gly  
 530 535 540

Ile Thr Tyr Glu Thr Ile Lys Glu Gly Met Met Glu Lys Gly Tyr Thr  
 545 550 555 560

Pro Asp Ile Arg Gly Cys Thr Val Ser Gln Ala Ser Glu Asn Pro Ala  
 565 570 575

Ser Ser

<210> 28

<211> 1365

<212> DNA

<213> Oryza sativa

<400> 28

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 aatagtattt tgcattggata ttgttcttca ggacagtgcgg aaaaggctat tggtattttc 300  
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 gatcatcatg tcttcaacat attaatatgt gcatacacta aacaagaaaa agtagacgag 600  
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aaggagatgg aaagcaatgg tgttaatcct gatattatta catataacat aattctgcat 1200  
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<210> 29

<211> 454

<212> PRT

<213> Oryza sativa

<400> 29

Met Ala Asp Asp Gly Arg Cys Pro Pro Asp Val Val Ser Tyr Asn Thr  
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Ile Ile Asp Gly Leu Phe Lys Glu Gly Asp Val Asp Lys Ala Tyr Ile  
 20 25 30

Thr Tyr His Glu Met Leu Asp Arg Arg Val Ser Pro Asp Ala Val Thr  
 35 40 45

Tyr Asn Ser Ile Ile Ala Ala Leu Ser Lys Ala Gln Ala Met Asp Arg  
 50 55 60

Ala Met Glu Val Leu Thr Val Met Val Met Pro Asn Cys Phe Thr Tyr  
 65 70 75 80

Asn Ser Ile Met His Gly Tyr Cys Ser Ser Gly Gln Ser Glu Lys Ala  
 85 90 95

Ile Gly Ile Phe Arg Lys Met Cys Ser Asp Gly Ile Glu Pro Asp Val  
 100 105 110

Val Thr Tyr Asn Ser Leu Met Asp Tyr Leu Cys Lys Asn Gly Lys Cys  
 115 120 125

Thr Glu Ala Arg Lys Ile Phe Asp Ser Met Val Lys Arg Gly Leu Lys  
 130 135 140

Pro Asp Ile Thr Thr Tyr Gly Thr Leu Leu His Gly Tyr Ala Ser Lys  
 145 150 155 160

Gly Ala Leu Val Glu Met His Asp Leu Leu Ala Leu Met Val Gln Asn  
 165 170 175

Gly Met Gln Leu Asp His His Val Phe Asn Ile Leu Ile Cys Ala Tyr  
 180 185 190

Thr Lys Gln Glu Lys Val Asp Glu Val Val Leu Val Phe Ser Lys Met  
195 200 205

Arg Gln Gln Gly Leu Thr Pro Asn Ala Val Asn Tyr Arg Thr Val Ile  
210 215 220

Asp Gly Leu Cys Lys Leu Gly Arg Leu Asp Asp Ala Met Leu Asn Phe  
225 230 235 240

Glu Gln Met Ile Asp Lys Gly Leu Thr Pro Asn Val Val Val Tyr Thr  
245 250 255

Ser Leu Ile His Ala Leu Cys Thr Tyr Asp Lys Trp Glu Lys Ala Glu  
260 265 270

Glu Leu Ile Phe Glu Ile Leu Asp Gln Gly Ile Asn Pro Asn Ile Val  
275 280 285

Phe Phe Asn Thr Ile Leu Asp Ser Leu Cys Lys Glu Gly Arg Val Ile  
290 295 300

Glu Ser Lys Lys Leu Phe Asp Leu Leu Gly His Ile Gly Val Asn Pro  
305 310 315 320

Asp Val Ile Thr Tyr Ser Thr Leu Ile Asp Gly Tyr Cys Leu Ala Gly  
325 330 335

Lys Met Asp Gly Ala Met Lys Leu Leu Thr Gly Met Val Ser Val Gly  
340 345 350

Leu Lys Pro Asp Ser Val Thr Tyr Ser Thr Leu Ile Asn Gly Tyr Cys  
355 360 365

Lys Ile Asn Arg Met Glu Asp Ala Leu Ala Leu Phe Lys Glu Met Glu  
370 375 380

Ser Asn Gly Val Asn Pro Asp Ile Ile Thr Tyr Asn Ile Ile Leu His  
385 390 395 400

Gly Leu Phe Arg Thr Arg Arg Thr Ala Ala Ala Lys Glu Leu Tyr Ala  
405 410 415

Arg Ile Thr Glu Ser Gly Thr Gln Leu Glu Leu Ser Thr Tyr Asn Ile  
420 425 430

Ile Leu Met Asp Phe Ala Lys Thr Asn Ser Leu Met Met His Phe Gly  
435 440 445

Cys Phe Arg Thr Tyr Val  
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<210> 30

<211> 1386

<212> DNA

<213> Oryza sativa

<400> 30

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cgtggcaagg ggcgcacacat ctacggcttg aaccgcgccc tcgacgacgt cgccgcgtcac 180  
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ggcttcttca aagaggggaa tccggacaaa gcttacgcta cataccatga aatgtttgac 660  
caggggattt tgccagatgt tgtgacttac agctcttattt tcgctgcctt atgcaaggct 720  
caagctatgg acaaggccat ggaggtactt aacaccatgg ttaagaatgg tgtcatgcct 780  
aattgcagga catataatag tattgtgcac ggatattgct cttcaaggca gttgacagag 840  
gctattggat ttctcaaaat gatgtgcagt gatgggtcg aaccagatgt tggtaacttgc 900  
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<210> 31

<211> 461

<212> PRT

<213> Oryza sativa

<400> 31

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Gln Gly Arg Gly Gly Arg Ala Gly Gly Asn Gly Ala Glu Asp Ala Arg  
20 25 30

His Val Phe Asp Glu Leu Leu Arg Arg Gly Lys Gly Ala Thr Ile Tyr  
 35 40 45

Gly Leu Asn Arg Ala Leu Asp Asp Val Ala Arg His Ser Pro Ala Ala  
 50 55 60

Ala Val Ser Arg Tyr Asn Arg Met Ala Arg Ala Gly Ala Asp Glu Val  
 65 70 75 80

Thr Pro Asn Leu Tyr Thr Tyr Ser Val Leu Ile Gly Cys Cys Cys Arg  
 85 90 95

Ala Gly Arg Leu Asp Leu Gly Phe Ala Ala Leu Gly Asn Val Ile Lys  
 100 105 110

Lys Gly Phe Arg Val Glu Ala Ile Thr Phe Thr Pro Leu Leu Lys Gly  
 115 120 125

Leu Cys Ala Asp Lys Arg Thr Ser Asp Ala Met Asp Ile Val Leu Cys  
 130 135 140

Arg Met Thr Gln Leu Gly Cys Ile Pro Asn Val Phe Ser Cys Thr Ile  
 145 150 155 160

Leu Leu Lys Gly Leu Cys Asp Glu Asn Arg Ser Gln Glu Ala Leu Glu  
 165 170 175

Leu Leu Gln Met Met Pro Asp Asp Gly Gly Asp Cys Pro Pro Asp Val  
 180 185 190

Val Leu Tyr Asn Thr Val Ile Asn Gly Phe Phe Lys Glu Gly Asp Pro  
 195 200 205

Asp Lys Ala Tyr Ala Thr Tyr His Glu Met Phe Asp Gln Gly Ile Leu  
 210 215 220

Pro Asp Val Val Thr Tyr Ser Ser Ile Ile Ala Ala Leu Cys Lys Ala  
 225 230 235 240

Gln Ala Met Asp Lys Ala Met Glu Val Leu Asn Thr Met Val Lys Asn  
 245 250 255

Gly Val Met Pro Asn Cys Arg Thr Tyr Asn Ser Ile Val His Gly Tyr  
 260 265 270

Cys Ser Ser Gly Gln Leu Thr Glu Ala Ile Gly Phe Leu Lys Met Met  
 275 280 285

Cys Ser Asp Gly Val Glu Pro Asp Val Val Thr Cys Asn Leu Leu Met  
 290 295 300

Asp Tyr Leu Cys Lys Asn Arg Arg Cys Thr Glu Ala Arg Lys Ile Phe  
 305 310 315 320

Asn Ser Met Thr Lys Cys Gly Leu Lys Pro Asp Ile Thr Thr Tyr Cys  
 325 330 335

Thr Leu Leu Gln Gly Tyr Ala Thr Lys Gly Ala Leu Val Glu Met His  
 340 345 350

Asp Leu Leu Asp Leu Met Val Trp Asn Gly Ile Gln Pro Asn His His  
 355 360 365

Val Phe Asn Ile Leu Ile Cys Ala Tyr Ala Lys Gln Glu Lys Val Asp  
 370 375 380

Glu Ala Met Leu Val Phe Ser Lys Met Arg Gln Gln Gly Leu Ser Pro  
 385 390 395 400

Asn Ala Val Asn Tyr Arg Thr Val Ile Asp Val Leu Cys Lys Leu Gly  
 405 410 415

Arg Val Tyr Asp Ala Val Leu Thr Leu Lys Gln Met Ile Asn Glu Gly  
 420 425 430

Leu Thr Pro Asp Ile Ile Val Tyr Thr Pro Leu Ile His Gly Phe Cys  
 435 440 445

Thr Cys Asp Lys Trp Glu Lys Ala Glu Glu Leu Ile Phe  
 450 455 460

<210> 32

<211> 1521

<212> DNA

<213> Oryza sativa

<400> 32

atggcacgcc gcgtcgctgc ccgcgcggc gcccgcggcc gccccgtccc gcgctcgagg 60  
 ggtacgatcc aagaccgagc acgcgttggg agcggtggcg cccgaggacgc actcgacgtg 120  
 ttcgacgaat tgctccggcg aggcacatggc gctccgatcc gcagcttcaa 180  
 cccgacgtcg cgcgcgacaa ccccgccggcc gctgtgtccc gcttcaaccg catggcacga 240  
 gctgggtgcca gcatggtaac tcccacccgtg cacacctatg gcatcctcat cggctgctgc 300  
 tgcagtgccg gcccgtttaga cctcggttgc gcccgttgg gccatgtcgt taagaaggaa 360  
 ttcagatggc aaccatcat cttaatcct ctgctcaagg gcctctgtgc agacaagagg 420  
 acggacgacg caatggacat agtgctccgt ggaatgacccg agtcagctg cgtgccaaat 480

gtcttctccc acaccattat tctcaaggga ctctgtcatg agaacagaag ccaagaagct 540  
 ctcgagctgc tccacatgtat ggctgtatggat ggaggaggct gcttacctaa tgttgtgtca 600  
 tacagcaccc tcacatcgatgg cctcttgaaa ggaggggatc cggacaaagc ctacgctaca 660  
 taccgtgaaa tgcttgaccg gaggatttg ccaaattgttg tgatttacag ctccattatt 720  
 gctgccctat gcaagggtca agcaatggac aaggccatgg aggtacacga taggatgggt 780  
 aagaatggag ttacacccaa ttgcttcacg tatactagtc ttgtgcatgg attttgctct 840  
 tcagggcagt tgacagaggc tattaaattt ctagaaaaga tgtgcagcaa tgggtgtgaa 900  
 ccaaattgttg ttacttatacg ctcgtttatg gactatctct gcaagaacgg aagatgcaca 960  
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 gtgtcgactc ctgcacatgcta a 1521

&lt;210&gt; 33

&lt;211&gt; 506

&lt;212&gt; PRT

&lt;213&gt; Oryza sativa

&lt;400&gt; 33

Met	Ala	Arg	Arg	Val	Ala	Ala	Arg	Ala	Arg	Ala	Gly	Gly	Val
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Pro	Arg	Ser	Glu	Gly	Thr	Ile	Gln	Asp	Arg	Ala	Arg	Val	Gly	Ser	Gly
						20				25				30	

Gly	Ala	Glu	Asp	Ala	Leu	Asp	Val	Phe	Asp	Glu	Leu	Leu	Arg	Arg	Gly
						35			40				45		

Ile	Gly	Ala	Pro	Ile	Arg	Ser	Leu	Asn	Gly	Ala	Leu	Ala	Asp	Val	Ala
							50		55				60		

Arg	Asp	Asn	Pro	Ala	Ala	Val	Ser	Arg	Phe	Asn	Arg	Met	Ala	Arg	
						65		70		75			80		

Ala	Gly	Ala	Ser	Met	Val	Thr	Pro	Thr	Val	His	Thr	Tyr	Gly	Ile	Leu
						85			90				95		

Ile	Gly	Cys	Cys	Cys	Ser	Ala	Gly	Arg	Leu	Asp	Leu	Gly	Phe	Ala	Ala
						100			105				110		

Leu	Gly	His	Val	Val	Lys	Lys	Gly	Phe	Arg	Val	Glu	Pro	Ile	Ile	Phe
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

115	120	125
Asn Pro Leu Leu Lys Gly Leu Cys Ala Asp Lys Arg Thr Asp Asp Ala		
130	135	140
Met Asp Ile Val Leu Arg Gly Met Thr Glu Leu Ser Cys Val Pro Asn		
145	150	155
160		
Val Phe Ser His Thr Ile Ile Leu Lys Gly Leu Cys His Glu Asn Arg		
165	170	175
Ser Gln Glu Ala Leu Glu Leu Leu His Met Met Ala Asp Asp Gly Gly		
180	185	190
Gly Cys Leu Pro Asn Val Val Ser Tyr Ser Thr Val Ile Asp Gly Leu		
195	200	205
Leu Lys Gly Gly Asp Pro Asp Lys Ala Tyr Ala Thr Tyr Arg Glu Met		
210	215	220
Leu Asp Arg Arg Ile Leu Pro Asn Val Val Ile Tyr Ser Ser Ile Ile		
225	230	235
240		
Ala Ala Leu Cys Lys Gly Gln Ala Met Asp Lys Ala Met Glu Val His		
245	250	255
Asp Arg Met Val Lys Asn Gly Val Thr Pro Asn Cys Phe Thr Tyr Thr		
260	265	270
Ser Leu Val His Gly Phe Cys Ser Ser Gly Gln Leu Thr Glu Ala Ile		
275	280	285
Lys Phe Leu Glu Lys Met Cys Ser Asn Gly Val Glu Pro Asn Val Val		
290	295	300
Thr Tyr Ser Ser Phe Met Asp Tyr Leu Cys Lys Asn Gly Arg Cys Thr		
305	310	315
320		
Glu Ala Arg Lys Ile Phe Asp Ser Met Val Lys Arg Gly Leu Lys Pro		
325	330	335
Asp Ile Thr Thr Tyr Ser Ser Leu Leu His Gly Tyr Ala Ile Glu Gly		
340	345	350
Ala Leu Val Glu Met His Gly Leu Phe Asp Leu Met Val Gln Ser Asp		
355	360	365
Met Gln Pro Asp His Tyr Val Phe Asn Thr Leu Ile Tyr Ala Ser Ala		

370	375	380
Lys Gln Gly Lys Val Asp Glu Ala Met Leu Val Phe Ser Lys Met Arg		
385	390	395
395		
400		
Gln Gln Gly Leu Lys Pro Asn Cys Val Thr Tyr Ser Thr Leu Ile Asn		
405	410	415
415		
Gly Tyr Cys Lys Ile Thr Arg Met Glu Asn Ala Leu Ala Leu Phe Gln		
420	425	430
430		
Glu Met Val Ser Asn Gly Val Ser Pro Asn Phe Ile Thr Tyr Asn Ile		
435	440	445
445		
Met Leu Gln Gly Leu Phe Arg Thr Gly Arg Thr Ala Thr Ala Lys Glu		
450	455	460
460		
Phe Tyr Val Gln Ile Ile Lys Ser Gly Lys Lys Asp Leu Ile Glu Gln		
465	470	475
475		
480		
Gly Leu Leu Glu Glu Leu Asp Asp Leu Phe Leu Ser Met Glu Asp Asn		
485	490	495
495		
Asp Cys Ser Thr Val Ser Thr Pro Ala Cys		
500	505	

<210> 34  
 <211> 1884  
 <212> DNA  
 <213> Oryza sativa

<400> 34  
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 gggcgagggg gccgcgcggg gggcagtggc ggtggcgcgg aggacgcacg ccacgtttc 120  
 gacgaattgc tccgtcgtgg cataccagat gtcttctcct acaatattct tctcaacggg 180  
 ctgtgtgtatc agaacagaag ccaagaagct ctcgagttac tgcacataat ggctgtatc 240  
 ggaggtgact gcccacactga tgggtgtcg tacagcaccc tcataatgg cttttcaag 300  
 gagggggatc tggacaaaat gcttgaccag aggatttcgc caaatgttgt gacctacaac 360  
 tctattatttgc ctgcgtatgc caaggctcaa actgtggaca aggccatgg ggtacttacc 420  
 accatggta agagtgggtt catgcctgtat tgcacatataatgtat tggcatggg 480  
 ttttgcattt cagggcagcc gaaagaggat attgtatttc taaaaagat ggcgcgtgtat 540  
 ggtgtcgaac cagatgttgt tacttataac tcgctcatgg attatcttgc caagaacgg 600  
 agatgcacgg aagcaagaaa gatttttgtat tctatgacca agagggccct aaagcctgtat 660  
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 catggctctt tggatttgc ggtacgaaac ggtatccacc ctaatcatta tggtttcagc 780  
 attcttagtat gtgcatacgc taaacaagag aaagtagaaag aggcaatgct tggattcagc 840  
 aaaatgagggc agcaaggatt gaatccgaat gcagtgacccat tataatgtat 900

ctttgcagg caggttagt agaagatgct atgctttattt ttgaggcagat gatcgatgaa 960  
 ggactaagac ctgacagcat tgtttataac tccttaattt atagtcctcg tatctttgac 1020  
 aatggggaga aggctgaaga gttatttctt gaaatgttgg atcgaggcat ctgtcttagc 1080  
 actatttctt ttaattcaat aattgacagt cattgcaaaag aaggggagggt tatagaatct 1140  
 gaaaaactct ttgacttgc ggtacgaatt ggtgtgaagc ccgatatcat tacccttggc 1200  
 aggttttgg ggagcgcgaag gcgcgactac tcactgttcg tcaacatcta cttcatctc 1260  
 accaacaatgt cgaacactgg agacaaggag aaggagactc ccgtcaacac caacggaggc 1320  
 aatactgctt caaactccag cggaggacca ttcttggca cataacaatcat aatccttcat 1380  
 ggactttgca aaaacaaact cactgatgat gcacttcgaa tgtttcagaa cctatgtttg 1440  
 atggatttga agcttgaggc taggacttc aacattatga ttgatgcatt gcttaaagtt 1500  
 ggcagaaatg atgaagccaa ggattttgtt gtgccttct cgtctaacgg ttttagtgcgc 1560  
 aattatttggc cgtacagatt gatggctgaa aatattatac gacaggggtt gctagaagaa 1620  
 ttggatcaac tctttcttc aatggaggac aatggctgta ctgttgcactc tggcatgcta 1680  
 aatttcatgt ttagggaact gttgcagaga ggtgagataa ccagggctgg cacttacatt 1740  
 tccatgatttgc atgagaagca ctttccttc gaagcatcca ctgcttcctt gtttataat 1800  
 ctttgtctg gggaaaata tcaagaatcatatatttc tccctgaaaa atacaagtcc 1860  
 ttatagaat ctttgagctg ctga 1884

<210> 35

<211> 627

<212> PRT

<213> Oryza sativa

<400> 35

Met Ala Arg Arg Ala Ala Ser Arg Ala Ala Gly Ala Leu Arg Ser Glu  
 1 5 10 15

Gly Ser Ile Gln Gly Arg Gly Arg Ala Gly Gly Ser Gly Gly Gly  
 20 25 30

Ala Glu Asp Ala Arg His Val Phe Asp Glu Leu Leu Arg Arg Gly Ile  
 35 40 45

Pro Asp Val Phe Ser Tyr Asn Ile Leu Leu Asn Gly Leu Cys Asp Glu  
 50 55 60

Asn Arg Ser Gln Glu Ala Leu Glu Leu Leu His Ile Met Ala Asp Asp  
 65 70 75 80

Gly Gly Asp Cys Pro Pro Asp Val Val Ser Tyr Ser Thr Val Ile Asn  
 85 90 95

Gly Phe Phe Lys Glu Gly Asp Leu Asp Lys Met Leu Asp Gln Arg Ile  
 100 105 110

Ser Pro Asn Val Val Thr Tyr Asn Ser Ile Ile Ala Ala Leu Cys Lys  
 115 120 125

Ala Gln Thr Val Asp Lys Ala Met Glu Val Leu Thr Thr Met Val Lys  
 130 135 140

Ser Gly Val Met Pro Asp Cys Met Thr Tyr Asn Ser Ile Val His Gly  
 145 150 155 160

Phe Cys Ser Ser Gly Gln Pro Lys Glu Ala Ile Val Phe Leu Lys Lys  
 165 170 175

Met Arg Ser Asp Gly Val Glu Pro Asp Val Val Thr Tyr Asn Ser Leu  
 180 185 190

Met Asp Tyr Leu Cys Lys Asn Gly Arg Cys Thr Glu Ala Arg Lys Ile  
 195 200 205

Phe Asp Ser Met Thr Lys Arg Gly Leu Lys Pro Asp Ile Thr Thr Tyr  
 210 215 220

Gly Thr Leu Leu Gln Gly Tyr Ala Thr Lys Gly Ala Leu Val Glu Met  
 225 230 235 240

His Gly Leu Leu Asp Leu Met Val Arg Asn Gly Ile His Pro Asn His  
 245 250 255

Tyr Val Phe Ser Ile Leu Val Cys Ala Tyr Ala Lys Gln Glu Lys Val  
 260 265 270

Glu Glu Ala Met Leu Val Phe Ser Lys Met Arg Gln Gln Gly Leu Asn  
 275 280 285

Pro Asn Ala Val Thr Tyr Gly Thr Val Ile Asp Val Leu Cys Lys Ser  
 290 295 300

Gly Arg Val Glu Asp Ala Met Leu Tyr Phe Glu Gln Met Ile Asp Glu  
 305 310 315 320

Gly Leu Arg Pro Asp Ser Ile Val Tyr Asn Ser Leu Ile His Ser Leu  
 325 330 335

Cys Ile Phe Asp Lys Trp Glu Lys Ala Glu Glu Leu Phe Leu Glu Met  
 340 345 350

Leu Asp Arg Gly Ile Cys Leu Ser Thr Ile Phe Phe Asn Ser Ile Ile  
 355 360 365

Asp Ser His Cys Lys Glu Gly Arg Val Ile Glu Ser Gly Lys Leu Phe  
 370 375 380

Asp Leu Met Val Arg Ile Gly Val Lys Pro Asp Ile Ile Thr Leu Gly  
 385 390 395 400

Arg Phe Leu Gly Ser Ala Arg Arg Asp Tyr Ser Leu Phe Val Asn Ile  
 405 410 415

Tyr Phe Ile Phe Thr Asn Met Ser Asn Thr Gly Asp Lys Glu Lys Glu  
 420 425 430

Thr Pro Val Asn Thr Asn Gly Gly Asn Thr Ala Ser Asn Ser Ser Gly  
 435 440 445

Gly Pro Phe Leu Gly Thr Tyr Asn Ile Ile Leu His Gly Leu Cys Lys  
 450 455 460

Asn Lys Leu Thr Asp Asp Ala Leu Arg Met Phe Gln Asn Leu Cys Leu  
 465 470 475 480

Met Asp Leu Lys Leu Glu Ala Arg Thr Phe Asn Ile Met Ile Asp Ala  
 485 490 495

Leu Leu Lys Val Gly Arg Asn Asp Glu Ala Lys Asp Leu Phe Val Ala  
 500 505 510

Phe Ser Ser Asn Gly Leu Val Pro Asn Tyr Trp Thr Tyr Arg Leu Met  
 515 520 525

Ala Glu Asn Ile Ile Gly Gln Gly Leu Leu Glu Glu Leu Asp Gln Leu  
 530 535 540

Phe Leu Ser Met Glu Asp Asn Gly Cys Thr Val Asp Ser Gly Met Leu  
 545 550 555 560

Asn Phe Ile Val Arg Glu Leu Leu Gln Arg Gly Glu Ile Thr Arg Ala  
 565 570 575

Gly Thr Tyr Leu Ser Met Ile Asp Glu Lys His Phe Ser Leu Glu Ala  
 580 585 590

Ser Thr Ala Ser Leu Phe Ile Asp Leu Leu Ser Gly Gly Lys Tyr Gln  
 595 600 605

Glu Tyr His Ile Phe Leu Pro Glu Lys Tyr Lys Ser Phe Ile Glu Ser  
 610 615 620

Leu Ser Cys  
 625

&lt;210&gt; 36

&lt;211&gt; 1554

&lt;212&gt; DNA

&lt;213&gt; Oryza sativa

&lt;400&gt; 36

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 gctccggtgg ctgggagggtg gcgacgtcgg cgccgaaatg tggccggag cgccgcgcgt 120  
 gagagccccg agctgcgacg gcatcacgccc gactaccggc cgtggccggc gcacatggag 180  
 gcaaagccgg tctacttcgc gtcgaggcgt gcctccgggc ggcccggact gcagcagcag 240  
 ctcgtccggc ccacccaaat ctgggcccattt ctggccgatc tcaagcttccc ggagcggaga 300  
 ccgatctggg ccgtccatcc gcccggccca gccaatcgga cggtggtgtt attactgtac 360  
 tgccaggtcg gtgaccctcc gcccggcg gcccggcg cggcggcagg catggcgcgc 420  
 cgtgtcacca cccttacccg cccccgcacc cgcgcggcg cggcggcggtt ccccgccg 480  
 cagggtggta cgacccaaga cctagggcgc gccccggca gtggcaccga gggcgcacgc 540  
 cacgtgctcg acgaattgcc gctacggggc tggggccctt cgatctacag cttcaaccgc 600  
 accctcaccg acgtcgcgcg tgacagccca gcccgcagcag ttgcgtctt caaccgcatt 660  
 gcccggccg gcccgcacga ggtaactccc gacttggca cctacagcat tctcatcggt 720  
 tgctgctgcc gcccggccg cttggacctc gtttgcgg cttggccaa tgtcattaag 780  
 aaggatttta gagtggaagc catcaccttc gtcctctgc tcaaggccctt ctgtgccac 840  
 aagaggacga ggcacgcattt ggacatagtg ctccgcagaa tgaccggactt cagctgcatt 900  
 ccagatgtt tctcctgcac cattttcttc aagggtctgt gtgatggaa cagaagccaa 960  
 gaagctctcg agctgctgca catgatggctt gatgatcgag gaggaggtt cccacactgt 1020  
 gtgggtcgat ataccactgtt catcaatggc ttcttcaaag agggggatttcc agacaaagct 1080  
 tacagtacat accatgaaat gcttgatcggtt aggatttcac caaatgtt gacctacacgc 1140  
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 ggcgtcgaac caaatgtt gtttgcgtt tacttataaa tcactgtatgattatctttt caagaatggc 1380  
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 attgcttacccctt atcgtaaccctt gcttcagggg tatgcttacca aaggagccctt tggttggatgt 1500  
 catgctctctt tggattttgtt ggttgcgtt gtttacaatgtt atttggagaa gtta 1554

&lt;210&gt; 37

&lt;211&gt; 517

&lt;212&gt; PRT

&lt;213&gt; Oryza sativa

&lt;400&gt; 37

Met	Ala	Arg	Arg	Gly	Arg	Arg	Tyr	Cys	Arg	Ala	Glu	Gly	Thr	Glu	Glu
1															

Arg	Gly	Thr	Gly	Ala	Pro	Val	Ala	Gly	Arg	Trp	Arg	Arg	Arg	Pro
20														

Asn Val Phe Pro Ser Ala Ala Leu Glu Ser Pro Glu Leu Arg Arg His  
35 40 45

His Ala Asp Tyr Arg Pro Trp Ala Ala His Met Glu Ala Lys Pro Val  
50 55 60

Tyr Phe Ala Ser Arg Arg Ala Ser Gly Arg Pro Glu Leu Gln Gln Gln  
65 70 75 80

Leu Val Arg Pro Thr Pro Ile Trp Ala Asp Trp Ala Asp Leu Ser Leu  
85 90 95

Pro Glu Arg Arg Pro Ile Trp Ala Val His Pro Arg Arg Pro Ala Asn  
100 105 110

Arg Thr Val Gly Val Leu Leu Tyr Cys Gln Val Gly Asp Pro Pro Pro  
115 120 125

Pro Ala Ala Ala Ala Ala Ala Gly Met Ala Arg Arg Val Thr Thr  
130 135 140

Leu Thr Arg Ala Arg Thr Arg Ala Arg Gly Gly Gly Val Pro Ser Ala  
145 150 155 160

Gln Gly Gly Thr Thr Gln Asp Leu Gly Arg Ala Gly Gly Ser Gly Thr  
165 170 175

Glu Gly Ala Arg His Val Leu Asp Glu Leu Pro Leu Arg Gly Trp Gly  
180 185 190

Ala Ser Ile Tyr Ser Phe Asn Arg Thr Leu Thr Asp Val Ala Arg Asp  
195 200 205

Ser Pro Ala Ala Ala Val Ser Leu Phe Asn Arg Met Ala Arg Ala Gly  
210 215 220

Ala Asp Glu Val Thr Pro Asp Leu Cys Thr Tyr Ser Ile Leu Ile Gly  
225 230 235 240

Cys Cys Cys Arg Ala Gly Arg Leu Asp Leu Gly Phe Ala Ala Leu Gly  
245 250 255

Asn Val Ile Lys Lys Gly Phe Arg Val Glu Ala Ile Thr Phe Ala Pro  
260 265 270

Leu Leu Lys Gly Leu Cys Ala Asp Lys Arg Thr Ser Asp Ala Met Asp  
275 280 285

Ile Val Leu Arg Arg Met Thr Glu Leu Ser Cys Met Pro Asp Val Phe  
 290 295 300  
  
 Ser Cys Thr Ile Leu Leu Lys Gly Leu Cys Asp Glu Asn Arg Ser Gln  
 305 310 315 320  
  
 Glu Ala Leu Glu Leu Leu His Met Met Ala Asp Asp Arg Gly Gly  
 325 330 335  
  
 Ser Pro Pro Asp Val Val Ser Tyr Thr Thr Val Ile Asn Gly Phe Phe  
 340 345 350  
  
 Lys Glu Gly Asp Ser Asp Lys Ala Tyr Ser Thr Tyr His Glu Met Leu  
 355 360 365  
  
 Asp Arg Arg Ile Ser Pro Asn Val Val Thr Tyr Ser Ser Ile Ile Ala  
 370 375 380  
  
 Ala Leu Cys Lys Ala Gln Ala Met Asp Lys Ala Met Glu Val Leu Asn  
 385 390 395 400  
  
 Thr Met Val Lys Asn Gly Val Met Pro Asp Cys Met Thr Tyr Asn Ser  
 405 410 415  
  
 Ile Leu His Gly Tyr Cys Ser Ser Gly Gln Pro Lys Glu Ala Ile Gly  
 420 425 430  
  
 Thr Leu Lys Lys Met Arg Ser Asp Gly Val Glu Pro Asn Val Val Thr  
 435 440 445  
  
 Tyr Arg Ser Leu Met Asn Tyr Leu Cys Lys Asn Gly Arg Cys Thr Glu  
 450 455 460  
  
 Ala Arg Lys Ile Phe Asp Ser Met Thr Lys Arg Gly Leu Glu Pro Asp  
 465 470 475 480  
  
 Ile Ala Thr Tyr Arg Thr Leu Leu Gln Gly Tyr Ala Thr Lys Gly Ala  
 485 490 495  
  
 Leu Val Glu Met His Ala Leu Leu Asp Leu Met Asp Pro Glu Phe Tyr  
 500 505 510  
  
 Lys Tyr Leu Glu Lys  
 515

&lt;210&gt; 38

&lt;211&gt; 2784

<212> DNA

<213> Oryza sativa

<400> 38

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tcggccggcgg cccgcggcggc ggaggtgacg gagtcgcagg aggacgcggc ggctgttggc 180  
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tgcaatggcg ggggggctgc cgatgacgag gaggtcgaga ggaaggcccc cgctgtcg 300  
cgatcaagc tctgccatga gcttctgcgg gagaggaggt ggccgcgcgt gcgggcagcc 360  
ttggcgcagc tggtgactga gcaagggtgag catgctatga attttccccca ttctgattat 420  
caactctact catgtggtat ctgaataact atggtgattt gtgtgaggag gcgttaggaat 480  
ggcatcggtt gtttgaact tctgatcgat atgaatgtgt gacacaggat atattgttt 540  
tccagaggca ttatcaattt atcattacca tataaaaaaa acgtaaagaaaa gggtcgaaag 600  
caatgcatac atagttgtat ttggtgtagt attattactg taattcgttt tttactagaa 660  
ggtctctgca agtatgacaa actagtaaca taaaaattgt tcgcgtttaa tcttattgcg 720  
cttcctgctg taggatctgg gtctgcagct gctctctgtg acatctttagt gaacagattc 780  
agagagtgtg attccaacgg ttgtgtatgg gatgctctag cgaacagttt tgctagagct 840  
cagatggttc atgatgccct ttacgttctt agtaaaaatgt gcagcctaaa catgcaaatac 900  
tcgggtttca cctatgacag tttattgcac ggcttaagga tgacagacgt ggcattggag 960  
ctttttgaag aaatggagtc ttgtgggttc tctcccagtg aatattcgca tagtattatt 1020  
attaatggcc tctgttaagca agataagggtt ggagaagctt tatctttcct tcaggaagct 1080  
aggaaggagg gaaagtttaa acccttggga atgaccctta acattctttagt gtctgcattt 1140  
tgtaatttggg ggtttgttca gtctgcacaaa tcattttat gcctgatgtt gaaatatgg 1200  
tttagtccctg acaggtatac cttttctacc cttatacacgt gtctatgtaa agtaggttca 1260  
atggaggaag cattggatct ttgcagagaga gtgacacaaag aaggaatggaa acttgagatt 1320  
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Ala Lys Ser Phe Leu Cys Leu Met Leu Lys Tyr Gly Leu Val Pro Asp		
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Arg Tyr Thr Phe Ser Thr Leu Ile His Gly Leu Cys Lys Val Gly Ser		
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Gln Met Ala Phe His Ile His Asp Ile Met Leu Cys Arg Gly Leu Val		
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Pro Thr Pro Val Thr Tyr Asn Leu Leu Ile Asn Val Leu Cys Leu Lys		
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Gly Ile Lys Leu Arg Lys Phe Ala Tyr Thr Thr Leu Ile Lys Ala Gln		

705

710

715

720

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Asn Arg Leu Cys Lys Arg Gln Phe Ala Lys Glu Ala Phe Met Phe Val  
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Pro Ile Met Leu Ser Val Gly Ile Tyr Pro Asp Thr Gln Ile Tyr Cys  
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Phe Ser Arg Phe Phe Ser Ala Ile Ala Arg Thr Lys Gln Phe Asn Leu  
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Val Leu Asp Phe Cys Lys Gln Leu Glu Leu Asn Gly Ile Ala His Asn  
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Thr Cys Phe Ala Tyr Ser Val Leu Gly Lys Val Met Lys Leu Gly Tyr  
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 tatacccttata aatataataa tctgggggg tagatgttagt aaatatttttta atgggttaggc 5520  
 cagataatat ttgaaatgtt tggatgttgc ctgttattat tttagatttgc ttgggttattt 5580  
 ttgttaagtgtt ctaatcaaca aatgcacgtc atttgcgtt aatacactac tttacttgc 5640  
 ccataattaa ttaatagaca ttctcttgc attacatcac attaccatag ttaatttgc 5700  
 tggtagtta tatataatccg gtgttagttaa atttttcatt ataaattatg gcaagacgag 5760  
 taaatatgaa acttacatgc agaggcagat aaatatttttgc ttttgcgtt gtttttgc 5820  
 aaaacaatgtt taaattatc gcgcaaaacc ctttcgtttt gttttatcg tggatgttcc 5880  
 tggatgttgc tttttttttt cattccatataa tggaaatgtt cttcattgtt ccacatttgc ttcttataat 5940  
 gtggctatttgc tggatgttgc cataatgttgc ccaacttgc acattgttaaa aacaatgtt 6000  
 cttttgc ttttttttttgc ttttttttttgc ttttttttttgc gttcacccgc 6060  
 gccaactccg gcccctacgtt catcaagtac ttatccatataa ttttttttttgc ttatccatataa 6120

aataacttaca atttgtttaa ttaaatcata gaatttagctg atacacacat atatagtcaa 6180  
 aatgagata gtaactgaag cagctcaagt tcaattttta ggtgcaaaat tcttctatca 6240  
 gttattatgt tttgcttca aattaataac atattcatat agccgacctc aactaattac 6300  
 gcattgatgc atagttcatt gtactaggaa aagtaaaatt tcattttaa gttagttat 6360  
 ttgagcaagt tatatatata tacacaatgc atgtgcttat atcccttcc aatgctaact 6420  
 ctgacttcat gaaaattaaa ttataggtgt tacttagtg agggacgcga attaatatta 6480  
 catcaactggt agtggcggag ccagtattt tactaaggag tatcaaata taaataagta 6540  
 aatatacgaa atattaaaag gatagtgaat ctccctttt aatgtacttc atttaaagt 6600  
 tagtttattt gagaaaagtta tatatataca atgtatgaac tgatattctt tgataatgat 6660  
 gatgcctatg tggatagtga atctccctt ttaatgtga tgaaaaataa a 6711

<210> 43  
 <211> 591  
 <212> PRT  
 <213> Petunia sp.

<400> 43  
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 Ser Phe Phe Ala Tyr Ser Ile Ala Pro Arg His Tyr Ser Thr Asn Thr  
 20 25 30  
 Cys Ser Ile Ser Val Lys Gly Asn Phe Gly Val Ser Asn Glu Phe Gln  
 35 40 45  
 Asn Val Lys Cys Leu Asp Asp Ala Phe Ser Leu Phe Arg Gln Met Val  
 50 55 60  
 Arg Thr Lys Pro Leu Pro Ser Val Ala Ser Phe Ser Lys Leu Leu Lys  
 65 70 75 80  
 Ala Met Val His Met Lys His Tyr Ser Ser Val Val Ser Leu Phe Arg  
 85 90 95  
 Glu Ile His Lys Leu Arg Ile Pro Val His Glu Phe Ile Leu Ser Ile  
 100 105 110  
 Val Val Asn Ser Cys Cys Leu Met His Arg Thr Asp Leu Gly Phe Ser  
 115 120 125  
 Val Leu Ala Ile His Phe Lys Lys Gly Ile Pro Tyr Asn Glu Val Thr  
 130 135 140  
 Phe Thr Thr Leu Ile Arg Gly Leu Phe Ala Glu Asn Lys Val Lys Asp  
 145 150 155 160

Ala Val His Leu Phe Lys Lys Leu Val Arg Glu Asn Ile Cys Glu Pro  
165 170 175

Asn Glu Val Met Tyr Gly Thr Val Met Asn Gly Leu Cys Lys Lys Gly  
180 185 190

His Thr Gln Lys Ala Phe Asp Leu Leu Arg Leu Met Glu Gln Gly Ser  
195 200 205

Thr Lys Pro Asn Thr Arg Thr Tyr Thr Ile Val Ile Asp Ala Phe Cys  
210 215 220

Lys Asp Gly Met Leu Asp Gly Ala Thr Ser Leu Leu Asn Glu Met Lys  
225 230 235 240

Gln Lys Ser Ile Pro Pro Asp Ile Phe Thr Tyr Ser Thr Leu Ile Asp  
245 250 255

Ala Leu Cys Lys Leu Ser Gln Trp Glu Asn Val Arg Thr Leu Phe Leu  
260 265 270

Glu Met Ile His Leu Asn Ile Tyr Pro Asn Val Cys Thr Phe Asn Ser  
275 280 285

Val Ile Asp Gly Leu Cys Lys Glu Gly Lys Val Glu Asp Ala Glu Glu  
290 295 300

Ile Met Arg Tyr Met Ile Glu Lys Gly Val Asp Pro Asp Val Ile Thr  
305 310 315 320

Tyr Asn Met Ile Ile Asp Gly Tyr Gly Leu Arg Gly Gln Val Asp Arg  
325 330 335

Ala Arg Glu Ile Phe Asp Ser Met Ile Asn Lys Ser Ile Glu Pro Asp  
340 345 350

Ile Ile Ser Tyr Asn Ile Leu Ile Asn Gly Tyr Ala Arg Gln Lys Lys  
355 360 365

Ile Asp Glu Ala Met Gln Val Cys Arg Glu Ile Ser Gln Lys Gly Leu  
370 375 380

Lys Pro Ser Ile Val Thr Cys Asn Val Leu Leu His Gly Leu Phe Glu  
385 390 395 400

Leu Gly Arg Thr Lys Ser Ala Gln Asn Phe Phe Asp Glu Met Leu Ser  
405 410 415

Ala Gly His Ile Pro Asp Leu Tyr Thr His Cys Thr Leu Leu Gly Gly  
 420 425 430

Tyr Phe Lys Asn Gly Leu Val Glu Glu Ala Met Ser His Phe His Lys  
                  435                 440                 445

Leu Glu Arg Arg Arg Glu Asp Thr Asn Ile Gln Ile Tyr Thr Ala Val  
 450 455 460

Ile Asp Gly Leu Cys Lys Asn Gly Lys Leu Asp Lys Ala His Ala Thr  
 465 470 475 480

Phe Glu Lys Leu Pro Leu Ile Gly Leu His Pro Asp Val Ile Thr Tyr  
 485 490 495

Thr Ala Met Ile Ser Gly Tyr Cys Gln Glu Gly Leu Leu Asp Glu Ala  
500 505 510

Lys Asp Met Leu Arg Lys Met Glu Asp Asn Gly Cys Leu Ala Asp Asn  
 515 520 525

Ser Glu Met Lys Ala Phe Leu Glu Glu Ile Ala Gly Lys Ser Phe Ser  
 545 550 555 560

Phe Glu Ala Ala Thr Val Glu Leu Leu Met Asp Ile Ile Ala Glu Asp  
 565 570 575

<210> 44  
<211> 1779  
<212> DNA  
<213> Petunia sp

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 tatggAACGG tcattGAATGG gctttGCAAa aaggGCCATA ctcaAAAAGC ttttgatttG 600  
 ctccggTTAA tggAACAAAGC aagtACTAAG cccaatacat gtatctatAG cattGTTATC 660  
 gatgcCTTTT gcaaAGATGG gatgCTAGAT ggtgCTACCA gcctttGAA tgAGATGAAA 720  
 caaaaaAGCA ttccCTCCGA catTTTact tatAGCactT taattGatGc tttgtGtaAG 780  
 ttaAGTCAGT gggAAAATGT taggACTTG ttccTTGAGA tgatacatCT taatATTTAT 840  
 ccaaATGTGT gcacCttCAA ctccGTCATT gatggACTAT gcaaAGAGGG gaaAGTAGAA 900  
 gacgCTGAGG aaATAATGAG atACATGATT gaaaaAGGTG tagaccCTGA tGtgatCACC 960  
 tataATATGA taattGACGG atATGGCTTG cgtggTCAG gggATAGAGC acgggAAATT 1020  
 tttgattCCA tGATCAATAA gAGCATTGAG cccaATATTa ttAGCTATAA tataCTAATA 1080  
 aatGGATATG ccaggcAAAA gaaaATAGAC gaggCAATGC aagtCTGCCG tGAAATTTC 1140  
 caaaaAGGGAT tGAAACCTAG tattGTTACC tGCAATGTTc tcttGcatGG tcttttGAA 1200  
 cttGGAAGAA ctaAAATCTGC acaAAATTc tttgatGAGA tGCTATCTGC ggggcACATA 1260  
 cctgatttt acactCATTG tactTTGCTT ggtggTTatt ttaAGAATGG acttGTTGAA 1320  
 gaggCTATGT cacactTCCA taAGTTGAA agaAGGAGAG aAGATAcAAA tattCAAATT 1380  
 tacacGGCTG tcattGATGG attGTCAAA aatGGTAAGC tcgacaAAAGC tcatGCTACG 1440  
 tttgAGAGC ttccCTTGTat aggCTTACAT CCTGATGTGA taACATACAC tGCAATGATT 1500  
 agtggatatt gtcaAGAAGG gttGTTAGAT gaAGCTAAAG atATGCTAAG gaaaATGGAG 1560  
 gacaATGGTT gtttGGCAGA caACCgAAcA tacaATGTTA ttGtGCGGGG atttCTCAGA 1620  
 agcaATAAAAG ttAGTGAAT gaAGGCTTT ctGAGGAAA tagCTGGAA gagCTTCTCA 1680  
 tttgAGGcAG ctactGTAGA gttattGATG gatATTATAG cAGAGGATCC ttctttGCTT 1740  
 aacatGATTc cagaATTcA ccGGGATAAT aagaAGTGA 1779

<210> 45  
 <211> 592  
 <212> PRT  
 <213> Petunia sp.

<400> 45  
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Ser Phe Phe Ala Tyr Ser Ile Ala Pro Arg His Tyr Ser Thr Asn Thr  
 20 25 30

Arg Ser Ile Ser Val Lys Gly Asn Phe Gly Val Ser Asn Glu Phe Glu  
 35 40 45

Asn Val Lys Cys Leu Asp Asp Ala Phe Ser Leu Phe Arg Gln Met Val  
 50 55 60

Arg Thr Lys Pro Leu Pro Ser Val Val Ser Phe Ser Lys Leu Leu Lys  
 65 70 75 80

Ala Leu Val His Met Lys His Tyr Ser Ser Val Val Ser Leu Phe Arg  
 85 90 95

Glu Ile His Lys Leu Arg Ile Pro Val His Glu Phe Ile Leu Ser Ile  
100 105 110

Val Val Asn Ser Cys Cys Leu Met His Arg Thr Asp Leu Gly Phe Ser  
115 120 125

Val Leu Ala Ile His Phe Lys Lys Gly Ile Pro Phe Asn Gln Val Ile  
130 135 140

Phe Asn Thr Leu Leu Arg Gly Leu Phe Ala Glu Asn Lys Val Lys Asp  
145 150 155 160

Ala Val His Leu Phe Lys Lys Leu Val Arg Glu Asn Ile Cys Glu Pro  
165 170 175

Asn Glu Val Met Tyr Gly Thr Val Met Asn Gly Leu Cys Lys Lys Gly  
180 185 190

His Thr Gln Lys Ala Phe Asp Leu Leu Arg Leu Met Glu Gln Gly Ser  
195 200 205

Thr Lys Pro Asn Thr Cys Ile Tyr Ser Ile Val Ile Asp Ala Phe Cys  
210 215 220

Lys Asp Gly Met Leu Asp Gly Ala Thr Ser Leu Leu Asn Glu Met Lys  
225 230 235 240

Gln Lys Ser Ile Pro Pro Asp Ile Phe Thr Tyr Ser Thr Leu Ile Asp  
245 250 255

Ala Leu Cys Lys Leu Ser Gln Trp Glu Asn Val Arg Thr Leu Phe Leu  
260 265 270

Glu Met Ile His Leu Asn Ile Tyr Pro Asn Val Cys Thr Phe Asn Ser  
275 280 285

Val Ile Asp Gly Leu Cys Lys Glu Gly Lys Val Glu Asp Ala Glu Glu  
290 295 300

Ile Met Arg Tyr Met Ile Glu Lys Gly Val Asp Pro Asp Val Ile Thr  
305 310 315 320

Tyr Asn Met Ile Ile Asp Gly Tyr Gly Leu Arg Gly Gln Val Asp Arg  
325 330 335

Ala Arg Glu Ile Phe Asp Ser Met Ile Asn Lys Ser Ile Glu Pro Asn  
340 345 350

Ile Ile Ser Tyr Asn Ile Leu Ile Asn Gly Tyr Ala Arg Gln Lys Lys  
355 360 365

Ile Asp Glu Ala Met Gln Val Cys Arg Glu Ile Ser Gln Lys Gly Leu  
370 375 380

Lys Pro Ser Ile Val Thr Cys Asn Val Leu Leu His Gly Leu Phe Glu  
385 390 395 400

Leu Gly Arg Thr Lys Ser Ala Gln Asn Phe Phe Asp Glu Met Leu Ser  
405 410 415

Ala Gly His Ile Pro Asp Leu Tyr Thr His Cys Thr Leu Leu Gly Gly  
420 425 430

Tyr Phe Lys Asn Gly Leu Val Glu Glu Ala Met Ser His Phe His Lys  
435 440 445

Leu Glu Arg Arg Arg Glu Asp Thr Asn Ile Gln Ile Tyr Thr Ala Val  
450 455 460

Ile Asp Gly Leu Cys Lys Asn Gly Lys Leu Asp Lys Ala His Ala Thr  
465 470 475 480

Phe Glu Lys Leu Pro Leu Ile Gly Leu His Pro Asp Val Ile Thr Tyr  
485 490 495

Thr Ala Met Ile Ser Gly Tyr Cys Gln Glu Gly Leu Leu Asp Glu Ala  
500 505 510

Lys Asp Met Leu Arg Lys Met Glu Asp Asn Gly Cys Leu Ala Asp Asn  
515 520 525

Arg Thr Tyr Asn Val Ile Val Arg Gly Phe Leu Arg Ser Asn Lys Val  
530 535 540

Ser Glu Met Lys Ala Phe Leu Glu Glu Ile Ala Gly Lys Ser Phe Ser  
545 550 555 560

Phe Glu Ala Ala Thr Val Glu Leu Leu Met Asp Ile Ile Ala Glu Asp  
565 570 575

Pro Ser Leu Leu Asn Met Ile Pro Glu Phe His Arg Asp Asn Lys Lys  
580 585 590

<210> 46  
<211> 25  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 46  
tgcacagtgt tatatttaca taccc 25

<210> 47  
<211> 25  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 47  
tttatgatac atggatttca acgac 25

<210> 48  
<211> 25  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 48  
tgaaaatgac aatcgtaaca gaaaa 25

<210> 49  
<211> 24  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 49  
aacattcctc cagacattat taca 24

<210> 50  
<211> 24  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 50  
gacgctgagg aaataatgag atac

24

<210> 51  
<211> 27  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 51  
tctagaaaaa atgaaggggg aatcaat

27

<210> 52  
<211> 31  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 52  
gaattcactt tgctctcactg ataaactaag a

31